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I, KAY WARD, TEAM LEADER EXAMINATION SUPPORT AND SALES hereby certify that annexed is a true copy of the Provisional specification in connection with Application No. PQ 0052 for a patent by COMMONWEALTH SCIENTIFIC AND INDUSTRIAL RESEARCH ORGANISATION, GOODMAN FIELDER LIMITED and GROUPE LIMAGRAIN PACIFIC PTY. LTD. filed on 29 April 1999.



WITNESS my hand this Eleventh day of May 2000

Kalard

KAY WARD

TEAM LEADER EXAMINATION SUPPORT AND SALES

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# A U S T R A L I A Patents Act 1990

# PROVISIONAL SPECIFICATION

for the invention entitled:

# "NOVEL GENES ENCODING WHEAT STARCH SYNTHASES AND USES THEREFOR"

The invention is described in the following statement:



# NOVEL GENES ENCODING WHEAT STARCH SYNTHASES AND USES THEREFOR

### FIELD OF THE INVENTION

5 The present invention relates generally to isolated nucleic acid molecules encoding wheat starch synthase enzymes and more particularly, to isolated nucleic acid molecules that encode wheat SSII and SSIII enzyme activities. The isolated nucleic acid molecules provide the means for modifying starch content and composition in plants, for example the ratio of amylose:amylopectin in the starch granule of the endosperm during the grain-filling phase of endosperm development. The isolated nucleic acid molecules of the present invention also provide the means for screening plant lines to determine the presence of natural and/or induced mutations in starch synthase genes which affect starch content and/or composition. The isolated nucleic acid molecules of the present invention further provide for the screening-assisted breeding of plants having desirable starch content and/or composition, in addition to providing for the direct genetic manipulation of plant starch content and/or composition.

#### **GENERAL**

Bibliographic details of the publications numerically referred to in this specification are collected at the end of the description.

This specification contains nucleotide and amino acid sequence information (SEQ ID Nos:) prepared using the programme PatentIn Version 2.0, presented herein at the end of the specification. Each nucleotide or amino acid sequence is identified in the sequence listing by the numeric indicator <210> followed by the sequence identifier (e.g. <210>1, <210>2, etc). The length, type of sequence (DNA, protein (PRT), etc) and source organism for each nucleotide or amino acid sequence are indicated by information provided in the numeric indicator fields <211>, <212> and <213>, respectively. Nucleotide and amino acid sequences (SEQ ID NOs:) referred to in the specification are defined by the information provided in numeric indicator field <400>

followed by the sequence identifier (eg. <400>1, <400>2, etc).

The designation of nucleotide residues referred to herein are those recommended by the IUPAC-IUB Biochemical Nomenclature Commission, wherein A represents 5 Adenine, C represents Cytosine, G represents Guanine, T represents thymine, Y represents a pyrimidine residue, R represents a purine residue, M represents Adenine or Cytosine, K represents Guanine or Thymine, S represents Guanine or Cytosine, W represents Adenine or Thymine, H represents a nucleotide other than Guanine, B represents a nucleotide other than Adenine, V represents a nucleotide other than 10 Thymine, D represents a nucleotide other than Cytosine and N represents any nucleotide residue.

The designations for naturally-occurring amino acid residues referred to herein are set forth in Table I. The designations for a non-limiting set of non-naturally-occurring amino acids is listed in Table 2.

As used herein the term "derived from" shall be taken to indicate that a specified integer may be obtained from a particular source albeit not necessarily directly from that source.

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Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated step or element or integer or group of steps or elements or integers but not the exclusion of any other step or element or integer or group of steps or elements or integers.

TABLE 1

Amino Acid	Three-letter Code	One-letter Code
Alanine	Ala	A
Arginine	Arg	R
Asparagine	Asn	N
Aspartic acid	Asp	D
Cysteine	Cys	C
) Glutamine	Gln	Q
Glutamic acid	Glu	E
Glycine	Gly	G
Histidine	His	H
Isoleucine	Ile	I
5 Leucine	Leu	L
Lysine	Lys	K
Methionine	Met	M
Phenylalanine	Phe	F
Proline	Pro	P
0 Serine	Ser	S
Threonine	Thr	T
Tryptophan	Trp	W
Tyrosine	Tyr	Y
Valine	Val	V
5 Aspartate/glutamate	Baa	В
Asparagine/glutamine		
Any amino acid as above	. Xaa	X

TABLE 2

_	Non-conventional amino acid	Code	Non-conventional amino acid	Code
5	α-aminobutyric acid	Abu	L-N-methylalanine	Nmala
	α-amino-α-methylbutyrate	Mgabu	L-N-methylarginine	Nmarg
	aminocyclopropane-	Cpro	L-N-methylasparagine	Nmasn
	carboxylate		L-N-methylaspartic acid	Nmasp
0	aminoisobutyric acid	Aib	L-N-methylcysteine	Nmcys
	aminonorbornyl-	Norb	L-N-methylglutamine	Nmgln
	carboxylate		L-N-methylglutamic acid	Nmglu
	cyclohexylalanine	Chexa	L-N-methylhistidine	Nmhis
	cyclopentylalanine	Cpen	L-N-methylisolleucine	Nmile
5	D-alanine	Dal	L-N-methylleucine	Nmleu
	D-arginine	Darg	L-N-methyllysine	Nmlys
	D-aspartic acid	Dasp	L-N-methylmethionine	Nmmet
	D-cysteine	Dcys	L-N-methylnorleucine	Nmnle
	D-glutamine	Dgln	L-N-methylnorvaline	Nmnva
0.	D-glutamic acid	Dglu	L-N-methylornithine	Nmorn
	D-histidine	Dhis	L-N-methylphenylalanine	Nmphe
	D-isoleucine	Dile	L-N-methylproline	Nmpro
	D-leucine	Dleu	L-N-methylserine	Nmser
	D-lysine	Dlys	L-N-methylthreonine	Nmthr
:5	D-methionine	Dmet	L-N-methyltryptophan	Nmtrp
	D-ornithine	Dorn	L-N-methyltyrosine	Nmtyr
	D-phenylalanine	Dphe	L-N-methylvaline	Nmval
	D-proline	Dpro	L-N-methylethylglycine	Nmetg
	D-serine	Dser	L-N-methyl-t-butylglycine	Nmtbug
0	D-threonine	Dthr	L-norleucine	Nle

	D-tryptophan	Dtrp	L-norvaline	Nva
	D-tyrosine	Dtyr	α-methyl-aminoisobutyrate	Maib
	D-valine	Dval	$\alpha$ -methyl- $\gamma$ -aminobutyrate	Mgabu
	D-α-methylalanine	Dmala	α-methylcyclohexylalanine	Mchexa
5	D-α-methylarginine	Dmarg	$\alpha$ -methylcylcopentylalanine	Mcpen
	D-α-methylasparagine	Dmasn	$\alpha$ -methyl- $\alpha$ -napthylalanine	Manap
	D-α-methylaspartate	Dmasp	$\alpha$ -methylpenicillamine	Mpen
	D-α-methylcysteine	Dmcys	N-(4-aminobutyl)glycine	Nglu
	D-α-methylglutamine	Dmgln	N-(2-aminoethyl)glycine	Naeg
10	D-α-methylhistidine	Dmhis	N-(3-aminopropyl)glycine	Norn
	D-α-methylisoleucine	Dmile	$N$ -amino- $\alpha$ -methylbutyrate	Nmaabu
	D-α-methylleucine	Dmleu	$\alpha$ -napthylalanine	Anap
	D-α-methyllysine	Dmlys	N-benzylglycine	Nphe
	D-α-methylmethionine	Dmmet	N-(2-carbamylethyl)glycine	Ngln
15	D-α-methylornithine	Dmorn	N-(carbamylmethyl)glycine	Nasn
	D-α-methylphenylalanine	Dmphe	N-(2-carboxyethyl)glycine	Nglu
	D-α-methylproline	Dmpro	N-(carboxymethyl)glycine	Nasp
	D-α-methylserine	Dmser	N-cyclobutylglycine	Ncbut
	D-α-methylthreonine	Dmthr	N-cycloheptylglycine	Nchep
20	D-α-methyltryptophan	Dmtrp	N-cyclohexylglycine	Nchex
	D-α-methyltyrosine	Dmty	N-cyclodecylglycine	Ncdec
	D-α-methylvaline	Dmval	N-cylcododecylglycine	Ncdod
	D-N-methylalanine	Dnmala	N-cyclooctylglycine	Ncoct
	D-N-methylarginine	Dnmarg	N-cyclopropylglycine	Ncpro
25	D-N-methylasparagine	Dnmasn	N-cycloundecylglycine	Ncund
	D-N-methylaspartate	Dnmasp	N-(2,2-diphenylethyl)	
			glycine	Nbhm
	D-N-methylcysteine	Dnmcys	N-(3,3-diphenylpropyl)	
			glycine	Nbhe

D-N-methylglutamine	Dnmgln	N-(3-guanidinopropyl)	
		glycine	Narg
D-N-methylglutamate	Dnmglu	N-(1-hydroxyethyl)glycine	Nthr
D-N-methylhistidine	Dnmhis	N-(hydroxyethyl))glycine	Nser
5 D-N-methylisoleucine	Dnmile	N-(imidazolylethyl))	
		glycine	Nhis
D-N-methylleucine	Dnmleu	N-(3-indolylyethyl)	
		glycine	Nhtrp
D-N-methyllysine	Dnmlys	N-methyl-γ-aminobutyrate	Nmgabu
10 N-methylcyclohexylalanine	Nmchexa	D-N-methylmethionine	Dnmmet
D-N-methylornithine	Dnmorn	N-methylcyclopentylalanine	Nmcpen
N-methylglycine	Nala	D-N-methylphenylalanine	Dnmphe
N-methylaminoisobutyrate	Nmaib	D-N-methylproline	Dnmpro
N-(1-methylpropyl)glycine	Nile	D-N-methylserine	Dnmser
15 N-(2-methylpropyl)glycine	Nleu	D-N-methylthreonine	Dnmthr
D-N-methyltryptophan	Dnmtrp	N-(1-methylethyl)glycine	Nval
D-N-methyltyrosine	Dnmtyr	N-methyla-napthylalanine	Nmanap
D-N-methylvaline	Dnmval	N-methylpenicillamine	Nmpen
γ-aminobutyric acid	Gabu	N-(p-hydroxyphenyl)glycine	Nhtyr
20 L-t-butylglycine	Tbug	N-(thiomethyl)glycine	Ncys
L-ethylglycine	Etg	penicillamine	Pen
L-homophenylalanine	Hphe	L-α-methylalanine	Mala
$L$ - $\alpha$ -methylarginine	Marg	L-α-methylasparagine	Masn
L-α-methylaspartate	Masp	L-α-methyl-t-butylglycine	Mtbug
25 L-α-methylcysteine	Mcys	L-methylethylglycine	Metg
L-α-methylglutamine	Mgln	L-α-methylglutamate	Mglu
L-α-methylhistidine	Mhis	L-α-methylhomo	
		phenylalanine	Mhphe
L-α-methylisoleucine	Mile	N-(2-methylthioethyl)	
30		glycine	Nmet

	L-α-methylleucine	Mleu	L-α-methyllysine	Mlys
	L-α-methylmethionine	Mmet	L-α-methylnorleucine	Mnle
	L-α-methylnorvaline	Mnva	L-α-methylornithine	Morn
	L-α-methylphenylalanine	Mphe	L-α-methylproline	Mpro
5	L-α-methylserine	Mser	L-α-methylthreonine	Mthr
	L-α-methyltryptophan	Mtrp	L-α-methyltyrosine	Mtyr
	L-α-methylvaline	Mval	L-N-methylhomo	
			phenylalanine	Nmhphe
	N-(N-(2,2-diphenylethyl)		N-(N-(3,3-diphenylpropyl)	
10	carbamylmethyl)glycine	Nnbhm	carbamylmethyl)glycine	Nnbhe
	1-carboxy-1-(2,2-diphenyl-			
	ethylamino)cyclopropane	Nmbc		

15 Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations or any two or more of said steps or features.

The present invention is not to be limited in scope by the specific embodiments described herein, which are intended for the purposes of exemplification only. Functionally-equivalent products, compositions and methods are clearly within the scope of the invention, as described herein.

#### **BACKGROUND TO THE INVENTION**

The biosynthesis of the starch granule is a complex process which involves the action of an array of isoforms of enzymes involved in the starch biosynthesis. Following the formation of glucose-1-phosphate, the enzyme activities required for the synthesis of granular starch include ADP glucose pyrophosphorylase (EC 2.7.7.27), starch synthases (EC 2.4.1.21), branching enzymes (EC 2.4.1.18) and debranching enzymes (EC 3.2.1.41 and EC 3.2.1.68) (Mouille *et al.*, 1996). Plants contain isozymes of each of these activities, and the definition of these isoforms and their roles has been conducted through investigation of the properties of the suite of soluble enzymes found in the stroma of the plastid, analysis of the proteins entrapped within the matrix of the starch granule, and mutational studies to identify genes and define linkages between individual genes and their specific roles.

Starch synthases extend regions of α-1,4 glucan through the transfer of the glucosyl moiety of ADPglucose to the non-reducing end of a pre-existing α-1,4 glucan. In addition to GBSS, 3 other classes of starch synthase have been identified in plants, SSI (wheat, Li *et al.*, 1999 and GenBank Accession No. U48227; rice, Baba *et al.*, 1993; potato, Genbank Accession No. STSTASYNT), SSII (pea, Dry *et al.* 1992; potato, Edwards *et al.*, 1995; maize, Harn *et al.* 1998 and GenBank Accession No. U66377) and SSIII (potato, Abel *et al.*, 1996; maize, Gao *et al.*, 1998). In the cereals, the most comprehensively studied species is maize, where in addition to GBSS, cDNAs encoding SSI, SSIIa, and SSIIb have been isolated, and both cDNA and genomic clones for *dull*1 have been characterised (Knight *et al.*, 1998; Harn *et al.*, 1998; Gao *et al.*, 1998). In maize, the product of the *du1* gene is known as maize SSII, however this gene is the homologue of potato SSIII.

The proteins within the matrix of the wheat starch granule have been extensively studied (Denyer et al., 1995; Rahman et al., 1995; Takaoka et al., 1997; Yamamori and Endo, 1996) and 60, 75, 85, 100, 104 and 105 kDa protein bands can be visualised following SDS-PAGE. The predominant 60 kDa protein is exclusively

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granule-bound and is analogous to the "waxy" granule bound starch synthase (GBSS) gene in maize (Rahman *et al.*, 1995). The combination of three null alleles for this enzyme from each of the wheat genomes (Nakamura *et al.*, 1995) results in the amylose-free "waxy" phenotype found in other species The 75 kDa starch synthase I (wSSI) is found in both the granule and the soluble fraction of wheat endosperm (Denyer *et al.*, 1995; Li *et al.*, 1999) and has been assigned to chromosomes 7A, 7B and 7D (Yamamori and Endo, 1996; Li *et al.*, 1999). The 85 kDa band contains a class II branching enzyme and an unidentified polypeptide (Rahman *et al.*, 1995). The 100, 104 and 105 kDa proteins of the wheat starch granule (designated Sgp-B1, Sgp-D1 and Sgp-A1 by Yamamori and Endo, 1996) have been shown to be encoded by a homeologous set of genes on the short arm of chromosome 7B, 7A and 7D respectively (Yamamori and Endo, 1996; Takaoka *et al.*, 1997). Denyer *et al.* (1995) concluded on the basis of enzyme activity assays that these proteins were also starch synthases. These genes are referred to hereinafter as the "wheat SSII genes".

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While GBSS has been established to be essential for amylose synthesis, the remaining starch synthases are thought to be primarily responsible for the elongation of amylopectin chains, although this does not preclude them from also having non-essential roles in amylose biosynthesis. Differences in kinetic properties between isoforms, and the analysis of mutants lacking various isoforms, suggests that each isoenzyme contributes to the extension of specific subsets of the available non-reducing ends. Accordingly, the production of plants that produce improved starches that are modified for particular purposes, for example starches having high or low amylose:amylopectin ratios, requires the availability of genes encoding the various starch synthase isoforms. Moreover, because of species-specific codon usages and variations in the kinetic parameters of these isoforms in different species, the production of modified starches may require the use of genes derived from particular species.

30 In work leading up to the present invention, the inventors sought to modify wheat

starch composition and content, by providing isolated nucleotide sequences encoding the wheat SSII (i.e. wSSII) and wheat SSIII (i.e. wSSIII) isoenzymes, and by introducing these nucleotide sequences into plants using recombinant DNA technology.

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### SUMMARY OF THE INVENTION

The present invention provides isolated nucleic acid molecules encoding the 100, 104 and 105 kDa SSII (Sgp-1) polypeptides of the wheat starch granule matrix, as determined using the SDS/PAGE system of Rahman et al. (1995), which polypeptides 10 are equivalent to the 100, 108 and 115 kDa polypeptides described by Yamamori and Endo (1996). The present invention further provides isolated nucleic acid molecules encoding the soluble dull1-type wheat starch synthase III polypeptide. Analysis of the polypeptides encoded by these nucleic acid molecules reveals several consensus amino acid sequence motifs (i.e., sequences having at least 25% sequence identity 15 to any one or more of the amino acid sequences selected from the group consisting (a)KVGGLGDVVTS;(b)GHTVEVILPKY;(c) of HDWSSAPVAWLYKEHY; (d) GILNGIDPDIWDPYTD: (e) DVPIVGIITRLTAQKG; (f)NGQVVLLGSA; (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and (h)TGGLVDTV ) that are highly conserved in wheat starch synthase isoenzymes, in addition to isoenzyme-specific 20 sequences, which sequences possess utility in isolating related starch synthaseencoding sequences and in assaying plants for their expression of one or more starch synthase isoenzymes.

Accordingly, one aspect of the present invention provides an isolated nucleic acid molecule which comprises a sequence of nucleotides which encodes, or is complementary to a nucleic acid molecule which encodes a wheat starch synthase polypeptide, protein or enzyme molecule or a functional subunit thereof selected from the following:

(i) a wheat starch synthase II (wSSII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence which is at

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least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, or <400>6;

- (ii) a wheat starch synthase III (wSSIII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>8 or <400>10;and
- (iii) a wheat starch synthase polypeptide, protein or enzyme or functional subunit thereof which comprises a conserved amino acid sequence having at least 25% identity to an amino acid sequence selected from the group consisting of:
  - (a) KVGGLGDVVTS;
  - (b) GHTVEVILPKY;
  - (c) HDWSSAPVAWLYKEHY;
  - (d) GILNGIDPDIWDPYTD;
  - (e) DVPIVGIITRLTAQKG;
  - (f) NGQVVLLGSA;
  - (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and
  - (h)TGGLVDTV

and wherein said wheat starch synthase polypeptide further comprises an amino acid sequence having at least about 85% identity overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, <400>6, <400>8 or <400>10.

In a preferred embodiment, the isolated nucleic acid molecule encodes a starch synthase polypeptide, protein or enzyme having at least about 90% amino acid sequence identity to any one of SEQ ID NOS:<400>2, <400>4, <400>6, <400>8 or <400>10, more preferably having at least about 95% or about 97% or about 99% identity to any one of said amino acid sequences.

30 In an alternative embodiment, the present invention provides an isolated nucleic acid

molecule which encodes a wheat starch synthase polypeptide, protein or enzyme molecule or a functional subunit thereof, wherein said nucleic acid molecule comprises a nucleotide sequence having at least about 85% nucleotide sequence identity to any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>10 to <400>16, or a complementary nucleotide sequence thereto.

In a preferred embodiment, the isolated nucleic acid molecule comprises the nucleotide sequence set forth in any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or is at least about 90% identical, more preferably at least about 95% or 97% or 99% identical to all or a protein-encoding part thereof.

In an alternative embodiment, the present invention provides an isolated nucleic acid molecule which encodes a wheat starch synthase polypeptide, protein or enzyme molecule or a functional subunit thereof, wherein said nucleic acid molecule comprises a nucleotide sequence that is capable of hybridising under at least moderate stringency hybridisation conditions to at least about 30 contiguous nucleotides derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto.

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A second aspect of the present invention provides a method of isolating a nucleic acid molecule that encodes a starch synthase polypeptide, protein or enzyme having at least about 85% amino acid sequence identity to any one SEQ ID NOS:<400>2, <400>4, <400>6, <400>8 or <400>10 and/or which comprises anamino acid sequence having at least 25% identity to an amino acid sequence selected from the group consisting of:

- (a) KVGGLGDVVTS;
- (b) GHTVEVILPKY;
- (c) HDWSSAPVAWLYKEHY;
- 30 (d) GILNGIDPDIWDPYTD;



- (e) DVPIVGIITRLTAQKG;
- (f) NGQVVLLGSA;
- (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and
- (h)TGGLVDTV,

# 5 said method comprising:

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- (i) hybridising a probe or primer comprising at least about 15 contiguous nucleotides in length derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto to single-stranded or double-stranded mRNA, cDNA or genomic DNA; and
- (ii) detecting the hybridised mRNA, cDNA or genomic DNA using a detecting means.

Preferably, the detecting means is a reporter molecule covalently attached to the probe or primer molecule or alternatively, a polymerase chain reaction format. Accordingly, the present invention clearly extends to the use of the nucleic acid molecules provided herein to isolate related starch synthase-encoding sequences using standard hybridisation and/or polymerase chain reaction techniques.

- 20 A third aspect of the invention provides an isolated probe or primer comprising at least about 15 contiguous nucleotides in length derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto.
- 25 Preferably, the probe or primer comprises a nucleotide sequence set forth in any one of SEQ ID NOS:<400>25 to <400>34.

A fourth aspect of the present invention is directed to an isolated or recombinant starch synthase polypeptide, protein or enzyme, preferably substantially free of conspecific or non-specific proteins, which comprises an amino acid sequence selected from the

## following:

- (i) a wheat starch synthase II (wSSII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, or <400>6;
- (ii) a wheat starch synthase III (wSSIII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>8 or <400>10;and
- (iii) a wheat starch synthase polypeptide, protein or enzyme or functional subunit thereof which comprises a conserved amino acid sequence having at least 25% identity to an amino acid sequence selected from the group consisting of:
  - (a) KVGGLGDVVTS;

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- (b) GHTVEVILPKY;
- (c) HDWSSAPVAWLYKEHY;
- (d) GILNGIDPDIWDPYTD;
- (e) DVPIVGIITRLTAQKG;
- (f) NGQVVLLGSA;

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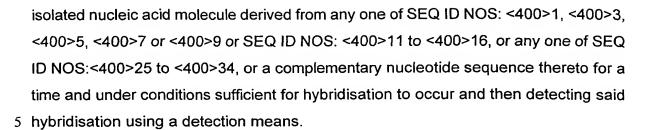
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- (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and
- (h)TGGLVDTV

and wherein said wheat starch synthase polypeptide further comprises an amino acid sequence having at least about 85% identity overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, <400>6, <400>8 or <400>10.

A further aspect of the invention provides a method of assaying for the presence or absence of a starch synthase isoenzyme or the copy number of a gene encoding same in a plant, comprising contacting a biological sample derived from said plant with an

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The detection means according to this aspect of the invention is any nucleic acid based hybridisation or amplification reaction.

10 A further aspect of the present invention utilises the above-mentioned assay method in the breeding and/or selection of plants which express or do not express particular starch sythase isoenzymes or alternatively, which express a particular starch synthase isoenzyme at a particular level in one or more plant tissues. This aspect clearly extends to the selection of transformed plant material which contains one or more of the isolated nucleic acid molecules of the present invention.

A further aspect of the present invention provides a method of modifying the starch content and/or starch composition of one or more tissues or organs of a plant, comprising expressing therein a sense molecule, antisense molecule, ribozyme 20 molecule, co-suppression molecule, or gene-targeting molecule having at least about 85% nucleotide sequence identity to any one of any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto for a time and under conditions sufficient for the enzyme activity of one or more starch synthase isoenzymes to be modified. 25 This aspect of the invention clearly extends to the introduction of the sense molecule, antisense molecule, ribozyme molecule, co-suppression molecule, or gene-targeting molecule to isolated plant cells, tissues or organs or organelles by cell fusion or transgenic means and the regeneration of intact plants therefrom.

30 A further aspect of the present invention provides an isolated promoter that is operable

in the endosperm of a monocotyledonous plant cell, tissue or organ, and preferably in the endosperm of a monocotyledonous plant cell, tissue or organ. For example, the HMG promoter from wheat, or the maize zein gene promoter are particularly preferred, as is the promoter derived from a starch synthase gene of the present invention, such as a promoter that is linked *in vivo* to any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto.

A still further aspect of the present invention contemplates a transgenic plant comprising an introduced sense molecule, antisense molecule, ribozyme molecule, cosuppression molecule, or gene-targeting molecule having at least about 85% nucleotide sequence identity to any one of any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto or a gentic construct comprising same, and to plant propagules, cells, tissues, organs or plant parts derived from said transgenic plant that also carry the introduced molecule(s).

#### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a copy of a photographic representation showing the distribution of wheat endosperm starch synthases between the starch granule and soluble fractions. Lane 1, SDS-PAGE of wheat endosperm starch granule proteins revealed by silver staining; lanes 2-7, immunoblot of wheat endosperm soluble phase and starch granule proteins separated by SDS-PAGE from various developmental stages and probed with an anti-(wheat wSSII peptide) monoclonal antibody. Lanes 2-4 contain proteins from the soluble fraction of wheat endosperm at 15 days post anthesis (Lane 2); 20 days post anthesis (Lane 3); and at 25 days post anthesis (Lane 4). Lanes 5-7 contain proteins from the starch granule of wheat endosperm at 15 days post anthesis (Lane 5); 20 days post anthesis (Lane 6); and at 25 days post anthesis (Lane 7).

30 Figure 2 is a copy of a schematic representation comparing the nucleotide sequences



of cDNA clones designated wSSIIA, wSSIIB and wSSIID, encoding the starch synthase II polypeptides from wheat, using the PILEUP programme of Devereaux *et al.* (1984).

5 Figure 3 is a copy of a schematic representation comparing the deduced amino acid sequences of starch synthase II from wheat (wSSIIA, wSSIIB and wSSIID), maize (maizeSSIIa and maizeSSIIb; Harn et al., 1998), pea (peaSSII; Dry et al., 1992) and potato (potatoSSII; van der Leij et al., 1991). Identical amino acid residues among each of these sequences are indicated below the sequences with "\*". The alignments of maize SSIIa with maize SSIIb, and pea SSII and potato SSII are essentially as described in Harn et al. (1998) and Edwards et al. (1995). All sequences are aligned to position the transit peptide cleavage site below the arrow (\$\bar{\psi}\$) between residues 59 and 60 of the wSSIIA sequence. The wSSIIp1 sequence, the sequence of SGP-B1 (peptide3), and of eight conserved regions are annotated and underlined.

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Figure 4 is a copy of a photographic representation of a northern blot showing the expression of wheat wSSII mRNA in wheat plants. Total RNAs were isolated from leaves pre-anthesis florets and endosperm of the wheat cultivar "Gabo", grown under a photoperiod comprising 16 hours daylength, and at 18 °C during theday, and at 13 °C during the night cycle, and probed with the wSSIIp2 DNA fragment. The source of each RNA is indicated at the top of the Figure as follows: Lane 1, leaf; Lane 2, pre-anthesis florets; Lanes 3-11, endosperm at: 4 days post-anthesis (Lane 3); 6 days post-anthesis (Lane 4); 8 days post-anthesis (Lane 5); 10 days post-anthesis (Lane 6);12 days post-anthesis (Lane 7); 15 days post-anthesis (Lane 8); 18 days post-anthesis (Lane 9); 21 days post-anthesis (Lane 10); and 25 days post-anthesis (Lane 11).

Figure 5 is a copy of a photographic representation showing the localization of wheat starch synthase II genes on the wheat genome by PCR, using the primers ssIIc, ssIId and ssIIe in the amplification reaction. The nullisomic-tetrasomic genomic DNA of

wheat cv. Chinese Spring was used as template DNA. Lane D, *Triticum tauschii*; Lane AB, Accession line N7DT7B having no 7D chromosome and four copies of the 7B chromosome; Lane AD, Accession line N7BT7A having no 7B chromosome and four copies of the 7A chromosome; Lane BD, Accession line N7AT7B having no 7A chromosome and four copies of the 7B chromosome; Lane ABD, wheat cv. Chinese Spring. PCR products derived from each cDNA clone are labelled. The results indicate that the cDNA clones, wSSIIB, wSSIIA and wSSIID are derived from the B-, A- and D-genomes of wheat, respectively.

10 **Figure 6** is a copy of a photographic representation showing the purification of a wheat SSII genomic clone from the *T. tauschii* var. Strangulata (Accession No. CPI 110799) genomic library. A genomic clone, designated wSSII-8, was identified by hybridisation with the wSSIIp2 probe and purified through successive rounds of selection and hybridisation.

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Figure 7 is a copy of a photographic representation showing a Southern blot of BamHI-digested genomic clone DNAs identified in a primary screening using the wSSIIp2 probe, following hybridisation with wSSIIp4 probe DNA which is derived from the 5'-end of the wSSIIA cDNA clone. Lane 8 contains DNA derived from genomic clone wSSII-8 (see Figure 6). Hybridisation of clone wSSII-8 to thw wSSIIp4 probe suggests that this genomic clone contains the promoter region of the wSSII gene.

**Figure 8** is a schematic representation comparing the deduced amino acid Sequences of the maize, potato and wheat SSIII polypeptides.

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Figure 9 is a copy of a photographic representation showing the purification of a wheat SSIII genomic clone from a *T. tauschii* genomic library. A plaque was identified by hybridisation with a PCR-derived from the wSSIII.B3 gene (a) and purified through successive rounds of selection and hybridisation. The hybridisation of plaques from a third round of plaque purification is shown in (b).



Figure 10 is a copy of a photographic representation showing the expression of wheat wSSIII mRNA in wheat. Total RNAs were isolated from the endosperm of the wheat cultivars Wyuna (Panel a) and Gabo (Panel b) leaves pre-anthesis florets and endosperm of the wheat cultivar "Gabo", grown under a photoperiod comprising 16 hours daylength, and at 18 °C during the day cycle, and at 13 °C during the night cycle, and probed with the wSSIIIp1 DNA fragment derived from wSSIII.B3 cDNA. The source of each RNA is indicated at the top of the Figure as follows: Lane 1, endosperm at: 4 days post-anthesis; Lane 2, endosperm at 6 days post-anthesis; Lane 4, endosperm at 8 days post-anthesis; Lane 4, endosperm at 10 days post-anthesis;

10 Lane 5, endosperm at 12 days post-anthesis; Lane 6, endosperm at 15 days post-anthesis; Lane 7, endosperm at 18 days post-anthesis; Lane 8, endosperm at 21 days post-anthesis; Lane 9, endosperm at 25 days post-anthesis; and Lane 10, endosperm at 31 days post-anthesis (Panel a only). In panel (c), L refers to leaf RNA, and P refers to RNA from pre-anthesis florets derived from the cultivar Gabo.

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Figure 11 is a schematic representation showing the relationships between the primary amino acid sequences of starch synthases (SS) and glycogen synthase of *E. coli* (GS). The dendrogram was generated by the program PILEUP (Devereaux *et al.*, 1984). The amino acid sequences used for the analysis are those of the wheat SSIIA, wheat SSIIB, wheat SSIID, and wheat SSIII polypeptides of the present invention compared to the deduced amino acid sequences of wheat GBSS (Clark *et al.*, 1991), wheat SSI (Li *et al.*, 1999), rice GBSS (Okagaki, 1992), rice SSI (Baba *et al.*, 1993), maize GBSS (Kloesgen *et al.*, 1986), maize SSI (Knight *et al.*, 1998), maize SSIIa and maize SSIIb (Harn *et al.*, 1998), maize SSIII (Gao *et al.*, 1998), pea GBSS (Dry *et al.*, 1992), pea SSII (Dry *et al.*, 1992), potato GBSS (van der Leij *et al.*, 1991), potato SSI (Genbank accession number: STSTASYNT), potato SSII (Edwards *et al.*, 1995), potato SSIII (Abel *et al.*, 1996), and *E. coli* glycogen synthase (GS) (Kumar *et al.*, 1986). Five groups of enzymes included in the alignment are granule-bound starch synthase (GBSS), starch synthase-I (SSI), starcg synthase-II (SSII), starch synthase-III (SSIII)



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Figure 12 is a schematic representation showing the position of conserved regions within cereal starch synthase genes. Comparisons of cereal starch synthases were made based on their deduced amino acid sequences and 8 conserved regions identified. Conserved regions are shown in bold and transit peptides (where defined) in grey. The sequences included in the alignment are the wheat SSII-A1 and wheat SSIII polypeptides of the present invention; wheat GBSS (Ainsworth *et al.*, 1993); wheat SSI (Li *et al.*, 1999); maize SSIIa (Harn *et al.*, 1998); and maize dull-1(Gao *et al.*, 1998).

Figure 13 is a schematic representation showing the position of conserved amino acid sequences within four wheat starch synthase proteins. The eight highly-conserved regions between the wheat starch synthase polypeptides are underlined and annotated at the top of each group of amino acid sequences. The sequences included in the alignment are the wheat SSII-A1 and wheat SSIII polypeptides of the present invention; wheat GBSS (wGBSS; Yan et al., 1999); wheat SSI (wSS1; Li et al., 1999); wheat SSII (wSS2; SEQ ID NO:<400>4); and wheat SSIII (wSS3; SEQ ID NO:<400>8).

Figure 14 is a copy of a schematic representation of a gene map showing the 20 alignment of fragments 1 to 6 of the genomic SSIII gene (lower line) with the corresponding SSIII cDNA clone (uper line). Raised regions in the genomic clone fragments (lower line) represent protein-encoding regions of the gene.

### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

- One aspect of the present invention provides an isolated nucleic acid molecule which comprises a sequence of nucleotides which encodes, or is complementary to a nucleic acid molecule which encodes a wheat starch synthase polypeptide, protein or enzyme molecule or a functional subunit thereof selected from the following:
  - (i) a wheat starch synthase II (wSSII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence set forth

in any one of SEQ ID NOS: <400>2, <400>4, or <400>6; and

(ii) a wheat starch synthase III (wSSIII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence set forth in any one of SEQ ID NOS: <400>8 or <400>10.

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Alternatively or in addition, the isolated nucleic acid molecule of the present invention encodes a wheat starch synthase II (wSSII) polypeptide, protein or enzyme or functional subunit thereof and comprises a nucleotide sequence set forth in any one of SEQ ID NOS: <400>1, <400>3, or <400>5.

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Alternatively or in addition, the isolated nucleic acid molecule of the present invention encodes a wheat starch synthase III (wSSIII) polypeptide, protein or enzyme or functional subunit thereof and comprises a nucleotide sequence set forth in any one of SEQ ID NOS: <400>7 or <400>9.

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As used herein, the term "starch synthase" shall be taken to refer to any enzymatically-active peptide, polypeptide, oligopeptide, polypeptide, protein or enzyme molecule that is at least capable of transferring a glucosyl moiety from ADP-glucose to an  $\alpha$ -1,4-glucan molecule, or a peptide, polypeptide, oligopeptide or polypeptide fragment of such an enzymatically-active molecule.

The term "wheat starch synthase" refers to a starch synthase derived from hexaploid wheat or barley or a progenitor species, or a relative thereto such as the diploid *Triticum tauschii* or other diploid, tetraploid, aneuploid, polyploid, nullisomic, or a wheat/barley addition line, amongst others, the only requirement that the genomic DNA is at least about 80% identical to the genome of a wheat plant as determined by standard DNA melting curve analyses.

The term "starch synthase II" or "wSSII" or similar term shall be taken to refer to a starch synthase as hereinbefore defined that is detectable in the starch granule of a

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plant seed endosperm and possesses one or more properties selected from the group consisting of:

- (i) it is immunologically cross-reactive with the wheat starch granule proteins designated Sgp-B1 and/or Sgp-D1 and/or Sgp-A1, having estimated molecular weights of about 85 kDa to about 115 kDa;
- (ii) it is encoded by one of a homeologous set of genes localised on wheat chromosomes 7B or 7A or 7D:
- (iii) it is encoded by a nucleotide sequence that comprises at least about 15 nucleotides in length derived from any one or more of SEQ ID NOS: <400>1, <400>3, or <400>5 or a complementary nucleotide sequence thereto;
- (iv) it is encoded by a nucleotide sequence that is at least about 85% identical to one or more of the nucleotide sequences set forth in SEQ ID NOS: <400>1, <400>3, or <400>5 or a complementary nucleotide sequence thereto;
- (v) it comprises an amino acid sequence having at least about 85% identity to one or more of SEQ ID NOS:<400>2 or <400>4;
- (vi) it comprises at least about 5 contiguous amino acids, preferably at least about 10 contiguous amino acids, more preferably at least about 15 contiguous amino acids, even more preferably at least about 20 contiguous amino acids and still even more preferably at least about 25-50 contiguous amino acids of the amino acid sequences set forth in SEQ ID NOS:<400>2 or <400>4 or <400>6;and
- (vii) it which comprises a conserved amino acid sequence having at least 25% identity to an amino acid sequence selected from the group consisting of:
  - (a) KVGGLGDVVTS;
  - (b) GHTVEVILPKY;
  - (c) HDWSSAPVAWLYKEHY;
  - (d) GILNGIDPDIWDPYTD;
  - (e) DVPIVGIITRLTAQKG;
  - (f) NGQVVLLGSA;
- 30 (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and



## (h)TGGLVDTV

in addition to any one or more of (i) to (vi).

The term "starch synthase III" or "wSSIII" or similar term shall be taken to refer to a starch synthase as hereinbefore defined that possesses one or more properties selected from the group consisting of:

- (i) it is encoded by a nucleotide sequence that comprises at least about 15 nucleotides in length derived from any one or more of SEQ ID NOS:<400>7 or <400>9 or any one or more of SEQ ID NOS: <400>11 to <400>16 or a complementary nucleotide sequence thereto;
- (ii) it is encoded by a nucleotide sequence that is at least about 85% identical to one or more of the nucleotide sequences set forth in SEQ ID NOS: <400>7 or <400>9 or any one or more of SEQ ID NOS: <400>11 to <400>16 or a complementary nucleotide sequence thereto; and
- (iii) it comprises an amino acid sequence having at least about 85% identity to one or more of SEQ ID NOS:<400>8 or <400>10;
- (iv) it comprises at least about 5 contiguous amino acids, preferably at least about 10 contiguous amino acids, more preferably at least about 15 contiguous amino acids, even more preferably at least about 20 contiguous amino acids and still even more preferably at least about 25-50 contiguous amino acids of the amino acid sequences set forth in SEQ ID NOS:<400>8 or <400>10;and
- (v) which comprises a conserved amino acid sequence having at least 25% identity to an amino acid sequence selected from the group consisting of:
  - (a) KVGGLGDVVTS;

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- (b) GHTVEVILPKY;
- (c) HDWSSAPVAWLYKEHY;
- (d) GILNGIDPDIWDPYTD;
- (e) DVPIVGIITRLTAQKG;
- (f) NGQVVLLGSA;
- 30 (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and

# (h)TGGLVDTV

in addition to any one or more of (i) to (iv).

In a more preferred embodiment, the WSSII or WSSIII polypeptide encoded by the nucleic acid molecule of the present invention will comprise a substantial contiguous region of any one of SEQ ID NOS: <400>2, <400>4, <400>6, <400>8 or <400>10 or <400>17 sufficient to possess the biological activity of a starch synthase polypeptide.

For the purposes of nomenclature, the nucleotide sequence set forth in SEQ ID NO: 10 <400>1 relates to the cDNA molecule encoding the WSSII (i.e. Sgp-B1) polypeptide of wheat. The amino acid sequence of the corresponding polypeptide is set forth herein as SEQ ID NO:<400>2. The nucleotide sequence set forth in SEQ ID NO: <400>3 relates to the cDNA molecule encoding the WSSII (i.e. Sgp-A1) polypeptide of wheat. The amino acid sequence of the corresponding polypeptide is set forth 15 herein as SEQ ID NO:<400>4. The nucleotide sequence set forth in SEQ ID NO: <400>5 relates to the cDNA molecule encoding the WSSII (i.e. Sgp-D1) polypeptide of wheat. The amino acid sequence of the corresponding polypeptide is set forth herein as SEQ ID NO:<400>6. The nucleotide sequences set forth in SEQ ID NOs: <400>7 and <400>9 relate, respectively, to full-length and partial cDNA molecules 20 encoding the WSSIII polypeptide of wheat. The amino acid sequences of the corresponding polypeptides are set forth herein as SEQ ID NOs:<400>8 and <400>10. respectively. The nucleotide sequences set forth in SEQ ID NOs: <400>11 to <400>16 relates to fragments of the genomic gene encoding the WSSIII polypeptide of wheat, significant protein-encoding regions of which are described by reference to Table 3 25 and Figure 14.

Preferably, the isolated nucleic acid molecule of the present invention comprises a sequence of nucleotides which encodes, or is complementary to a nucleic acid molecule which encodes a wheat starch synthase III polypeptide, protein or enzyme molecule or a functional subunit thereof which comprises an amino acid sequence



which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>8 or <400>10 and more preferably, which additionally comprises which comprises one or more conserved amino acid sequences selected from the group consisting of:

- 5 (a) KVGGLGDVVTS;
  - (b) GHTVEVILPKY;
  - (c) HDWSSAPVAWLYKEHY;
  - (d) GILNGIDPDIWDPYTD;
  - (e) DVPIVGIITRLTAQKG;
- 10 (f) NGQVVLLGSA;
  - (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and
  - (h)TGGLVDTV.
- 15 The present invention clearly extends to homologues, analogues and derivatives of the wheat starch synthase II and III genes exemplified by the nucleotide sequences set forth herein as SEQ ID NOs:<400>1, <400>3, <400>5, <400>7, <400>9 and <400>11 to <400>16.
- 20 Preferred starch synthase genes may be derived from a naturally-occurring starch synthase gene by standard recombinant techniques. Generally, a starch synthase gene may be subjected to mutagenesis to produce single or multiple nucleotide substitutions, deletions and/or additions. Nucleotide insertional derivatives of the starch synthase gene of the present invention include 5' and 3' terminal fusions as well as intra-sequence insertions of single or multiple nucleotides. Insertional nucleotide sequence variants are those in which one or more nucleotides are introduced into a predetermined site in the nucleotide sequence although random insertion is also possible with suitable screening of the resulting product. Deletional variants are characterised by the removal of one or more nucleotides from the sequence. Substitutional nucleotide variants are those in which at least one nucleotide

in the sequence has been removed and a different nucleotide inserted in its place. Such a substitution may be "silent" in that the substitution does not change the amino acid defined by the codon. Alternatively, substituents are designed to alter one amino acid for another similar acting amino acid, or amino acid of like charge, polarity, or 5 hydrophobicity.

For the present purpose, "homologues" of a nucleotide sequence shall be taken to refer to an isolated nucleic acid molecule which is substantially the same as the nucleic acid molecule of the present invention or its complementary nucleotide sequence, notwithstanding the occurrence within said sequence, of one or more nucleotide substitutions, insertions, deletions, or rearrangements.

"Analogues" of a nucleotide sequence set forth herein shall be taken to refer to an isolated nucleic acid molecule which is substantially the same as a nucleic acid molecule of the present invention or its complementary nucleotide sequence, notwithstanding the occurrence of any non-nucleotide constituents not normally present in said isolated nucleic acid molecule, for example carbohydrates, radiochemicals including radionucleotides, reporter molecules such as, but not limited to DIG, alkaline phosphatase or horseradish peroxidase, amongst others.

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"Derivatives" of a nucleotide sequence set forth herein shall be taken to refer to any isolated nucleic acid molecule which contains significant sequence similarity to said sequence or a part thereof. Generally, the nucleotide sequence of the present invention may be subjected to mutagenesis to produce single or multiple nucleotide substitutions, deletions and/or insertions. Nucleotide insertional derivatives of the nucleotide sequence of the present invention include 5' and 3' terminal fusions as well as intra-sequence insertions of single or multiple nucleotides or nucleotide analogues. Insertional nucleotide sequence variants are those in which one or more nucleotides or nucleotide analogues are introduced into a predetermined site in the nucleotide sequence of said sequence, although random insertion is also possible with suitable



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screening of the resulting product being performed. Deletional variants are characterised by the removal of one or more nucleotides from the nucleotide sequence. Substitutional nucleotide variants are those in which at least one nucleotide in the sequence has been removed and a different nucleotide or nucleotide analogue 5 inserted in its place.

The present invention extends to the isolated nucleic acid molecule when integrated into the genome of a cell as an addition to the endogenous cellular complement of starch synthase genes, irrespective of whether or not the introduced nucleotide sequence is translatable or non-translatable to produce a polypeptide. The present invention clearly contemplates the introduction of additional copies of starch synthase genes into plants, particularly wheat plants, in the antisense orientation to reduce the expression of particular wheat starch synthase genes. As will be known to those skilled in the art, such antisense genes are non-translatable, notwithstanding that they can be expressed to produce antisense mRNA molecules.

The said integrated nucleic acid molecule may, or may not, contain promoter sequences to regulate expression of the subject genetic sequence.

- 20 Accordingly, the present invention clearly encompasses preferred homologues, analogues and derivatives that comprise a sequence of nucleotides which encodes, or is complementary to a nucleic acid molecule which encodes a wheat starch synthase polypeptide, protein or enzyme molecule or a functional subunit thereof selected from the following:
- 25 (i) a wheat starch synthase II (wSSII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, or <400>6;
  - (ii) a wheat starch synthase III (wSSIII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence which is at

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least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>8 or <400>10;and

- (iii) a wheat starch synthase polypeptide, protein or enzyme or functional subunit thereof which comprises a conserved amino acid sequence having at least 25% identity to an amino acid sequence selected from the group consisting of:
  - (a) KVGGLGDVVTS;
  - (b) GHTVEVILPKY;
  - (c) HDWSSAPVAWLYKEHY;
  - (d) GILNGIDPDIWDPYTD;
  - (e) DVPIVGIITRLTAQKG;
  - (f) NGQVVLLGSA;
  - (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and
  - (h)TGGLVDTV
- and wherein said wheat starch synthase polypeptide further comprises an amino acid sequence having at least about 85% identity overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, <400>6, <400>8 or <400>10.
- 20 Preferably, the isolated nucleic acid molecule encodes a starch synthase polypeptide, protein or enzyme that comprises two, more preferably three, more preferably four, more preferably five, more preferably six, more preferably seven and even more preferably eight of the conserved amino acid motifs listed *supra*. Even more preferably, the said amino acid motifs are located in a relative configuration such as that shown for the wheat SSII or wheat SSIII polypeptides listed in Figure 13 herein.

In a preferred embodiment, the isolated nucleic acid molecule encodes a starch synthase polypeptide, protein or enzyme having at least about 90% amino acid sequence identity to any one of SEQ ID NOS:<400>2, <400>4, <400>6, <400>8 or 30 <400>10, more preferably having at least about 95% or about 97% or about 99%



identity to any one of said amino acid sequences.

In an alternative embodiment, the present invention provides an isolated nucleic acid molecule which encodes a wheat starch synthase polypeptide, protein or enzyme 5 molecule or a functional subunit thereof, wherein said nucleic acid molecule comprises a nucleotide sequence having at least about 85% nucleotide sequence identity to any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>10 to <400>16, or a degenerate nucleotide sequence thereto or a complementary nucleotide sequence thereto.

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By "degenerate nucleotide sequence" is meant a nucleotide sequence that encodes a substantially identical amino acid sequence as a stated nucleotide sequence.

In a preferred embodiment, the isolated nucleic acid molecule comprises the nucleotide sequence set forth in any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or is at least about 90% identical, more preferably at least about 95% or 97% or 99% identical to all or a protein-encoding part thereof.

In an alternative embodiment, preferred homologues, analogues and derivatives of the nucleic acid molecule of the present invention encodes a wheat starch synthase polypeptide, protein or enzyme molecule or a functional subunit thereof and comprises a nucleotide sequence that is capable of hybridising under at least moderate stringency hybridisation conditions to at least about 30 contiguous nucleotides derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto.

For the purposes of defining the level of stringency, a low stringency is defined herein as being a hybridisation and/or a wash carried out in 6xSSC buffer, 0.1% (w/v) SDS at 28°C. Generally, the stringency is increased by reducing the concentration of SSC

buffer, and/or increasing the concentration of SDS and/or increasing the temperature of the hybridisation and/or wash. A moderate stringency comprises a hybridisation and/or a wash carried out in 0.2 x SSC-2 x SSC buffer, 0.1% (w/v) SDS at 42°C to 65°C, while a high stringency comprises a hybridisation and/or a wash carried out in 5 0.1xSSC-0.2 x SSC buffer, 0.1% (w/v) SDS at a temperature of at least 55°C. Conditions for hybridisations and washes are well understood by one normally skilled in the art. For the purposes of further clarification only, reference to the parameters affecting hybridisation between nucleic acid molecules is found in pages 2.10.8 to 2.10.16. of Ausubel *et al.* (1987), which is herein incorporated by reference.

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Those skilled in the art will be aware of procedures for the isolation of further wheat starch synthase genes to those specifically described herein or homologues, analogues or derivatives of said genes, for example further cDNA sequences and genomic gene equivalents, when provided with one or more of the nucleotide sequences set forth in SEQ ID NOs: <400>1, <400>3, <400>5, <400>7, <400>9, or <400>11 to <400>16. In particular, amplifications and/or hybridisations may be performed using one or more nucleic acid primers or hybridisation probes comprising at least 10 contiguous nucleotides and preferably at least about 20 contiguous nucleotides or 50 contiguous nucleotides derived from the nucleotide sequences set forth herein, to isolate cDNA clones, mRNA molecules, genomic clones from a genomic library (in particular genomic clones containing the entire 5' upstream region of the gene including the promoter sequence, and the entire coding region and 3'-untranslated sequences), and/or synthetic oligonucleotide molecules, amongst others. The present invention clearly extends to such related sequences.

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Accordingly, a second aspect of the present invention provides a method of isolating a nucleic acid molecule that encodes a starch synthase polypeptide, protein or enzyme having at least about 85% amino acid sequence identity to any one SEQ ID NOS:<400>2, <400>4, <400>6, <400>8 or <400>10 and/or which comprises a conserved amino acid sequence having at least 25% identity to an amino acid

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sequence selected from the group consisting of:

- (a) KVGGLGDVVTS;
- (b) GHTVEVILPKY;
- (c) HDWSSAPVAWLYKEHY;
- 5 (d) GILNGIDPDIWDPYTD;
  - (e) DVPIVGIITRLTAQKG;
  - (f) NGQVVLLGSA;
  - (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and
  - (h)TGGLVDTV,

## 10 said method comprising:

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- (i) hybridising a probe or primer comprising at least about 15 contiguous nucleotides in length derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto to single-stranded or double-stranded mRNA, cDNA or genomic DNA; and
- (ii) detecting the hybridised mRNA, cDNA or genomic DNA using a detecting means.

Preferably, the detecting means is a reporter molecule covalently attached to the probe or primer molecule or alternatively, a polymerase chain reaction format.

An alternative method contemplated in the present invention involves hybridising two nucleic acid "primer molecules" to a nucleic acid "template molecule" which comprises a related starch synthase gene or related starch synthase genetic sequence or a functional part thereof, wherein the first of said primers comprises contiguous nucleotides derived from any one or more of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16 and the second of said primers comprises contiguous nucleotides complementary to any one or more of SEQ ID NOS: <400>1, <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>9 or SEQ ID NOS: <400>11 to <400>16 and the second of SEQ ID NOS: <400>11 to <400>16 Specific nucleic acid molecule copies of the template molecule are amplified

enzymatically in a polymerase chain reaction, a technique that is well known to one skilled in the art.

In a preferred embodiment, each nucleic acid primer molecule is at least 10 nucleotides in length, more preferably at least 20 nucleotides in length, even more preferably at least 30 nucleotides in length, still more preferably at least 40 nucleotides in length and even still more preferably at least 50 nucleotides in length.

Furthermore, the nucleic acid primer molecules consists of a combination of any of the nucleotides adenine, cytidine, guanine, thymidine, or inosine, or functional analogues or derivatives thereof which are at least capable of being incorporated into a polynucleotide molecule without having an inhibitory effect on the hybridisation of said primer to the template molecule in the environment in which it is used.

15 Furthermore, one or both of the nucleic acid primer molecules may be contained in an aqueous mixture of other nucleic acid primer molecules, for example a mixture of degenerate primer sequences which vary from each other by one or more nucleotide substitutions or deletions. Alternatively, one or both of the nucleic acid primer molecules may be in a substantially pure form.

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The nucleic acid template molecule may be in a recombinant form, in a virus particle, bacteriophage particle, yeast cell, animal cell, or a plant cell. Preferably, the nucleic acid template molecule is derived from a plant cell, tissue or organ, in particular a cell, tissue or organ derived from a wheat or barley plant or a progenitor species, or a relative thereto such as the diploid *Triticum tauschii* or other diploid, tetraploid, aneuploid, polyploid, nullisomic, or a wheat/barley addition line, amongst others.

Those skilled in the art will be aware that there are many known variations of the basic polymerase chain reaction procedure, which may be employed to isolate a related starch synthase gene or related starch synthase genetic sequence when provided with



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the nucleotide sequences set forth herein. Such variations are discussed, for example, in McPherson *et al* (1991). The present invention extends to the use of all such variations in the isolation of related starch synthase genes or related starch synthase genetic sequences using the nucleotide sequences embodied by the present invention.

As exemplified herein, the present inventors have isolated several wheat starch synthase genes using both hybridisation and polymerase chain reaction approaches, employing novel probes and primer sequences to do so.

- 10 Accordingly, a third aspect of the invention provides an isolated probe or primer comprising at least about 15 contiguous nucleotides in length derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto.
- 15 Preferably, the probe or primer comprises a nucleotide sequence set forth in any one of SEQ ID NOS:<400>25 to <400>34.

The isolated nucleic acid molecule of the present invention may be introduced into and expressed in any cell, for example a plant cell, fungal cell, insect cell. animal cell, yeast cell or bacterial cell. Those skilled in the art will be aware of any modifications which are required to the codon usage or promoter sequences or other regulatory sequences, in order for expression to occur in such cells.

A further aspect of the invention provides a method of assaying for the presence or absence of a starch synthase isoenzyme or the copy number of a gene encoding same in a plant, comprising contacting a biological sample derived from said plant with an isolated nucleic acid molecule derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16, or any one of SEQ ID NOS:<400>25 to <400>34, or a complementary nucleotide sequence thereto for a time and under conditions sufficient for hybridisation to occur and then

detecting said hybridisation using a detection means.

The detection means according to this aspect of the invention is any nucleic acid based hybridisation or amplification reaction.

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The hexaploid nature of wheat prevents the straightforward identification of starch synthase allelic variants by hybridisation using the complete starch synthase-encoding sequence, because the similarities between the various alleles generally results in significant cross-hybridisation. Accordingly, sequence-specific hybridisation probes are required to distinguish between the various alleles. Similarly, wherein PCR is used to amplify specific allelelic variants of a starch synthase gene, one or more sequence-specific amplification primers are generally required. As will be apparent from the amino acid sequence comparisons provided herein, such as in Figures 3 and 13, non-conserved regions of particular wheat starch synthase polypeptides are particularly useful for the design of probes and primers that are capable of distinguishing between one or more starch synthase polypeptide isoenzyme or allelic variant. The present invention clearly contemplates the design of such probes and primers based upon the sequence comparisons provided herein.

20 In the performance of this embodiment of the present invention, the present inventors particularly contemplate the identification of wheat starch synthase null alleles or alternatively, mutations wherein specific amino acids are inserted or deleted or substituted, compared to one or more of the wheat SSII or SSIII alleles disclosed herein. Such null alleles and other allelic variants are readily identifiable using PCR screeening which employs amplification primers based upon the nucleotide and amino acid sequences disclosed herein for SSII and/or SSIII. Once identified, the various

mutations can be stacked or pyramided into one or more new wheat lines, such as by introgression and/or standard plant breeding and/or recombinant approaches (eg. transformation, transfection, etc) thereby producing a novel germplasm which exhibits

30 altered starch properties compared to existing lines. DNA markers based upon the

nucleotide and amino acid sequences disclosed herein for SSII and/or SSIII can be employed to monitor the stacking of genes into the new lines and to correlate the presence of particular genes with starch phenotypes of said lines.

of new DNA markers that reveal polymorphisms such as, for example, length polymorphisms, restriction site polymorphisms, and single nucleotide polymorphisms, amongst others, between wheat starch synthases and, in particular, between wheat GBSS and/or SSI and/or SSII and/or SSIII, or between allelic variants of one or more of said starch synthases, that can be used to identify the three genomes of hexaploid wheats (i.e., the A, B and D genomes).

Preferably, such DNA markers are derived from the intron region of a starch synthase gene disclosed herein, more preferably the wheat SSII and/or the wheat SSIII gene.

15 Those skilled in the art will be aware that such regions generally have a higher degree of variation than in the protein-encoding regions and, as a consequence, are particularly useful in identifying specific allelic variants of a particular gene, such as allelic variants contained in any one of the three wheat genomes, or alternatively or in addition, for the purpose of distinguishing between wheat GBSS, SSI, SSII or SSIII 20 genes.

A further approach contemplated by the present inventors is the design of unique isoenzyme-specific and/or allele-specific peptides based upon the amino acid sequence disclosed herein as SEQ ID NOS:<400>2 and/or <400>4 and/or <400>6 and/or <400>8 and/or <400>10, which peptides are then used to produce polyclonal or monoclonal antibodies by conventional means. Alternatively, the genes encoding these polypeptides or unique peptide regions thereof can be introduced in an expressible format into an appropriate prokaryotic or eukaroyotic expression system, where they can be expressed to produce the isoenzyme-specific and/or allele-specific peptides for antibody production. Such antibodies may also be used as markers for the

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purpose of both identifying parental lines and germplasms and monitoring the stacking of genes in new lines, using conventional immunoassays such as, for example, ELISA and western blotting.

5 A further aspect of the present invention utilises the above-mentioned nucleic acid based assay method in the breeding and/or selection of plants which express or do not express particular starch synthase isoenzymes or alternatively, which express a particular starch synthase isoenzyme at a particular level in one or more plant tissues. This aspect clearly extends to the selection of transformed plant material which 10 contains one or more of the isolated nucleic acid molecules of the present invention.

Yet another aspect of the present invention provides for the expression of the nucleic acid molecule of the present invention in a suitable host (e.g. a prokaryote or eukaryote) to produce full length or non-full length recombinant starch synthase gene products.

Hereinafter the term "starch synthase gene product" shall be taken to refer to a recombinant product of a starch synthase gene of the present invention.

20 Preferably, the recombinant starch synthase gene product comprises an amino acid sequence having the catalytic activity of a starch synthase polypeptide or a functional mutant, derivative part, fragment, or analogue thereof.

In a particularly preferred embodiment of the invention, the recombinant starch synthase gene product is selected from the following:

- (i) a wheat starch synthase II (wSSII) polypeptide, protein or enzyme or functional subunit thereof which comprises an amino acid sequence which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, or <400>6;
- (ii) a wheat starch synthase III (wSSIII) polypeptide, protein or enzyme or



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functional subunit thereof which comprises an amino acid sequence which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>8 or <400>10;and

- (iii) a wheat starch synthase polypeptide, protein or enzyme or functional subunit thereof which comprises a conserved amino acid sequence having at least 25% identity to an amino acid sequence selected from the group consisting of:
  - (a) KVGGLGDVVTS;
  - (b) GHTVEVILPKY;
  - (c) HDWSSAPVAWLYKEHY;
  - (d) GILNGIDPDIWDPYTD;
  - (e) DVPIVGIITRLTAQKG;
  - (f) NGQVVLLGSA;
  - (g)AGSDFIIVPSIFEPCGLTQLVAMRYGS; and
- 15 (h)TGGLVDTV

and which is at least about 85% identical overall to an amino acid sequence set forth in any one of SEQ ID NOS: <400>2, <400>4, <400>6, <400>8 or <400>10.

- 20 Accordingly, the present invention clearly extends to homologues, analogues and derivatives of the amino acid sequences set forth herein as SEQ ID NOS: <400>2, <400>4. <400>6, <400>8 and <400>10.
- In the present context, "homologues" of an amino acid sequence refer to those polypeptides, enzymes or proteins which have a similar catalytic activity to the amino acid sequences described herein, notwithstanding any amino acid substitutions, additions or deletions thereto. A homologue may be isolated or derived from the same or another plant species as the species from which the polypeptides of the invention are derived.

"Analogues" encompass polypeptides of the invention notwithstanding the occurrence of any non-naturally occurring amino acid analogues therein.

"Derivatives" include modified peptides in which ligands are attached to one or more of the amino acid residues contained therein, such as carbohydrates, enzymes, proteins, polypeptides or reporter molecules such as radionuclides or fluorescent compounds. Glycosylated, fluorescent, acylated or alkylated forms of the subject peptides are particularly contemplated by the present invention. Additionally, derivatives of an amino acid sequence described herein which comprises fragments or parts of the subject amino acid sequences are within the scope of the invention, as are homopolymers or heteropolymers comprising two or more copies of the subject polypeptides. Procedures for derivatizing peptides are well-known in the art.

Substitutions encompass amino acid alterations in which an amino acid is replaced with a different naturally-occurring or a non-conventional amino acid residue. Such substitutions may be classified as "conservative", in which an amino acid residue contained in a starch synthase gene product is replaced with another naturally-occurring amino acid of similar character, for example Gly↔Ala, Val↔Ile↔Leu, Asp↔Glu, Lys↔Arg, Asn↔Gln or Phe↔Trp↔Tyr.

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Substitutions encompassed by the present invention may also be "non-conservative", in which an amino acid residue which is present in a starch synthase gene product described herein is substituted with an amino acid with different properties, such as a naturally-occurring amino acid from a different group (eg. substituted a charged or hydrophobic amino acid with alanine), or alternatively, in which a naturally-occurring amino acid is substituted with a non-conventional amino acid.

Non-conventional amino acids encompassed by the invention include, but are not limited to those listed in Table 2.



Amino acid substitutions are typically of single residues, but may be of multiple residues, either clustered or dispersed.

Amino acid deletions will usually be of the order of about 1-10 amino acid residues, 5 while insertions may be of any length. Deletions and insertions may be made to the N-terminus, the C-terminus or be internal deletions or insertions. Generally, insertions within the amino acid sequence will be smaller than amino- or carboxy-terminal fusions and of the order of 1-4 amino acid residues.

10 A homologue, analogue or derivative of a starch synthase gene product as referred to herein may readily be made using peptide synthetic techniques well-known in the art, such as solid phase peptide synthesis and the like, or by recombinant DNA manipulations. Techniques for making substituent mutations at pre-determined sites using recombinant DNA technology, for example by M13 mutagenesis, are also well-known. The manipulation of nucleic acid molecules to produce variant peptides, polypeptides or proteins which manifest as substitutions, insertions or deletions are well-known in the art.

The starch synthase gene products described herein may be derivatized further by the inclusion or attachment thereto of a protective group which prevents, inhibits or slows proteolytic or cellular degradative processes. Such derivatization may be useful where the half-life of the subject polypeptide is required to be extended, for example to increase the amount of starch produced in the endosperm or alternatively, to increase the amount of protein produced in a bacterial or eukaryotic expression system.

25 Examples of chemical groups suitable for this purpose include, but are not limited to, any of the non-conventional amino acid residues listed in Table 2, in particular a D-stereoisomer or a methylated form of a naturally-occurring amino acid listed in Table 1. Additional chemical groups which are useful for this purpose are selected from the list comprising aryl or heterocyclic N-acyl substituents, polyalkylene oxide moieties, desulphatohirudin muteins, alpha-muteins, alpha-aminophosphonic acids, water-

soluble polymer groups such as polyethylene glycol attached to sugar residues using hydrazone or oxime groups, benzodiazepine dione derivatives, glycosyl groups such as beta-glycosylamine or a derivative thereof, isocyanate conjugated to a polyol functional group or polyoxyethylene polyol capped with diisocyanate, amongst others.

5 Similarly, a starch synthase gene product or a homologue, analogue or derivative thereof may be cross-linked or fused to itself or to a protease inhibitor peptide, to reduce susceptibility of said molecule to proteolysis.

In a particularly preferred embodiment, the percentage similarity to in any one of SEQ ID NOS: <400>2, <400>4, <400>6, <400>8 or <400>10 is at least about 90%, more preferably at least about 95%, even more preferably at least about 97% and even more preferably at least about 98%, or about 99% or 100%.

In a related embodiment, the present invention provides a "sequencably pure" form of the amino acid sequence described herein. "Sequencably pure" is hereinbefore described as substantially homogeneous to facilitate amino acid determination.

In a further related embodiment, the present invention provides a "substantially homogeneous" form of the subject amino acid sequence, wherein the term 20 "substantially homogeneous" is hereinbefore defined as being in a form suitable for interaction with an immunologically interactive molecule. Preferably, the polypeptide is at least 20% homogeneous, more preferably at least 50% homogeneous, still more preferably at least 75% homogeneous and yet still more preferably at least about 95-100% homogeneous, in terms of activity per microgram of total protein in the protein preparation.

To produce the recombinant polypeptide of the present invention, the coding region of a starch synthase gene described herein or a functional homologue, analogue or derivative thereof is placed operably in connection with a promoter sequence in the sense orientation, such that a starch synthase gene product is capable of being



expressed under the control of said promoter sequence.

In the present context, the term "in operable connection with" means that expression of the isolated nucleotide sequence is under the control of the promoter sequence with 5 which it is connected, regardless of the relative physical distance of the sequences from each other or their relative orientation with respect to each other.

Reference herein to a "promoter" is to be taken in its broadest context and includes the transcriptional regulatory sequences of a classical genomic gene, including the TATA box which is required for accurate transcription initiation, with or without a CCAAT box sequence and additional regulatory elements (i.e. upstream activating sequences, enhancers and silencers) which alter gene expression in response to developmental and/or external stimuli, or in a tissue-specific manner. A promoter is usually, but not necessarily, positioned upstream or 5', of a structural gene, the expression of which it regulates. Furthermore, the regulatory elements comprising a promoter are usually positioned within 2 kb of the start site of transcription of the gene.

In the present context, the term "promoter" is also used to describe a synthetic or fusion molecule, or derivative which confers, activates or enhances expression of a structural gene or other nucleic acid molecule, particularly in a plant cell and more preferably in a wheat plant or other monocotyledonous plant cell, tissue or organ. Preferred promoters may contain additional copies of one or more specific regulatory elements, to further enhance expression and/or to alter the spatial expression and/or temporal expression. For example, regulatory elements which confer copper inducibility may be placed adjacent to a heterologous promoter sequence, thereby conferring copper inducibility on the expression of said molecule.

Those skilled in the art will be aware that in order to obtain optimum expression of the starch synthase gene of the present invention, it is necessary to position said gene in an appropriate configuration such that expression is controlled by the promoter

sequence. Promoters are generally positioned 5' (upstream) to the genes that they control. In the construction of heterologous promoter/structural gene combinations it is generally preferred to position the promoter at a distance from the gene transcription start site that is approximately the same as the distance between that promoter and the gene it controls in its natural setting, i.e., the gene from which the promoter is derived. As is known in the art, some variation in this distance can be accommodated without loss of promoter function. Similarly, the preferred positioning of a regulatory sequence element with respect to a heterologous gene to be placed under its control is defined by the positioning of the element in its natural setting, i.e., the genes from which it is derived. Again, as is known in the art, some variation in this distance can also occur.

Examples of promoters suitable for expressing the starch synthase gene of the present invention include viral, fungal, bacterial, animal and plant derived promoters capable of functioning in prokaryotic or eukaryotic cells. Preferred promoters are those capable of regulating the expression of the subject starch synthase genes in plants cells, fungal cells, insect cells, yeast cells, animal cells or bacterial cells, amongst others. Particularly preferred promoters are capable of regulating expression of the subject nucleic acid molecules in monocotyledonous plant cells. The promoter may regulate the expression of the said molecule constitutively, or differentially with respect to the tissue in which expression occurs or, with respect to the developmental stage at which expression occurs, or in response to external stimuli such as physiological stresses, or plant pathogens, or metal ions, amongst others.

25 Accordingly, strong constitutive promoters are particularly preferred for the purposes of the present invention.

Examples of preferred promoters include the bacteriophage T7 promoter, bacteriophage T3 promoter, SP6 promoter, *lac* operator-promoter, *tac* promoter, SV40 late promoter, SV40 early promoter, RSV-LTR promoter, CMV IE promoter, CaMV 35S



promoter, SCSV promoter, SCBV promoter and the like.

Particularly preferred promoters operable in plant cells include, for example the CaMV 35S promoter, and the SCBV promoter. Those skilled in the art will readily be aware of additional promoter sequences other than those specifically described.

In a particularly preferred embodiment, the promoter may be derived from a genomic starch synthase gene. Preferably, the promoter sequence comprises nucleotide sequences that are linked *in vivo* to nucleotide sequences set forth in any one of SEQ ID NOs: <400>1, <400>3, <400>5, <400>7, <400>9, or any one of SEQ ID NOs: <400>11 to <400>16. By "linked *in vivo*" means that the promoter is present in its native state in the genome of a wheat plant where it controls expression of the starch synthase gene of the present invention.

- 15 Conveniently, genetic constructs are employed to facilitate expression of a starch synthase genetic sequence of the present invention or a functional derivative, part, homologue, or analogue thereof. To produce a genetic construct, the starch synthase gene of the invention is inserted into a suitable vector or episome molecule, such as a bacteriophage vector, viral vector or a plasmid, cosmid or artificial chromosome vector which is capable of being maintained and/or replicated and/or expressed in the host cell, tissue or organ into which it is subsequently introduced. The said genetic construct comprises the subject nucleic acid molecule placed operably under the control of a promoter sequence and optionally, a terminator sequence.
- 25 The term "terminator" refers to a DNA sequence at the end of a transcriptional unit which signals termination of transcription. Terminators are 3'-non-translated DNA sequences containing a polyadenylation signal, which facilitates the addition of polyadenylate sequences to the 3'-end of a primary transcript. Terminators active in bacteria, yeasts, animal cells and plant cells are known and described in the literature.
- 30 They may be isolated from bacteria, fungi, viruses, animals and/or plants.

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Examples of terminators particularly suitable for use in expressing the nucleic acid molecule of the present invention in plant cells include the nopaline synthase (NOS) gene terminator of *Agrobacterium tumefaciens*, the terminator of the Cauliflower mosaic virus (CaMV) 35S gene, and the *zein* gene terminator from *Zea mays*.

Genetic constructs will generally further comprise one or more origins of replication and/or selectable marker gene sequences.

The origin of replication can be functional in a bacterial cell and comprise, for example, the pUC or the CoIE1 origin. Alternatively, the origin of replication is operable in a eukaryotic cell, tissue and more preferably comprises the 2 micron ( $2\mu$ m) origin of replication or the SV40 origin of replication.

As used herein, the term "selectable marker gene" includes any gene which confers a phenotype on a cell in which it is expressed to facilitate the identification and/or selection of cells which are transfected or transformed with a genetic construct of the invention or a derivative thereof.

Suitable selectable marker genes contemplated herein include the ampicillin-resistance 20 gene (Amp'), tetracycline-resistance gene (Tc'), bacterial kanamycin-resistance gene (Kan'), is the zeocin resistance gene (Zeocin is a drug of bleomycin family which is trademark of InVitrogen Corporation), the AURI-C gene which confers resistance to the antibiotic aureobasidin Α, phosphinothricin-resistance gene, neomycin phosphotransferase gene (nptII), hygromycin-resistance gene, β-glucuronidase (GUS) 25 gene, chloramphenicol acetyltransferase (CAT) gene, green fluorescent proteinencoding gene or the luciferase gene, amongst others. Those skilled in the art will be aware of other selectable marker genes useful in the performance of the present invention and the subject invention is not limited by the nature of the selectable marker gene.



Usually, an origin of replication or a selectable marker gene suitable for use in bacteria is physically-separated from those genetic sequences contained in the genetic construct which are intended to be expressed or transferred to a eukaryotic cell, or integrated into the genome of a eukaryotic cell.

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Standard methods can be used to introduce genetic constructs into a cell, tissue or organ for the purposes of modulating gene expression. Particularly preferred methods suited to the introduction of synthetic genes and genetic constructs comprising same to eukaryotic cells include liposome-mediated transfection or transformation, 10 transformation of cells with attenuated virus particles or bacterial cells and standard procedures for the transformation of plant and animal cells, tissues, organs or organisms. Any standard means may be used for their introduction including cell mating, transformation or transfection procedures known to those skilled in the art or described by Ausubel *et al.* (1992).

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In a further embodiment of the present invention, the starch synthase genes of the present invention and genetic constructs comprising same are adapted for integration into the genome of a cell in which it is expressed. Those skilled in the art will be aware that, in order to achieve integration of a genetic sequence or genetic construct into the genome of a host cell, certain additional genetic sequences may be required. In the case of plants, left and right border sequences from the T-DNA of the *Agrobacterium tumefaciens* Ti plasmid will generally be required.

The invention further contemplates increased starch and/or modified starch composition in transgenic plants expressing the nucleic acid molecule of the invention in the sense orientation such that the activity of one or more starch synthase isoenzymes is increased therein. By increasing the level of one or more starch synthase isoenzymes, the deposition of starch in the amyloplast or chloroplast is increased and/or a modified starch granule structure is produced and/or starch composition is modified and/or the amylose/amylopectin ratio is altered in the plant.

Wherein it is desired to increase the synthesis of a particular starch synthase isoenzyme in a plant cell, the coding region of a starch synthase gene is placed operably behind a promoter, in the sense orientation, such that said starch synthase is expressed under the control of said promoter sequence. In a preferred embodiment, the starch synthase genetic sequence is a starch synthase genomic sequence, cDNA molecule or protein-coding sequence.

Wherein it is desirable to reduce the level of a particular starch synthase isoenzyme in a plant cell, the nucleic acid molecule of the present invention can be expressed in the antisense orientation, as an antisense molecule or a ribozyme molecule, under the control of a suitable promoter.

Alternatively, the nucleic acid molecule of the present invention may also be expressed in the sense orientation, in the form of a co-suppression molecule, to reduce the level of a particular starch synthase isoenzyme in a plant cell. As will be known to those skilled in the art, co-suppression molecules that comprise inverted repeat sequences of a target nucleic acid molecule provide optimum efficiency at reducing expression of said target nucleic acid molecule and, as a consequence, the present invention clearly contemplates the use of inverted repeat sequences of any one or more of the starch synthase genetic sequences exemplified herein, or inverted repeat sequences of a homologue, analogue or derivative of said starch synthase genetic sequences, to reduce the level of a starch synthase isoenzyme in a plant.

The expression of an antisense, ribozyme or co-suppression molecule comprising a starch synthase gene in a cell such as a plant cell, fungal cell, insect cell. animal cell, yeast cell or bacterial cell, may also increase the availability of carbon as a precursor for a secondary metabolite other than starch (e.g. sucrose or cellulose). By targeting the endogenous starch synthase gene, expression is diminished, reduced or otherwise lowered to a level that results in reduced deposition of starch in the amyloplast or chloroplast and/or leads to modified starch granule structure and/or composition



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and/or altered amylose/amylopectin ratio.

Accordingly, a further aspect of the present invention provides a method of modifying the starch content and/or starch composition of one or more tissues or organs of a plant, comprising expressing therein a sense molecule, antisense molecule, ribozyme molecule, co-suppression molecule, or gene-targeting molecule having at least about 85% nucleotide sequence identity to any one of any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto for a time and under conditions sufficient for the enzyme activity of one or more starch synthase isoenzymes to be modified. This aspect of the invention clearly extends to the introduction of the sense molecule, antisense molecule, ribozyme molecule, co-suppression molecule, or genetargeting molecule to isolated plant cells, tissues or organs or organelles by cell fusion or transgenic means and the regeneration of intact plants therefrom.

Co-suppression is the reduction in expression of an endogenous gene that occurs when one or more copies of said gene, or one or more copies of a substantially similar gene are introduced into the cell, preferably in the form of an inverted repeat structure.

The present inventors have discovered that the genetic sequences disclosed herein are capable of being used to modify the level of starch when expressed, particularly when expressed in plants cells. Accordingly, the present invention clearly extends to the modification of starch biosynthesis in plants, in particular wheat or barley plants or a progenitor plant species, or a relative thereto such as the diploid *Triticum tauschii* or other diploid, tetraploid, aneuploid, polyploid, nullisomic, or a wheat/barley addition line, amongst others.

In particular, the present invention contemplates decreased starch production and/or modified starch composition in transgenic plants expressing the nucleic acid molecule of the invention in the antisense orientation or alteratively, expressing a ribozyme or

co-suppression molecule comprising the nucleic acid sequence of the invention such that the activity of one or more starch synthase isoenzymes is decreased therein.

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In the context of the present invention, an antisense molecule is an RNA molecule which is transcribed from the complementary strand of a nuclear gene to that which is normally transcribed to produce a "sense" mRNA molecule capable of being translated into a starch synthase polypeptide. The antisense molecule is therefore complementary to the mRNA transcribed from a sense starch synthase gene or a part thereof. Although not limiting the mode of action of the antisense molecules of the present invention to any specific mechanism, the antisense RNA molecule possesses the capacity to form a double-stranded mRNA by base pairing with the sense mRNA, which may prevent translation of the sense mRNA and subsequent synthesis of a polypeptide gene product.

Ribozymes are synthetic RNA molecules which comprise a hybridising region complementary to two regions, each of at least 5 contiguous nucleotide bases in the target sense mRNA. In addition, ribozymes possess highly specific endoribonuclease activity, which autocatalytically cleaves the target sense mRNA. A complete description of the function of ribozymes is presented by Haseloff and Gerlach (1988) and contained in International Patent Application No. WO89/05852.

The present invention extends to ribozyme which target a sense mRNA encoding a native starch synthase gene product, thereby hybridising to said sense mRNA and cleaving it, such that it is no longer capable of being translated to synthesise a functional polypeptide product.

According to this embodiment, the present invention provides a ribozyme or antisense molecule comprising at least 5 contiguous nucleotide bases derived from any one of



SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto or a homologue, analogue or derivative thereof, wherein said antisense or ribozyme molecule is able to form a hydrogen-bonded complex with a sense mRNA encoding a starch synthase gene product to reduce translation thereof.

In a preferred embodiment, the antisense or ribozyme molecule comprises at least 10 to 20 contiguous nucleotides derived from any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto or a homologue, analogue or derivative thereof. Although the preferred antisense and/or ribozyme molecules hybridise to at least about 10 to 20 nucleotides of the target molecule, the present invention extends to molecules capable of hybridising to at least about 50-100 nucleotide bases in length, or a molecule capable of hybridising to a full-length or substantially full-length mRNA encoded by a starch synthase gene.

Those skilled in the art will be aware of the necessary conditions, if any, for selecting or preparing the antisense or ribozyme molecules of the invention.

- 20 It is understood in the art that certain modifications, including nucleotide substitutions amongst others, may be made to the antisense and/or ribozyme molecules of the present invention, without destroying the efficacy of said molecules in inhibiting the expression of a starch synthase gene. It is therefore within the scope of the present invention to include any nucleotide sequence variants, homologues, analogues, or fragments of the said gene encoding same, the only requirement being that said nucleotide sequence variant, when transcribed, produces an antisense and/or ribozyme molecule which is capable of hybridising to a sense mRNA molecule which encodes a starch synthase gene product.
- 30 Gene targeting is the replacement of an endogenous gene sequence within a cell by

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a related DNA sequence to which it hybridises, thereby altering the form and/or function of the endogenous gene and the subsequent phenotype of the cell. According to this embodiment, at least a part of the DNA sequence defined by any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16 may be introduced into target cells containing an endogenous gene that encodes a particular starch synthase isoenzyme, thereby replacing said endogenous gene. According to this embodiment, the polypeptide product of the gene targetting molecule generally encodes a starch synthase isoenzyme that possesses different catalytic activity to the polypeptide product of the endogenous gene, producing in turn modified starch content and/or composition in the target cell.

The present invention extends to genetic constructs designed to facilitate expression of a sense molecule, an antisense molecule, ribozyme molecule, co-suppression molecule, or gene targeting molecule of the present invention. The requirements for expressing such molecules are similar to those for expressing a recombinant polypeptide as described *supra*.

The present invention further extends to the production and use of starches produced by the application of the novel genes described herein.

Starch hydrolysates or undegraded starches are both useful in industry and, as a consequence, the present invention is useful in applications relating to the use of both

starch hydrolysates and undegraded starches. By "starch hydrolysates" is meant the glucose and glucan components that are obtainable by the enzymatic or chemical

25 degradation of starch in chemical modifications and processes, such as fermentation.

For example, starch produced by plants expressing the sense, antisense, cosuppression, gene-targetting or ribozyme molecules of the present invention may exhibit modified viscosities and/or gelling properties of its glues when compared to 30 starch derived from wild-type plants. Native starches produced by the performance of



the inventive method are useful as an additive in the following: (i) foodstuffs, for the purpose of increasing the viscosity or gelling properties of food; (ii) in non-foodstuffs. such as an adjuvant or additive in the paper and cardboard industries, for retention or as a size filler, or as a solidifying substance or for dehydration, or film coating. 5 amongst others; (iii) in the adhesive industry as pure starch glue, as an additive to synthetic resins and polymer dispersions, or as an extenders for synthetic adhesives: (iv) in the textile and textile care industries to strengthen woven products and reduce burring or to thicken dye pastes; (v) in the building industry, such as a binding agent in the production of gypsum plaster boards, or for the deceleration of the sizing 10 process; (vi) in ground stabilization or for the temporary protection of ground particles against water in artificial earth shifting; (vii) as a wetting agent in plant protectants and fertilizers; (viii) as a binding agent in drugs, pharmaceuticals and medicated foodstuff such as vitamins, etc; (ix) as an additive in coal and briquettes; (xi) as a flocculent in the processing of coal ore and slurries; (xii) as a binding agent in casting processes 15 to increase flow resistance and improve binding strength; and (xiii) to improve the technical and optical quality of rubber and plastic products. Additional applications are not excluded.

A further aspect of the present invention provides an isolated promoter that is operable in the endosperm of a monocotyledonous plant cell, tissue or organ, and preferably in the endosperm of a monocotyledonous plant cell, tissue or organ. According to this embodiment, it is preferred that the promoter is derived from a starch synthase gene of the present invention, such as a promoter that is linked *in vivo* to any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto.

In a particularly preferred embodiment, the promoter comprises a nucleotide sequence derivable from the 5'-upstream region of SEQ ID NO:<400>11 or a complementary nucleotide sequence thereto, an more preferably comprises nucleotides 1 to about 287 of SEQ ID NO:<400>11 or nucleotides 1 to about 287 of SEQ ID NO:<400>11 or a

complementary nucleotide sequence thereto. The present invention clearly extends to promoter sequences that comprise further nucleotide sequences in the region upstream of the stated nucleotide sequence that are linked *in vivo* to said nucleotide sequence in the wheat genome.

5

In a related embodiment, the promoter sequence of the present invention will further comprise an exon sequence derived from a starch synthase gene, for example nucleotides 260 to 385 of SEQ ID NO:<400>11 or a complementary nucleotide sequence thereto. Those skilled in the art will be aware that the inclusion of such nucleotide sequences may increase the expression of a heterologous structural gene, the expression of which is controlled thereby.

The present invention further extends to the expression of any structural gene operably under the control of the starch synthase promoter sequence exemplified herein or a functional homologue, analogue or derivative of said promoter sequence.

As with other embodiments described herein for expression in cells, a genetic construct may be employed to effect said expression and the present invention clearly extends to said genetic constructs.

20

The polypeptide encoded by the structural gene component may be a reporter molecule which is encoded by a gene such as the bacterial β-glucuronidase gene or chloramphenicol acetyltransferase gene or alternatively, the firefly luciferase gene. Alternatively, wherein it is desirable to alter carbon partitioning within the endosperm, the polypeptide may be an enzyme of the starch sucrose biosynthetic pathways. Preferably, the promoter sequence is used to regulate the expression of one or more of the starch synthase genes of the present invention or a sense, antisense, ribozyme, co-suppression or gene-targetting molecule comprising or derived from same.

30 Recombinant DNA molecules carrying the aforesaid nucleic acid molecule of the



present invention or a sense, antisense, ribozyme, gene-targetting or co-suppression molecule and/or genetic construct comprising same, may be introduced into plant tissue, thereby producing a "transgenic plant", by various techniques known to those skilled in the art. The technique used for a given plant species or specific type of plant 5 tissue depends on the known successful techniques. Means for introducing recombinant DNA into plant tissue include, but are not limited to, transformation (Paszkowski et al., 1984), electroporation (Fromm et al., 1985), or microinjection of the DNA (Crossway et al., 1986), or T-DNA-mediated transfer from Agrobacterium to the plant tissue. Representative T-DNA vector systems are described in the following 10 references: An et al.(1985); Herrera-Estrella et al. (1983a,b); Herrera-Estrella et al. (1985). Once introduced into the plant tissue, the expression of the introduced gene may be assayed in a transient expression system, or it may be determined after selection for stable integration within the plant genome. Techniques are known for the in vitro culture of plant tissue, and in a number of cases, for regeneration into whole 15 plants. Procedures for transferring the introduced gene from the originally transformed plant into commercially useful cultivars are known to those skilled in the art.

In general, plants are regenerated from transformed plant cells or tissues or organs on hormone-containing media and the regenerated plants may take a variety of forms, such as chimeras of transformed cells and non-transformed cells; clonal transformants (e.g., all cells transformed to contain the expression cassette); grafts of transformed and untransformed tissues (e.g., a transformed root stock grafted to an untransformed scion in citrus species). Transformed plants may be propagated by a variety of means, such as by clonal propagation or classical breeding techniques. For example, a first generation (or T1) transformed plants may be selfed to give homozygous second generation (or T2) transformed plants, and the T2 plants further propagated through classical breeding techniques.

Accordingly, a still further aspect of the present invention contemplates a transgenic 30 plant comprising an introduced sense molecule, antisense molecule, ribozyme

molecule, co-suppression molecule, or gene-targeting molecule having at least about 85% nucleotide sequence identity to any one of any one of SEQ ID NOS: <400>1, <400>3, <400>5, <400>7 or <400>9 or any one of SEQ ID NOS: <400>11 to <400>16, or a complementary nucleotide sequence thereto or a genetic construct comprising same.

The present invention further extends to those plant parts, propagules and progeny of said transgenic plant or derived therefrom, the only requirement being that said propagules and progeny also carry the introduced nucleic acid molecule(s).

10

The present invention is further described by reference to the following non-limiting examples.

15

### **EXAMPLE 1**

### Plant material

Genetic stocks of hexaploid bread wheat *Triticum aestivum* cv. Chinese Spring with various chromosome additions and deletions were kindly supplied by Dr E. Lagudah (CSIRO Plant Industry, Canberra) and derived from stocks described in Sears and Miller (1985). The hexaploid (*Triticum aestivum*) wheats cv Gabo and cv Wyuna were grown in controlled growth cabinet conditions (18 °C day and 13° C night, with a photoperiod of 16 h). Wheat leaves and florets prior to anthesis, and endosperm were collected over the grain filling period, immediately frozen in liquid nitrogen and stored at -80°C until required.

25

### **EXAMPLE 2**

## Gel Electrophoresis, Antibodies and Immunoblotting

Monoclonal antibodies against the Sgp-1 proteins, and their use in the immunoblotting of SDS-PAGE gels have been described previously (Rahman *et al.*, 1995).



# Preparation of total RNA from wheat

Total RNA was isolated from the leaf, floret and endosperm tissues of wheat essentially as described by Higgins *et al.* (1976) or Rahman *et al.* (1998). RNA was quantified by UV absorption and by separation in 1.4% (w/v) agarose-formaldehyde gels which were then visualised under UV light after staining with ethidium bromide.

### **EXAMPLE 4**

## Construction and screening of cDNA libraries

10 A first cDNA library, an expression cDNA library of wheat endosperm, was constructed from mRNA isolated from wheat cv Chinese Spring. RNA from 5, 7, 9, 11 and 13 days after anthesis was pooled and random primers were used for the first strand of cDNA synthesis. Monoclonal antibodies against 100 -105 kDa proteins in wheat starch granules (Rahman *et al.*, 1995) were used for immunoscreening of the expression 15 cDNA library.

A second cDNA library was constructed from the endosperm mRNA of the hexaploid *Triticum aestivum* cultivar Wyuna, 8 - 12 days after anthesis, as described by Rahman *et al.* (1997). This library was screened with a 85-bp cDNA fragment, wSSIIp1, which was obtained by immunoscreening of the expression cDNA library as described above. The wSSIIp1 probe corresponded to nucleotide positions 988 to 1072 of wSSIIB (SEQ ID NO:<400>1) at the hybridisation conditions as described earlier (Rahman *et al.*, 1998).

25 A third cDNA library was constructed from RNA from the endosperm of the hexaploid *Triticum aestivum* cultivar Rosella as described by Rahman *et al.* (1997). This library was screened with a 347-bp cDNA fragment, wSSIIIp1 for the first screening and a 478-bp cDNA fragment wSSIIIp3 for the second screening ( PLEASE ADVISE-nucleotides 2469 to 2947 of SEQ ID NO:<400>7) using the hybridisation conditions described herein.

## Construction and screening of Triticum tauschii genomic library

The genomic library used in this study, prepared from *Triticum tauschii*, var strangulata, (Accession Number CPI 110799), has been described in Rahman et al., 5 (1997). Of all the accessions of *T. tauschii* surveyed, DNA marker analysis suggests that the genome of CPI 110799 is the most closely related to the D genome of hexaploid wheat (Lagudah et al., 1991).

Hybridisations were carried out in 25% formamide, 6 x SSC, 0.1% SDS at 42°C for 16 hours, then filters were washed 3 times using 2 x SSC containing 0.1% SDS at 65°C for 1 hour per wash.

For the isolation of a genomic wSSII clone, the probe comprised the PCR-derived DNA fragment wSSIIp2 and positive-hybridising plaques were digested using the restriction enzyme *BamHI*, separated on a 1% agarose gel, transferred to nitrocellulose membrane and hybridised to probe wSSIIp4 comprising nucleotides 1 to 367 of the wSSIIA cDNA clone, using the conditions described by Rahman *et al.* (1997).

For the isolation of a genomic wSSIII clone, plaques hybridising to the PCR-derived 20 DNA fragment wSSIIIp1 from clone wSSIII.B3 (i.e. nucleotides 3620 to 3966 of SEQ ID NO:<400>7) were selected and re-screened until plaque-purified.

#### **EXAMPLE 6**

# 25 **DNA sequencing and analysis**

DNA sequencing was performed using the automated ABI system with dye terminators as described by the manufacturers. DNA sequences were analysed using the GCG suite of programs (Devereaux *et al.*, 1984).



## DNA and RNA analysis

DNA was isolated and analysed as previously described (Maniatis *et al.*, 1982; Rahman *et al.*, 1998). Approximately 20 µg of DNA was digested with restriction enzymes *Bam*HI, *Dra*I and *Eco*RI, separated on a 1% agarose gel and transferred to reinforced nitrocellulose membranes (BioRad) and hybridised with <sup>32</sup>P-labelled DNA probe, either wSSIIIp1, corresponding to nucleotides 3620 to 3966 of the wheat SSIII gene, or alternatively, with the entire wSSII cDNA clone. DNA fragment probes were labelled with the Rapid Multiprime DNA Probe Labelling Kit (Promega).

10

The hybridisation and wash conditions were performed as described in Rahman *et al.* (1997). For RNA analysis, 10 μg of total RNA was separated in a 1.4% agarose-formaldehyde gel and transferred to a Hybond N+ membrane (Amersham), and hybridised with cDNA probe at 42°C as previously described by Khandjian *et al.*, (1987) or Rahman *et al.*, (1998). After washing for 30 minutes at 65°C with 2x SSC, 0.1% SDS; followed by three washes of 40 minutes at 65°C with 0.2x SSC, 1% SDS, the membranes were visualised by overnight exposure at -80°C with Kodak MR X-ray film.

20

## **EXAMPLE 8**

# Expression of wheat Sgp-1 polypeptides in the wheat endosperm

The development and use of monoclonal antibodies to the Sgp-1 proteins has been described previously (Rahman *et al.*, 1995). These antibodies were used by the present inventors to characterise the expression and localisation of the Sgp-1 proteins.

The proteins found in the matrix of the wheat starch granule are shown in Figure 1, lane1. The remaining lanes show an immunoblot of proteins from the soluble phase (Figure 1; lanes 2-4) and the starch granule (Figure 1; lanes 5-7), respectively, 30 following SDS-PAGE. In addition to cross-reactivity with the 100-105 kDa proteins, a

weak cross-reaction with a 50 kDa protein in both the granule and the soluble fractions were observed (Figure 1). The Sgp-1 polypeptides are present in the starch granule throughout endosperm development (Figure 1; lanes 5-7, also see Rahman *et al.*, 1995). However, as the endosperms matures, there is a reduction in the amount of Sgp-1 protein found in the soluble fraction. Lane 4 shows that by 25 days after anthesis, the level of these proteins in the soluble fraction is substantially reduced. This observation is consistent with previous results from Rahman *et al.*, (1995), who suggested that the Sgp-1 proteins were exclusively granule bound based on studies of granules from endosperm in mid-late stages endosperm development, however, these results suggest that the partitioning of these proteins between the granule and the soluble phase changes during development.

### **EXAMPLE 9**

# Isolation of cDNA clones encoding wheat starch synthase II (wSSII) proteins:

15 Monoclonal antibodies against Sgp-1 polypeptides (Rahman *et al.*, 1995) were used to probe the expression library described in Example 4 (i.e. the first cDNA library). Three immunoreactive plaques were identified and sequenced. One clone, designated wSSIIp1, contained an 85-bp cDNA insert with homology to maize SSIIa (Harn *et al.*, 1998).

20

DNA from the wSSIIp1 clone was used as a probe in the hybridisation screening of the second cDNA library, prepared from *Triticum aestivum* cultivar Wyuna endosperm RNA as described in Example 4. Ten hybridising cDNA clones were selected and sequenced. On the basis of the DNA sequences obtained, the 10 cDNA clones can be classified into three groups. Group 1 contains 7 cDNA clones, group 2 contains 2 cDNA clones and group 3 contains 1 cDNA clone.

The longest clone from group 1 (designated wSSIIB) is 2939 bp in length (SEQ ID NO:<400>1) and encodes a 798 -amino acid polypeptide starting at nucleotide 176 and terminating at nucleotide 2572 (SEQ ID NO:<400>2).



The longest clone from group 2 (designated wSSIIA) is 2807 bp in length (SEQ ID NO:<400>3) and encodes a 799 -amino acid polypeptide starting at nucleotide 89 and terminating at nucleotide 2488 (SEQ ID NO:<400>4).

5 The cDNA from group 3 is a partial cDNA clone (designated wSSIID), which is 2107 bp in length (SEQ ID NO:<400>5) and encodes a 597 -amino acid polypeptide starting at nucleotide 1 and terminating at nucleotide 1794 (SEQ ID NO:<400>6). The encoded polypeptide is approximately a 200 amino acid residues shorter than that of polypeptides encoded by longest clones of group 1 or 2 clones, respectively (Figure 10 2).

Comparison of the three cDNA clones, wSSIIB, wSSIIA and wSSIID shows that they share 95.7% to 96.6% identity at amino acid level, with variation at 44 amino acid positions between the three sequences (Figure 3). Of the 44 amino acid changes 15 between these sequences, 31 changes occur in the N-terminal region (residues 1 to 300), 10 changes occur in the central region (residues 301 to 729) and 3 changes occur in the C-terminal region (residues 730 to 799). The wSSIIA polypeptide (799 amino acid residues) and wSSIIB polypeptide (798 amino acid residues) sequences differ in length by a single amino acid residue, due to the deletion of Asp-69 from the wSSIIB polypeptidesequence.

A comparison of the nucleotide sequences of the wSSIA, wSSIIB and wSSIID cDNA clones with the nucleotide sequence of the wSSIIp1 cDNA obtained by immunoscreening confirms that the wSSIIp1 sequence is found in each cDNA. The 25 peptide encoded by the wSSIIp1 cDNA clone corresponds to amino acid residues in the region from residue 272 to residue 298 of the wSSIIA polypeptide, and to amino acid residues in the region from residue 271 to residue 297 of the wSSIIB polypeptide (see Figure 3). Thus, the peptide epitope encoded by wSSIIp1 that reacts with the anti-Sgp-1 monoclonal antibodies can therefore be localised to this region of the wSSIID wSSIIA and wSSIIB polypeptides and to the corresponding region of the wSSIID

polypeptide.

Notwithstanding that a region having about 63% amino acid sequence identity to the peptide epitope encoded by clone wSSIIp1 is found in the maize SSIIa polypeptide 5 (Figure 3), the degree of amino acid conservation between maize and wheat sequences in this region of the polypeptide is insufficient for immunological cross-reactivity to occur between these species using the monoclonal antibodies to the wheat Sgp-1 proteins described by Rahman *et al.* (1995). Additionally, this peptide epitope is not found in granule-bound starch synthases, SSI, or SSIII (data not shown).

10

The wSSIIB cDNA (SEQ ID NO:<400>1) encodes an amino acid sequence comprising the peptide motif AAGKKDAGID (SEQ ID NO:<400>18) between residues 60 and 69 of SEQ ID NO:<400>2 (Figure 3) which, with the exception of the second residue, is identical to the N-terminal of the 100 kDa (A<sup>T</sup>/<sub>L</sub>GKKDAGID: SEQ ID NOs:<400>19 and 20) protein (Sgp-B1) from the wheat starch granule (note that the sequence given in Rahman *et al.*, 1995 (A<sup>T</sup>/<sub>L</sub>GKKDAL: SEQ ID NOs:<400>21 and 22 ) has been revised following further amino acid sequence analysis).

The wSSIIA cDNA clone (SEQ ID NO:<400>3) encodes an amino acid sequence comprising the peptide motif AAGKKDARVDDAA (SEQ ID NO: <400>23) at residues 60 to 73 of SEQ ID NO:<400>4, which is about 66% identical to the N-terminal amino acid sequence (i.e. ALGKKDAGIVDGA: SEQ ID NO: <400>24) of the 104 kDa and 105 kDa starch granule proteins, Sgp-D1 and Sgp-A1 respectively, as determined by sequence analysis of isolated protein (Rahman *et al.*, 1995).

25

Furthermore, Takaoka *et al.* (1997) reported the amino acid sequences of 3 polypeptides obtained from sequencing starch granule proteins derived from the Sgp-1 proteins. Peptide 3 described by Takaoka *et al.* (1997) corresponds to amino acid residues 378 to 387 of the amino acid sequence of the wSSIIA cDNA (SEQ ID NO:<400>4; Figure 3). Peptides 1 and 2 described by Takaoka *et al.* (1997) could not



be detected in the amino acid sequences of the wSSII cDNA clones of the present invention, however peptide 1 of Takaoka *et al.* (1997) can be found in the amino acid sequences of SSI from maize, rice, wheat and potato (data not shown).

5 Denyer et al. (1995) demonstrated that the Sgp-1 proteins possess starch synthase activity and, as a consequence, the wSSIIB, wSSIA and wSSIID cDNA clones encode starch synthase enzymes that are differentially expressed in a developmentally-regulated manner in both the soluble and granule-bound fractions of the endosperm (Figure 1). Based on the nomenclature suggested by Harn et al. (1998), it is appropriate to describe the Sgp-1 proteins as "starch synthases" rather than "granule-bound starch synthases".

#### **EXAMPLE 10**

# Analysis of wheat starch synthase II mRNA expression

The mRNA for wheat starch synthase II could be detected in leaves, pre-anthesis florets and endosperm of wheat when total RNAs isolated from these tissue were probed with a PCR probe, wSSIIp2, corresponding to nucleotide positions 1435 to 1835 bp of wSSIIB-cDNA (SEQ ID NO:<400>1; Figure 4). Unlike wSSI, which could not be detected in wheat leaves derived from plants grown under the same conditions, wSSII genes are highly-expressed in the leaves (Figure 4, lane 1), and expressed at an intermediate level in pre-anthesis florets (Figure 4, lane 2), and at much lower levels in developing wheat endosperm cells (Figure 4, lanes 3-11). In contrast, the maize SSIIa is expressed predominantly in the endosperm, whilst the maize SSIIb is detected mainly in the leaf, albeit at low levels (Harn *et al.*, 1998).

The wSSII mRNA was detectable in the endosperm 6 days after anthesis and mRNA levels increase between 8 and 18 days post-anthesis, after which time levels of mRNA decline.

15

Southern blotting experiments in wheat demonstrated that the wSSIIp2 probe used detected only a single copy of the SSII gene in each genome (data not shown). Thus, it is unlikely that this probe cross-hybridised with mRNAs encoded by genes other than wSSII.

5

### **EXAMPLE 11**

## Chromosomal localization of the wheat wSSII genes.

10 I. Amplification of specific cDNA regions of wheat starch synthase II using PCR Two PCR products, wSSIIp2 and wSSIIp3 were amplified from the cDNA clone wSSIIB and used for the northern hybridisation and Southern hybridisation, respectively.

The primers sslla (5' TGTTGAGGTTCCATGGCACGTTC 3': SEQ ID NO: <400>25)

15 and ssllb (5' AGTCGTTCTGCCGTATGATGTCG 3': SEQ ID NO: <400>26) were used to amplify the cDNA fragment wSSllp2 (i.e. nucleotide positions 1435 to 1835 of SEQ ID NO: <400>1).

The primers ssllc (5' CCAAGTACCAGTGGTGAACGC 3': SEQ ID NO: <400>27) and sslld (5' CGGTGGGATCCAACGGCCC 3': SEQ ID NO: <400>28) were used to amplify the cDNA fragment wSSllp3 (i.e. nucleotide positions 2556 to 2921 of SEQ ID NO:<400>1).

The amplification reactions were performed using a FTS-1 thermal sequencer (Corbett, 25 Australia) for 1 cycle of 95°C for 2 minutes; 35 cycles of 95°C for 30 seconds, 60°C for 1 minutes, 72°C for 2 minutes and 1 cycle of 25°C for 1 minute.

II. PCR and nucleotide sequence analysis of 3' sequences of wheat SSII genes
Genomic DNA was extracted from wild-type Chinese Spring wheat, and from three
nullisomic-tetrasomic lines of chromosome 7 of Chinese Spring wheat, and from



Triticum tauschii (var strangulata, accession number CPI 100799), and used as a template for the amplification and nucleotide sequence analysis of wheat SSII genes.

RFLP analysis of *Bam*HI and *Eco*RI restricted DNA from each wheat or *T. Tauschii* line was carried out using the wSSIIp3 fragment as a probe. Three hybridising bands were obtained which could be assigned to chromosomes 7A, 7B and 7D, respectively (data not shown). This analysis indicates that there is a single copy of the wSSII gene in each genome in hexaploid wheat, consistent with the findings of Yamamori and Endo (1996) who located the SGP-A1, B1 and D1 proteins to the short arm of chromosome 7.

PCR analysis was used to assign each of the cDNA clones to the individual wheat genomes. A single 365 bp PCR fragment was obtained from nullisomic-tetrasomic genomic DNA of Chinese Spring when primers ssllc and sslld were used for the PCR amplification (Figure 5, right panel). This PCR product is obtained only from lines bearing the B genome. The fragment was cloned and sequenced and shown to be identical to a 365 bp region of the wSSIIB cDNA. An identical fragment is obtained by PCR amplification of the wSSIIB cDNA clone, but not by amplification of the wSSIIA or wSSIID clones, supporting the conclusion that the wSSIIB cDNA is the product of a gene located on chromosome 7 of the B genome of hexaploid wheat.

Two PCR products were also amplified from nullisomic-tetrasomic genomic DNA of Chinese Spring using the primers ssIIc and ssIIe (Figure 5, left panel). One PCR fragment, approximately 350 bp is only amplified when the A genome is present, and a second 322 bp product is only amplified when the D-genome is present. The 350 and 322 bp PCR products were also cloned and sequenced and shown to be identical to the wSSIIA and wSSIID cDNAs, respectively, supporting the conclusion that the wSSIIA and wSSIID cDNAs are the products of genes located on chromosomes 7A and 7D, respectively.

## Isolation of genomic wSSII clones

Screening of a genomic library from the D-genome donor of wheat, *T. tauschii*, was performed as described in Example 5, using the the PCR-derived DNA fragment wSSIIp2 as a hybridisation probe. Figure 6 shows an example of a plaque lift showing the positive-hybridising clone wSSII-8, a putative *T. tauschii* homologue of the wSSII gene.

Positive-hybridising plaques were digested using the restriction enzyme *BamHI*, separated on a 1% agarose gel, transferred to nitrocellulose membrane and hybridised to probe wSSIIp4 comprising nucleotides 1 to 367 of the wSSIIA cDNA clone, using the conditions described by Rahman *et al.* (1997). As shown in Figure 7,clone wSSII-8 also hybridises strongly to this probe, confirming its identity as a genomic wSSII gene. Furthermore, in light of the fact that the wSSIIp4 probe comprises the 5'-end of the cDNA clone, it is likely that genomic clone wSSII-8 comprises the promoter region of the wSSII gene.



15

## **EXAMPLE 13**

Cloning of specific cDNA regions of wheat starch synthase III using RT-PCR PCR primers were used to amplify sequences of starch synthase III from wheat endosperm cDNA. The design of PCR primers was based on the sequences of starch synthase III from potato and the *du1* starch synthase III gene of maize.

First-strand cDNAs were synthesised from 1  $\mu$ g of total RNA (derived from endosperm of the cultivar Rosella, 12 days after anthesis) as described by Maniatis *et al.* (1982), and then used as templates to amplify two specific cDNA regions, wSSIIIp1 and wSSIIIp2, of wheat starch synthase III by PCR.

The primers used to obtain the cDNA clone wSSIIIp1 were as follows:

Primer wSS3pa (5' GGAGGTCTTGGTGATGTTGT 3': SEQ ID NO: <400>29); and

Primer wSS3pb (5' CTTGACCAATCATGGCAATG 3': SEQ ID NO: <400>30).

The primers used to obtain the cDNA clone wSSIIIp2 were as follows:

Primer wSS3pc (5' CATTGCCATGATTGGTCAAG 3': SEQ ID NO: <400>31); and

Primer wSS3pd (5' ACCACCTGTCCGTTCCGTTGC 3': SEQ ID NO: <400>32).

20 The amplified clones wSSIIIp1 and wSSIIIp2 were used as probes to screen the third cDNA library and *T. tauschii* genomic DNA library as described in Example 4.

A further probe designated wSSIIIp3 was used for screening the third cDNA library, as described in Example 4. Probe wSSIIIp3 was amplified by PCR from a cDNA clone produced from the first screening using the following amplification primers:

Primer wSS3pe (5' GCACGGTCTATGAGAACAATGGC 3': SEQ ID NO: <400>33); and Primer wSS3pf (5' TCTGCATACCACCAATCGCCG 3': SEQ ID NO: <400>34).

The amplification reactions were performed using a FTS-1 or FTS4000 thermal sequencer (Corbett, Australia) for 1 cycle of 95°C for 2 minutes; 35 cycles of 95°C for

30 seconds, 60°C for 1 minutes, 72°C for 2 minutes and 1 cycle of 25°C for 1 minute.

Amplified sequences of the expected length were obtained, cloned and sequenced, and shown to contain DNA sequences highly homologous to the maize and potato SSIII genes. PCR fragments were subsequently used to probe a wheat cDNA library by DNA hybridisation and 8 positive clones were obtained, including one 3 kb cDNA. A region from the 5' end of this cDNA was amplified by PCR and used a probe for a second round of screening the cDNA library, obtaining 8 cDNA clones. Of these, one cDNA was demonstrated to be full length (wSSIII.B3, 5.36 kb insert). The sequence of the 5.36 kb wSSIII.B3 cDNA clone is given in SEQ ID NO:<400>7.

Sequencing of the 8 cDNA clones obtained from the second round screening of the wheat cDNA library revealed that there were at least 2 classes of cDNA encoding SSIII present, possibly being encoded by homeologous genes on different wheat genomes.

- 15 The sequence of a representative of this second class of cDNA clones, wSSIII.B1, is shown in SEQ ID NO:<400>9. The 3664 bp clone wSSIII.B1 is not full length, spanning only the region from nucleotides 1690 to 5363 of the homeologous clone wSSIII.B3, with an open reading frame between nucleotide positions 1 and 3180.
- 20 An open reading frame is found between the ATG translation start codon at position 29 and the stop codon at position 4921 of the cDNA clone wSSIII.B3 (SEQ ID NO:<400>7). The amino acid sequence deduced from this open reading frame is shown in SEQ ID NO:<400>8.
- 25 An alignment of the deduced amino acid sequences of SSIII from maize, potato and wheat is shown in Figure 8. There is 56.6% identity between the maize SSIII and wheat wSSIII.B3 sequences at the amino acid level.

The C-terminal domain of starch synthases comprise the catalytic domain, and a characteristic amino acid sequence motif KVGGLGDVVTSLSRAVQDLGHNVEV (SEQ



ID NO: <400> 35) in maize, or alternatively KVGGLGDVVTSLSRAIQDLGHTVEV (SEQ ID NO: <400>36) in wheat, marking the first conserved region in the C-terminal domain.

5 The amino acid identity between maize dull1 and wSSIII.B3 in the N-terminal region (i.e. amino acids 1 to 600 in Figure 8) is only 32.2%; whilst the amino acid identity in the central region (i.e. amino acids 601 to 1248 in Figure 8) is 68.4%; and in the C-terminal region (i.e. amino acids 1249 to 1631 in Figure 8) is 84.6%. Accordingly, the SSIII starch synthases are much more highly conserved between maize and wheat in the region comprising the catalytic domain of the proteins.

## **EXAMPLE 14**

# Isolation of genomic clones for SSIII

Screening of a genomic library from the D-genome donor of wheat, *T. tauschii*, 15 identified a number of clones which hybridised to the wSSIII PCR fragment. Positive plaques in the genomic library were selected as those hybridising with a probe that had been generated by PCR (amplifying between nucleotide positions 3620 to 3966) from the SSIII cDNA as template. The primer sequences used were as follows: wSS3pa (5' GGAGGTCTTGGTGATGTTGT 3': SEQ ID NO: <400>29); and 20 wSS3pb (5' CTTGACCAATCATGGCAATG 3': SEQ ID NO: <400>30).

Hybridisation was carried out in 25% formamide, 6 x SSC, 0.1% SDS at 42 °C for 16 hour, then washed three times with 2 x SSC containing 0.1% SDS at 65 °C, for 1 hour per wash. Figure 9 shows an example of a plaque lift showing positive and negative hybridisations for plaques containing the *T. tauschii* homologue of the wSSIII.B3 gene.

DNA was isolated from positive-hybridising λ clones using methods described by Maniatis *et al.* Briefly, DNA was digested using *Bam*HI or *BgI*I and sub-cloned in to the vector pJKKmfm. DNA sequencing was performed using the automated ABI system 30 with dye terminators as described by the manufacturers. DNA sequences were

analysed using the GCG suite of programs (Devereaux et al., 1984).

Nucleotide equences of the genomic SSIII clone from *T. tauschii* are provided gerein as 6 contiguous sequences designated fragments 1 to 6 (SEQ ID 5 NOs:<400>11 to <400>16, respectively). Table 3 defines the relative positions of these fragments with respect to the SSIII cDNA and describes the positions of exons. Figure 1 shows this information schematically.

10 EXAMPLE 15

# Analysis of wheat starch synthase III mRNA expression

Figure 10 shows the expression of wSSIII mRNA during endosperm development in two wheat varieties grown under defined environmental conditions. The expression of the gene is seen very early in endosperm development in both cultivars, 4 days after anthesis (Figure 10, panels a and b). Expression in the leaf of the variety Gabo is very weak (Figure 10, panel c, Lane L) whereas strong expression is seen in pre-anthesis florets (Figure 10, panel c, Lane P).

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TABLE 3

Summary of the Wheat Starch Synthase III Genomic Sequence

	Fragment in genomic DNA	Length	Features in SEQ ID NOs:<400>11 to <400>16	Corresponding region in cDNA sequence
	clone	(pb)		
2	Fragment 1	728	Translation start codon (nucleotides 287 to 289);	
	(SEQ ID NO:<400>11)		Exon 1.1 (nucleotides 260 to385).	Exon 1.1: nucleotides 1 to 126
<del></del>	Fragment 2	2446	Exon 2.1 ( nucleotides 1 to 1938);	Exon 2.1: nucleotides 1008 to 2948;
	(SEQ ID NO:<400>12)		Exon 2.2 (nucleotides 2197 to 2418).	Exon 2.2: nucleotides 2949 to 3171
-	Fragment 3	1032	Exon 3.1 (nucleotides 310 to 580)	Exon 3.1: nucleotides 3172 to 3440
10	(SEQ ID NO:<400>13)			
-	Fragment 4	892	Exon 4.1 (nucleotides 678 to 853)	Exon 4.1: nucleotides 3441 to 3616
	(SEQ ID NO:<400>14)			
	Fragment 5	871	Partial Exon 5.1 (nucleotides 1 to 29)	Exon 5.1: nucleotides 3908 to 3937 (partial)
	(SEQ ID NO:<400>15)		Exon 5.2 (nucleotides 293 to 463)	Exon 5.2: nucleotides 3938 to 4108
			Exon 5.3 (nucleotides 589 to 695)	Exon 5.3: nucleotides 4109 to 4215
15	Fragment 6	1583	Exon 6.1 (nucleotides 471 to 653);	Exon 6.1: nucleotides 4238 to 4420
	(SEQ ID NO:<400>16)		Exon 6.2 (nucleotides 770 to 902);	Exon 6.2: nucleotides 4421 to 4552
			Exon 6.3 (nucleotides 999 to 1110);	Exon 6.3: nucleotides 4553 to 4664
			Exon 6.4 (nucleotides 1201 to 1328);	Exon 6.4: nucleotides 4665 to 4793
			Partial Exon 6.5 (nucleotides 1408 to 1583);	Exon 6.5: nucleotides 4794 to 4966 (partial)
			Translation stop codon (nucleotides 1536 to 1538)	

#### **EXAMPLE 16**

# Amino acid sequence comparisons between wheat SSII and SSIII polypeptides

5 Amino acid sequence comparisons between wheat BSSS, SSI, SSII and SSIII polypeptides, as indicated in Figure 13, reveals eight highly-conserved domains. The amino acid sequences of these domains are represented in the wheat SSIII amino acid sequence by the following sequence motifs:

(a) Region1:

KVGGLGDVVTS;

10

(b) Region 2:

GHTVEVILPKY;

(c) Region 3:

HDWSSAPVAWLYKEHY;

(d) Region 4:

GILNGIDPDIWDPYTD;

(e) Region 5:

DVPIVGIITRLTAQKG;

(f) Region 5a:

NGQVVLLGSA;

15

(g) Region 6:

AGSDFIIVPSIFEPCGLTQLVAMRYGS; and

(h) Region 7:

TGGLVDTV.

As shown in Table 4 below, there is at least about 25% amino acid sequence identity, preferably at least about 30% amino acid sequence identity, more preferably at least about 35% amino acid sequence identity, more preferably at least about 40% amino acid sequence identity, more preferably at least about 45% amino acid sequence identity, more preferably at least about 50% amino acid sequence identity, more preferably at least about 55% amino acid sequence identity, more preferably at least about 60% amino acid sequence identity, more preferably at least about 65% amino acid sequence identity, more preferably at least about 75% amino acid sequence identity, more preferably at least about 80% amino acid sequence identity, more preferably at least about 85% amino acid sequence identity, more preferably at least about 85% amino acid sequence identity and even more preferably at least about 95% amino acid sequence identity between the



amino acid sequences of plant starch synthase enzymes, in particular wheat starch synthases.

TABLE 4

Identities between conserved motifs of plant starch synthases

	Conserved Region	Number of conserved	Number of conserved
1		residues between wheat	residues between
		starch synthases	wheat SSII and SSIII
			polypeptides
	Region1:		
į	KVGGLGDVVTS	6/11 residues	6/11 residues
	Region 2:		
10	GHTVEVILPKY	6/11 residues	6/11 residues
	Region 3:		
	HDWSSAPVAWLYKEHY	4/16 residues	5/16 residues
	Region 4:		
	GILNGIDPDIWDPYTD	7/16 residues	8/16 residues
15	Region 5:		
	DVPIVGIITRLTAQKG	8/16 residues	10/16 residues
	Region 5a:		
	NGQVVLLGSA	4/10 residues	4/10 residues
	Region 6:		
20	AGSDFIIVPSIFEPCGLT	15/27 residues	17/27 residues
	QLVAMRYGS		
	Region 7:		
	TGGLVDTV	5/9 residues	5/9 residues

25

The most conserved regions of the wheat SSII and SSIII polypeptides are a region of 6 or 7 identical amino acids in Region 1 (Table 4; Figure 13) and a region of 8 or

9 identical amino acids in Region 6 (Table 4; Figure 13). The lowest regions of identity are found in regions 3 and 5a.

5

#### **EXAMPLE 17**

#### Discussion

Early work on the Sgp-1 starch synthase proteins (Denyer *et al.*, 1995; Rahman *et al.*, 1995) was based on the localisation of these proteins in the wheat starch granule, and no definitive conclusion concerning their presence or absence in soluble extracts of the wheat endosperm was presented. We have now demonstrated that a monoclonal antibody against the Sgp-1 proteins cross reacts strongly with those starch synthase proteins having apparent molecular weights of 100-105 kDa in soluble extracts, however, the appearance of these proteins in soluble extracts is dependant on the developmental stage of the endosperm material. Whilst the proteins can be detected in the soluble phase in early to mid endosperm development, little or no soluble protein remains in late endosperm development (Figure 1). This observation accounts for the failure of Rahman *et al.* (1995) to detect the protein in soluble extracts in a previous report.

20 Based upon the localisation of the Sgp-1 starch synthase proteins in the wheat endosperm, the following nomenclature is suggested for wheat starch synthase enzymes: wGBSS for the 60 kDa granule bound starch synthase (Wx); wSSI for the 75 kDa starch synthase I (Sgp-3); wSSII for the 100 - 105 kDa proteins (Sgp-1); and wSSIII for a soluble high molecular starch synthase.

25

The present invention provides cDNA clones encoding the wSSII and wSSIII polypeptides and the corresponding genomic clones.

The wSSIII cDNA clone described herein is clearly related to the maize and potato 30 SSIII polypeptides.



Comparison of the amino acid sequences of all available starch synthases with the deduced amino acid sequences of the three wSSII cDNA clones of the present invention (i.e. wSSIIB, wSSIIA and wSSIID) was conducted using PILEUP analysis (Devereaux et al., 1984) and data are presented as a dendrogram (Figure 11). The sequence of the glycogen synthase of *E. coli* was also included. Based upon their amino acid similarities, four classes of plant starch synthases can be defined: GBSS, SSI, SSII and SSIII.

Based upon sequence identities and the function of the Sgp-1 proteins in wheat, the wSSIIB, wSSIIA and wSSID cDNA clones are members of the starch synthase II (SSII) group and are more similar in sequence to maize SSIIa than maize SSIIb. Table 5 shows that levels of identity at the amino acid level between the wSSII sequences, as determined using the BESTFIT programme in GCG (Devereaux *et al.*, 1984), and other class II starch synthases range from 70% identity with potato SSII to 85% identity with maize SSIIa. Both wSSIIB and wSSIID showed significantly higher homology to maize SSIIa than wSSIIA.

**TABLE 5** 

wSSII-A wSSII-B wssII-D 20 wssi-A 100% 95.9% 100% wSSII-B wssII-D 96.3% 100% 96.7% maize SSIIa 76.1% 85.2% 84.7% maize SSIIb 76.3% 76.7% 75.9% 25 pea SSII 72.0% 72.2% 71.8% potato SSII 70.9% 71.1% 70.3%

Whilst the evidence is compelling that the wSSIIA, wSSIIB and wSSIID cDNAs encode the Sgp-A1, Sgp-B1 and Sgp-D1 proteins of the wheat starch granule, molecular weights calculated from the deduced amino acid sequences of the clones

are considerably lower than estimates obtained from SDS-PAGE. The molecular weight of the precursor wSSIIA protein is 87,229 Da, and the mature protein 81,164 Da, yet the estimated molecular weight in our experience is 105 kDa. The assignment of the wSSIIA cDNA to the A-genome of wheat is demonstrated in 5 Figure 5, and the assignement of the 105 kDa protein to the A-genome in Denyer et al. (1995) and Yamamori and Endo (1996). Similarly, the molecular weight of the wSSIIB protein is 86,790 Da and the mature protein 80,759 Da, yet the molecular weight of the Sgp-B1 protein is estimated to be 100 kDa. No comparison can be made of the wSSIID sequences as a full length cDNA clone was not obtained. The 10 wSSIIA and wSSIIB amino acid sequences differ by just a single amino acid residue, yet there is an apparent difference of 5 kDa in molecular weight when estimated by SDS-PAGE. Several possibilities can be advanced to account for this apparent discrepancy in molecular weights. Firstly, the wSSII proteins may not migrate in SDS-PAGE in accordance with their molecular weight because they 15 retain some conformation under the denaturing conditions used. Secondly, the proteins may be glycosylated. It is also possible that the proteins may be noncovalently linked to starch through a high affinity starch binding site which survives denaturation and SDS-PAGE. Differences between the apparent molecular weights and those calculated from the deduced amino acid sequences will have to be 20 defined in establishing the relationship between the wSSII proteins and proteins encoded by the analogous SSII genes of other species.

The catalytic domain of the starch synthases is found at the C-terminal end of the protein (Gao *et al.*, 1998; Harn *et al.*, 1998). Harn *et al.* (1998) identified 7 conserved regions among SSIIa, SSIIb, SSI and GBSS sequences.

We have identified include an additional conserved region (designated region 5a in Figure 3 and Figure 12) comprising the amino acid sequence motif DVQLVMLGTG. Comparison of the wSSII sequences of the present invention with differing isoforms of starch synthases (GBSS, SS1, SSII and SSIII) identified a total of 8 regions of



the deduced amino acid sequences which were conserved amongst starch synthases from each class. Figure 12 shows an alignment of plant starch synthase sequences, in which the position of the first homologous region is used as the basis of the alignment. This first homologous region contains the consensus motif KXGG which is believed to be present in the ADPglucose binding site of starch and glycogen synthases (Furukawa *et al.*, 1990).

The conservation of eight peptide regions among the 4 classes of starch synthases is striking, in terms of their sequence homologies and their alignment. The major 10 differences in structure between the classes of genes are found in the length of the N-terminal region between the transit peptide and the first conserved region. At one extreme, the GBSS genes have a very short N-terminal arm, whereas the du1 starch synthase contains a very long N-terminal extension containing several distinct regions (Figure 12). The wSSII genes contain an N-terminal extension 15 which is longer than either GBSS,SSI, or SSIIb, and slightly longer than the maize SSIIa gene (Figure 12). Analysis of the wheat SSII genes shows that there is a motif, PVNGENK, which is repeated. The area surrounding the repeated PVNGENK motif is not homologous to maize SSIIa and the insertion of this region is responsible for the difference in length between the wheat SSII and maize SSIIa 20 genes. In pea and potato SSII polypeptides, a PPP motif (Figure 3; residues 251-253 and 287-289 respectively) has been suggested to mark the end of the Nterminal region and to facilitate the flexibility of an "N-terminal arm". This motif is not found in either the maize or wheat SSII sequences.

25 The generation of a wheat line combining null alleles at each of the three wSSII loci, wSSIIA, wSSIIB and wSSIID, has been reported recently by Yamamori (1998). In this triple null line, the large starch granules were reported to be mostly deformed and a novel starch with high blue value was observed when stained with iodine, indicating that wSSII is a key enzyme for the synthesis of starch in wheat. Further analysis of the starch derived from this triple null mutant is in progress.

Mutations in starch synthases are known in three other species. In pea, mutation in SSII gives rise to starch with altered granule morphology and an amylopectin which yields an oligosaccharide distribution with reduced chain length on debranching. compared to the wild type (Craig et al., 1998). A similar mutation in a gene 5 designated SSII is known in Chlamydomonas (the sta-3 mutation) and similar effects on granule morphology and amylopectin structure are observed (Fontaine et al., 1993). In maize, two mutations affecting starch synthases are known. First, the dull1mutation has been shown to be caused by a lesion within the du1 SSIII-type starch synthase gene (Gao et al., 1998). A second mutation, the sugary-2 mutation 10 yields a starch with reduced amylopectin chain lengths on debranching (this mutation co-segregates with the SSIIa locus (Harn et al., 1998) although direct evidence that the sugary-2 mutation is caused by a lesion in the SSIIa gene is lacking). In the SSII mutants of each of these species, amylose biosynthesis capacity is retained, suggesting different roles in amylose and amylopectin 15 synthesis for the GBSS and SSII genes. Given the conservation in overall organisation of the GBSS and SSII genes (see Figures 11 and 12), when an alignment is made based on the KTGGL motif of the first conserved region, this focuses attention on the role(s) of the N-terminal region in defining substrate specificity and the localisation of the proteins as the N-terminal region is the major 20 area of divergence between the 4 classes of starch synthases. However, it is premature to exclude the influence of more subtle mutations in central and Cterminal regions of the gene.

The cloning of the wSSII and wSSIII cDNAs and genomic clones described herein provides useful tools for the further study of the roles of the starch synthases in wheat. Firstly, they provide a source of markers which can be used to recover and combine null or divergent alleles. Secondly, genetic manipulation of wheat by gene suppression or over-expression can be carried out, and the genes may be used for overexpression in other species. The promoter regions of these genes are also useful in regulating the expression of starch synthase genes and other

P:\OPER\MRO\PI-WSS.PRV - 29/4

heterologous genes in the developing wheat endosperm and in pre-anthesis florets of wheat.

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## SEQUENCE LISTING

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gtg d	cag	ctg	gtc	atg	ctg	ggc	acc	aaa ,	cgc	cac	gac	ctg	gag	ggc	atg	2146



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gcc atg gcc tac Ala Met Ala Tyr				
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tgg acg ttc gac Trp Thr Phe Asp 740	cgc gca gag Arg Ala Glu	gcg cag aag c Ala Gln Lys Lo 745	tg atc gag gcg eu Ile Glu Ala 750	ctc ggg 2434 Leu Gly
cac tgc ctc cgc His Cys Leu Arg 755				
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Gly Gln Asp Met Glu Val Asn Tyr Phe His Ala Tyr Ile Asp Gly Val

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Leu Arg Asp Thr Val Pro Pro Phe Asp Pro Phe Asn His Ser Gly Leu Gly Trp Thr Phe Asp Arg Ala Glu Ala Gln Lys Leu Ile Glu Ala Leu Gly His Cys Leu Arg Thr Tyr Arg Asp Tyr Lys Glu Ser Trp Arg Gly Leu Gln Glu Arg Gly Met Ser Gln Asp Phe Ser Trp Glu His Ala Ala Lys Leu Tyr Glu Asp Val Leu Val Lys Ala Lys Tyr Gln Trp 790 <210> 3 <211> 2800 <212> DNA <213> Triticum aestivum <220> <221> CDS <222> (89)..(2488) <400> 3 gctgccacca cctccgcctg cgccgcgctc tgggcggagg accaacccgc gcatcgtacc 60 ategeeegee eegateeegg eegeegee atg teg teg geg gte geg tee gee 112 Met Ser Ser Ala Val Ala Ser Ala gcg tcc ttc ctc gcg ctc gcc tcc gcc tcc ccc ggg aga tca cgc agg 160 Ala Ser Phe Leu Ala Leu Ala Ser Ala Ser Pro Gly Arg Ser Arg Arg 10 15 208 egg geg agg gtg age geg eeg eec eec eac gee ggg gee gge agg etg Arg Ala Arg Val Ser Ala Pro Pro Pro His Ala Gly Ala Gly Arg Leu cac tgg ccg ccg tgg ccg ccg cag cgc acg gct cgc gac gga ggt gtg 256 His Trp Pro Pro Trp Pro Pro Gln Arg Thr Ala Arg Asp Gly Gly Val 50 gcc gcg cgc gcc gcc ggg aag aag gac gcg agg gtc gac gac gcc 304 Ala Ala Arg Ala Ala Gly Lys Lys Asp Ala Arg Val Asp Asp Asp Ala gcg tcc gcg agg cag ccc cgc gca cgc cgc ggt ggc gcc gcc acc aag 352 Ala Ser Ala Arg Gln Pro Arg Ala Arg Arg Gly Gly Ala Ala Thr Lys 75 80 gtc gcg gag cgg agg gat ccc gtc aag acg ctc gat cgc gac gcc gcg 400 Val Ala Glu Arg Arg Asp Pro Val Lys Thr Leu Asp Arg Asp Ala Ala 448 Glu Gly Gly Ala Pro Ala Pro Pro Ala Pro Arg Gln Asp Ala Ala Arg cca ccg agt atg aac ggc acg ccg gtg aac ggt gag aac aaa tct acc 496 Pro Pro Ser Met Asn Gly Thr Pro Val Asn Gly Glu Asn Lys Ser Thr 125 130 135

gge gge gge geg ace aaa gae age ggg etg eec gea eec gea ege

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Lys	Thr	Leu	Asp 100	Arg	Asp	Ala	Ala	Glu 105	Gly	Gly	Ala	Pro	Ala 110	Pro	Pro
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Arg	Val	Pro	Val	Asn 165	Gly	Glu	Asn	Lys	Ala 170	Asn	Val	Ala	Ser	Pro 175	Pro
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Ile	Ser	Asp 195	Lys	Ala	Pro	Glu	Ser 200	Val	Val	Pro	Ala	Glu 205	Lys	Pro	Pro
Pro	Ser 210	Ser	Gly	Ser	Asn	Phe 215	Val	Val	Ser	Ala	Ser 220	Ala	Pro	Arg	Leu
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Val	Glu	Glu	Ala	Pro 245	Asn	Pro	Lys	Ala	Leu 250	Ser	Pro	Pro	Ala	Ala 255	Pro
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Glu	Pro	Val 275	Glu	Ala	Lys	Asp	Asp 280	Gly	Trp	Ala	Val	Ala 285	Asp	Asp	Ala
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Asp	Tyr	Glu 355	Glu	Ala	Tyr	Asp	Val 360	Gly	Val	Arg	Lys	Tyr 365	Tyr	Lys	Ala
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170

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576

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435 440 445 ggg cgc cac gac ctg gag agc atg ctg cag cac ttc gag cgg gag cac 1392 Gly Arg His Asp Leu Glu Ser Met Leu Gln His Phe Glu Arg Glu His cac gac aag gtg cgc ggg tgg gtg ggg ttc tcc gtg cgc ctg gcg cac 1440 His Asp Lys Val Arg Gly Trp Val Gly Phe Ser Val Arg Leu Ala His 470 475 cgg atc acg gcg ggg gcg gac gcg ctc ctc atg ccc tcc cgg ttc gtg 1488 Arg Ile Thr Ala Gly Ala Asp Ala Leu Leu Met Pro Ser Arg Phe Val 485 490 495 ccg tgc ggg ctg aac cag ctc tac gcc atg gcc tac ggc acc gtc ccc 1536 Pro Cys Gly Leu Asn Gln Leu Tyr Ala Met Ala Tyr Gly Thr Val Pro 500 505 gtc gtg cac gcc gtc ggc ggc ctc agg gac acc gtg ccg ccg ttc gac Val Val His Ala Val Gly Gly Leu Arg Asp Thr Val Pro Pro Phe Asp 520 ccc ttc aac cac tcc ggg ctc ggg tgg acg ttc gac cgc gcc gag gcg 1632 Pro Phe Asn His Ser Gly Leu Gly Trp Thr Phe Asp Arg Ala Glu Ala cac aag ctg atc gag gcg ctc ggg cac tgc ctc cgc acc tac cga gac 1680 His Lys Leu Ile Glu Ala Leu Gly His Cys Leu Arg Thr Tyr Arg Asp ttc aag gag agc tgg agg gcc ctc cag gag cgc ggc atg tcg cag gac 1728 Phe Lys Glu Ser Trp Arg Ala Leu Gln Glu Arg Gly Met Ser Gln Asp 565 tte age tgg gag cae gee gee aag ete tae gag gae gte ete gte aag 1776 Phe Ser Trp Glu His Ala Ala Lys Leu Tyr Glu Asp Val Leu Val Lys 580 gcc aag tac cag tgg tga acgctagctg ctagccgctc cagccccgca 1824 Ala Lys Tyr Gln Trp tgcgtgcatg acaggatgga actgcattgc gcacgcagga aagtgccatg gagcgccggc 1884 atccgcgaag tacagtgaca tgaggtgtgt gtggttgaga cgctgattcc aatccggccc 1944 gtagcagagt agagcggagg tatatgggaa tcttaacttg gtattgtaat ttgttatgtt 2004 gtgtgcatta ttacaatgtt gttacttatt cttgttaagt cggaggccaa gggcgaaagc 2064 2107 <210> 6 <211> 597 <212> PRT <213> Triticum aestivum Pro Ala Glu Lys Thr Pro Pro Ser Ser Gly Ser Asn Phe Glu Ser Ser Ala Ser Ala Pro Gly Ser Asp Thr Val Ser Asp Val Glu Gln Glu Leu



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Pro	Cys	Gly	Leu 500	Asn	Gln	Leu	Tyr	Ala 505	Met	Ala	Tyr	Gly	Thr 510	Val	Pro	
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		_		_					_	_			gct Ala	_		2308
	_		-	-	_				_				tgg Trp			2356
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Lys Lys Glu Val Asp Ala Ala Asp Lys Ala Arg Val Lys Glu Asp Ala

Phe Glu Leu Asp Xaa Ala Ser Thr Thr Leu Arg Ser Val Ile Val Asp Val Met Asp His Xaa Trp Asp Cys Gln Glu Thr Leu Arg Ser Val Ile Val Asp Val Met Asp His Asn Gly Thr Val Gln Glu Thr Leu Arg Ser Val Ile Val Asp Val Met Asp Asp Ala Ala Asp Lys Ala Arg Val Glu Glu Asp Val Phe Glu Leu Asp Leu Ser Gly Asn Ile Ser Ser Ser Ala 250 Thr Thr Val Glu Leu Asp Ala Val Asp Glu Val Gly Pro Val Gln Asp Lys Phe Glu Ala Thr Ser Ser Gly Asn Val Ser Asn Ser Ala Thr Val 280 Arg Glu Val Asp Ala Ser Asp Glu Ala Gly Asn Asp Gln Gly Ile Phe Arg Ala Asp Leu Ser Gly Asn Val Phe Ser Ser Ser Thr Thr Val Glu Val Gly Ala Val Asp Glu Ala Gly Ser Ile Lys Asp Arg Phe Glu Thr Asp Ser Ser Gly Asn Val Ser Thr Ser Ala Pro Met Trp Asp Ala Ile 345 Asp Glu Thr Val Ala Asp Gln Asp Thr Phe Glu Ala Asp Leu Ser Gly 360 Asn Ala Ser Ser Cys Ala Thr Tyr Arg Glu Val Asp Asp Val Val Asp Glu Thr Arg Ser Glu Glu Glu Thr Phe Ala Met Asp Leu Phe Ala Ser 390 395 Glu Ser Gly His Glu Lys His Met Ala Val Asp Tyr Val Gly Glu Ala Thr Asp Glu Glu Glu Thr Tyr Gln Gln Tyr Pro Val Pro Ser Ser 425 Phe Ser Met Trp Asp Lys Ala Ile Ala Lys Thr Gly Val Ser Leu Asn Pro Glu Leu Arg Leu Val Arg Val Glu Glu Gln Gly Lys Val Asn Phe Ser Asp Lys Lys Asp Leu Ser Ile Asp Asp Leu Pro Gly Gln Asn Gln 470 Ser Ile Ile Gly Ser Tyr Lys Gln Asp Lys Ser Ile Ala Asp Val Ala Gly Pro Thr Gln Ser Ile Phe Gly Ser Ser Lys Gln His Arg Ser Ile 505 Val Ala Phe Pro Lys Gln Asn Gln Ser Ile Val Ser Val Thr Glu Gln



Lys Gln Ser Ile Val Gly Phe Arg Ser Gln Asp Leu Ser Ala Val Ser 535 Leu Pro Lys Gln Asn Val Pro Ile Val Gly Tyr Val Glu Arg Gly Ser 550 Asn Xaa Lys Gln Val Pro Val Val Asp Arg Gln Asp Ala Leu Tyr Val Asn Gly Leu Glu Ala Lys Glu Gly Asp His Thr Ser Glu Lys Thr Asp Glu Asp Ala Leu His Val Lys Phe Asn Val Asp Asn Val Leu Arg Lys His Gln Ala Asp Arg Thr Gln Ala Val Glu Lys Lys Thr Trp Lys Lys Val Asp Glu Glu His Leu Tyr Met Thr Glu His Gln Lys Arg Ala Ala Glu Gly Gln Met Val Val Asn Glu Asp Glu Leu Ser Ile Thr Glu Ile 645 650 Gly Met Gly Arg Gly Asp Lys Ile Gln His Val Leu Ser Glu Glu Glu 665 Leu Ser Trp Ser Glu Asp Glu Val Gln Leu Ile Glu Asp Asp Gly Gln 680 Tyr Glu Val Asp Glu Thr Ser Val Ser Val Asn Val Glu Gln Asp Ile Gln Gly Ser Pro Gln Asp Val Val Asp Pro Gln Ala Leu Lys Val Met Leu Gln Glu Leu Ala Glu Lys Asn Tyr Ser Met Arg Asn Lys Leu Phe Val Phe Pro Glu Val Val Lys Ala Asp Ser Val Ile Asp Leu Tyr Leu Asn Arg Asp Leu Thr Ala Leu Ala Asn Glu Pro Asp Val Val Ile Lys Gly Ala Phe Asn Gly Trp Lys Trp Arg Leu Phe Thr Glu Arg Leu His Lys Ser Asp Leu Gly Gly Val Trp Trp Ser Cys Lys Leu Tyr Ile Pro Lys Glu Ala Tyr Arg Leu Asp Phe Val Phe Phe Asn Gly Arg Thr Val Tyr Glu Asn Asn Gly Asn Asn Asp Phe Cys Ile Gly Ile Glu Gly Thr 825 Met Asn Glu Asp Leu Phe Glu Asp Phe Leu Val Lys Glu Lys Gln Arg 840 Glu Leu Glu Lys Leu Ala Met Glu Glu Ala Glu Arg Arg Thr Gln Thr 855 Glu Glu Gln Arg Arg Lys Glu Ala Arg Ala Ala Asp Glu Ala Val

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Glu	Ala	Ser 915		Asp	Thr	Arg	Gly 920	Asp	Thr	Ile	Arg	Leu 925	Tyr	Tyr	Asn
Arg	Asn 930	Ser	Arg	Pro	Leu	Ala 935	His	Ser	Thr	Glu	Ile 940	Trp	Met	His	Gly
Gly 945	Tyr	Asn	Asn	Trp	Thr 950	Asp	Gly	Leu	Ser	11e 955		Glu	Ser	Phe	Val 960
Lys	Суѕ	Asn	Asp	Lys 965	Asp	Gly	Asp	Trp	Trp 970	_	Ala	Asp	Val	Ile 975	Pro
Pro	Glu	Lys	Ala 980	Leu	Val	Leu	Asp	Trp 985	Val	Phe	Ala	Asp	Gly 990	Pro	Ala
Gly	Asn	Ala 995	Arg	Asn	Tyr	_	Asn 1000	Asn	Ala	Arg		Asp 1005	Phe	His	Ala
	Leu .010	Pro	Asn	Asn		Val 1015	Thr	Glu	Glu		Phe 1020	Trp	Ala	Gln	Glu
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Glu	Thr	Met	_	Arg 1045	Lys	Ala	Glu	_	Ser 1050	Ala	Asn	Ile	_	Ala 1055	Glu
Met	Lys		Lys 1060	Thr	Met	Arg	-	Phe 1065	Leu	Leu	Ser		Lys 1070	His	Ile
Val		Thr 1075	Arg	Thr	Xaa		Lys 1080	Tyr	Val	Pro		Thr 1085	Thr	Val	Asp
	Leu .090	Tyr	Asn	Pro		Asn 1095	Thr	Val	Leu		Gly 1100	Lys	Ser	Glu	Gly
Trp 105	Phe	Arg	Cys	Ser	Phe 1110	Asn	Leu	Trp		His 1115	Ser	Ser	Gly		Leu 1120
Pro	Pro	Gln		Met 1125	Val	Lys	Ser		Asp L130	Gly	Pro	Leu		Lys 1135	Ala
Thr	Val		Val .140	Pro	Pro	Asp		Tyr .145	Met	Met	Asp		Val 1150	Phe	Ser
∃lu		Glu 155	Glu	Asp	Gly		Tyr .160	Asp	Asn	Arg		Gly 165	Met	Asp	Tyr
	Ile 170	Pro	Val	Ser		Ser .175	Ile	Glu	Thr		Asn L180	Tyr	Met	Arg	Ile
le 185	His	Ile	Ala	Val 1	Glu .190	Met	Ala	Pro		Ala L195	Lys	Val	Gly		Leu 1200
Sly	Asp	Val		Thr 205	Ser	Leu	Ser		Ala 210	Ile	Gln	Asp		Gly	His



- Thr Val Glu Val Ile Leu Pro Lys Tyr Asp Cys Leu Asn Gln Ser Ser 1220 1225 1230
- Val Lys Asp Leu His Leu Tyr Gln Ser Phe Ser Trp Gly Gly Thr Glu 1235 1240 1245
- Ile Lys Val Trp Val Gly Arg Val Glu Asp Leu Thr Val Tyr Phe Leu 1250 1255 1260
- Glu Pro Gln Asn Gly Met Phe Gly Val Gly Cys Val Tyr Gly Arg Asn 265 1270 1275 1280
- Asp Asp Arg Arg Phe Gly Phe Phe Cys His Ser Ala Leu Glu Phe Ile 1285 1290 1295
- Leu Gln Asn Glu Phe Ser Pro His Ile Ile His Cys His Asp Trp Ser 1300 1305 1310
- Ser Ala Pro Val Ala Trp Leu Tyr Lys Glu His Tyr Ser Gln Ser Arg 1315 1320 1325
- Met Ala Ser Thr Arg Val Val Phe Thr Ile His Asn Leu Glu Phe Gly 1330 1340
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- Val Ser Pro Thr Tyr Ser Arg Asp Val Ala Gly His Gly Ala Ile Ala 1365 1370 1375
- Pro His Arg Glu Lys Phe Tyr Gly Ile Leu Asn Gly Ile Asp Pro Asp 1380 1385 1390
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- Glu Asn Val Val Glu Gly Lys Arg Ala Lys Arg Ala Leu Gln Gln 1410 1415 1420
- Lys Phe Gly Leu Gln Gln Thr Asp Val Pro Ile Val Gly Ile Ile Thr 425 1430 1435 1440
- Arg Leu Thr Ala Gln Lys Gly Ile His Leu Ile Lys His Ala Ile His 1445 1450 1455
- Arg Thr Leu Glu Ser Asn Gly His Val Val Leu Leu Gly Ser Ala Pro 1460 1465 1470
- Asp His Arg Ile Gln Gly Asp Phe Cys Arg Leu Ala Asp Ala Leu His 1475 1480 1485
- Gly Val Tyr His Gly Arg Val Lys Leu Val Leu Thr Tyr Asp Glu Pro 1490 1495 1500
- Leu Ser His Leu Ile Tyr Ala Gly Ser Asp Phe Ile Ile Val Pro Ser 505 1510 1515 1520
- Ile Phe Glu Pro Cys Gly Leu Thr Gln Leu Val Ala Met Arg Tyr Gly
  1525 1530 1535
- Ser Ile Pro Ile Val Arg Lys Thr Gly Gly Leu His Asp Thr Val Phe 1540 1545 1550
- Asp Val Asp Asn Asp Lys Asp Arg Ala Arg Ser Leu Gly Leu Glu Pro 1555 1560 1565

Asn Gly Phe Ser Phe Asp Gly Ala Asp Ser Asn Gly Val Asp Tyr Ala
1570
1580

Leu Asn Arg Ala Ile Gly Ala Trp Phe Asp Ala Arg Asp Trp Phe His
585
1590
1595
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agg aac aag ctg ttt gtt ttt cca gag gta gtg aaa gct gat tca gtt

528



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					gaa Glu											912
					agg Arg 310											960
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	Glu				aag Lys 390											1200
					cct Pro											1248
_	_			_	GJÀ aaa		_				-				_	1296

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Lys Thr Trp Lys Lys Val Asp Glu Glu His Leu Tyr Met Thr Glu His 55 60



Gln Lys Arg Ala Ala Glu Gly Gln Met Val Val Asn Glu Asp Glu Leu Ser Ile Thr Glu Ile Gly Met Gly Arg Gly Asp Lys Ile Gln His Val Leu Ser Glu Glu Glu Leu Ser Trp Ser Glu Asp Glu Val Gln Leu Ile Glu Asp Asp Gly Gln Tyr Glu Val Asp Glu Thr Ser Val Ser Val Asn Val Glu Gln Asp Ile Gln Gly Ser Pro Gln Asp Val Val Asp Pro Gln Ala Leu Lys Val Met Leu Gln Glu Leu Ala Glu Lys Asn Tyr Ser Met Arg Asn Lys Leu Phe Val Phe Pro Glu Val Val Lys Ala Asp Ser Val Ile Asp Leu Tyr Leu Asn Arg Asp Leu Thr Ala Leu Ala Asn Glu Pro 185 Asp Val Val Ile Lys Gly Ala Phe Asn Gly Trp Lys Trp Arg Leu Phe Thr Glu Arg Leu His Lys Ser Asp Leu Gly Gly Val Trp Trp Ser Cys Lys Leu Tyr Ile Pro Lys Glu Ala Tyr Arg Leu Asp Phe Val Phe Phe Asn Gly Arg Thr Val Tyr Glu Asn Asn Gly Asn Asn Asp Phe Cys Ile Gly Ile Glu Gly Thr Met Asn Glu Asp Leu Phe Glu Asp Phe Leu Val Lys Glu Lys Gln Arg Glu Leu Glu Lys Leu Ala Met Glu Glu Ala Glu Arg Arg Thr Gln Thr Glu Glu Gln Arg Arg Arg Lys Glu Ala Arg Ala Ala Asp Glu Ala Val Arg Ala Gln Ala Lys Ala Glu Ile Glu Ile Lys 310 Lys Lys Leu Gln Ser Met Leu Ser Leu Ala Arg Thr Cys Val Asp Asn Leu Trp Tyr Ile Glu Ala Ser Thr Asp Thr Arg Gly Asp Thr Ile 345 Arg Leu Tyr Tyr Asn Arg Asn Ser Arg Pro Leu Ala His Ser Thr Glu Ile Trp Met His Gly Gly Tyr Asn Asn Trp Ser Asp Gly Leu Ser Ile Val Glu Ser Phe Val Lys Cys Asn Asp Lys Asp Gly Asp Trp Trp Tyr Ala Asp Val Ile Pro Pro Glu Lys Ala Leu Val Leu Asp Trp Val Phe 405 410

Ala Asp Gly Pro Ala Gly Asn Ala Arg Asn Tyr Asp Asn Asn Ala Arg Gln Asp Phe His Ala Ile Leu Pro Asn Asn Asn Val Thr Glu Glu Gly 440 Phe Trp Ala Gln Glu Gln Asn Ile Tyr Thr Arg Leu Leu Gln Glu 455 Arg Arg Glu Lys Glu Glu Thr Met Lys Arg Lys Ala Glu Arg Ser Ala Asn Ile Lys Ala Glu Met Lys Ala Lys Thr Met Arg Arg Phe Leu Leu 490 485 Ser Gln Lys His Ile Val Tyr Thr Arg Thr Xaa Leu Lys Tyr Val Pro Gly Thr Thr Val Asp Val Leu Tyr Asn Pro Ser Asn Thr Val Leu Asn Gly Lys Ser Glu Gly Trp Phe Arg Cys Ser Phe Asn Leu Trp Met His Ser Ser Gly Ala Leu Pro Pro Gln Lys Met Val Lys Ser Gly Asp Gly 550 Pro Leu Leu Lys Ala Thr Val Asp Val Pro Pro Asp Ala Tyr Met Met 570 Asp Phe Val Phe Ser Glu Trp Glu Glu Asp Gly Ile Tyr Asp Asn Arg Asn Gly Met Asp Tyr His Ile Pro Val Ser Asp Ser Ile Glu Thr Glu 600 Asn Tyr Met Arg Ile Ile His Ile Ala Val Glu Met Ala Pro Val Ala 615 Lys Val Gly Gly Leu Gly Asp Val Val Thr Ser Leu Ser Arg Ala Ile Gln Asp Leu Gly His Thr Val Glu Val Ile Leu Pro Lys Tyr Asp Cys 650 Leu Asn Gln Ser Ser Val Lys Asp Leu His Leu Tyr Gln Ser Phe Ser 665 Trp Gly Gly Thr Glu Ile Lys Val Trp Val Gly Arg Val Glu Asp Leu 680 Thr Val Tyr Phe Leu Glu Pro Gln Asn Gly Met Phe Gly Val Gly Cys Val Tyr Gly Arg Asn Asp Asp Arg Arg Phe Gly Phe Phe Cys His Ser Ala Leu Glu Phe Ile Leu Gln Asn Glu Phe Ser Pro His Ile Ile His 725 730 Cys His Asp Trp Ser Ser Ala Pro Val Ala Trp Leu Tyr Lys Glu His Tyr Ser Gln Ser Arg Met Ala Ser Thr Arg Val Val Phe Thr Ile His P:\OPER\MRO\PI-WSS.PRV - 29/4

765 755 760 Asn Leu Glu Phe Gly Ala His Tyr Ile Gly Lys Ala Met Thr Tyr Cys 775 Asp Lys Ala Thr Thr Val Ser Pro Thr Tyr Ser Arg Asp Val Ala Gly His Gly Ala Ile Ala Pro His Arg Glu Lys Phe Tyr Gly Ile Leu Asn Gly Ile Asp Pro Asp Ile Trp Asp Pro Tyr Thr Asp Asn Phe Ile Pro 825 Val Pro Tyr Thr Cys Glu Asn Val Val Glu Gly Lys Arg Ala Ala Lys Arg Ala Leu Gln Gln Lys Phe Gly Leu Gln Gln Thr Asp Val Pro Ile Val Gly Ile Ile Thr Arg Leu Thr Ala Gln Lys Gly Ile His Leu Ile 870 Lys His Ala Ile His Arg Thr Leu Glu Ser Asn Gly Gln Val Val Leu Leu Gly Ser Ala Pro Asp His Arg Ile Gln Gly Asp Phe Cys Arg Leu Ala Asp Ala Leu His Gly Val Tyr His Gly Arg Val Lys Leu Val Leu Thr Tyr Asp Glu Pro Leu Ser His Leu Ile Tyr Ala Gly Ser Asp Phe Ile Ile Val Pro Ser Ile Phe Glu Pro Cys Gly Leu Thr Gln Leu Val Ala Met Arg Tyr Gly Ser Ile Pro Ile Val Arg Lys Thr Gly Gly Leu 970 Tyr Asp Thr Val Phe Asp Val Asp Asn Asp Lys Asp Arg Ala Arg Ser 985. Leu Gly Leu Glu Pro Asn Gly Phe Ser Phe Asp Gly Ala Asp Ser Asn Gly Val Asp Tyr Ala Leu Asn Arg Ala Ile Gly Ala Trp Phe Asp Ala 1015 1020 Arg Asp Trp Phe His Ser Leu Cys Lys Arg Val Met Glu Gln Asp Trp 1030 Ser Trp Asn Arg Pro Ala Leu Asp Tyr Ile Glu Leu Tyr His Ala Ala 1050 Arg Lys Phe

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aacatcttta catgactgaa catcagatag gtgctgccga aggacagatg gtagttaacg 960



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<sup>&</sup>lt;211> 1032

<sup>&</sup>lt;212> DNA

<sup>&</sup>lt;213> Triticum sp.

<sup>&</sup>lt;400> 13

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accacagtgg atgtgctata caatccctct aacacagtgc taaatggaaa gccggaggtt 480
tggtttagat gctctttaa cctttggatg catccaagtg gagcattgcc accccagaag 540
atggtgaaat caggggatgg gccgctctta aaagccacag gtttattgcg ttattacatc 600
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<211> 892

<212> DNA

<213> Triticum sp.

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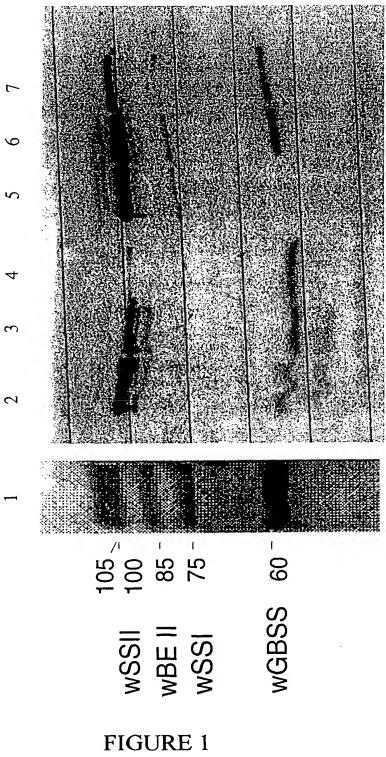
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<211> 25

Gln Asp Leu Gly His Thr Val Glu Val



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/SSIID wssiia	~~~~~~~	~~~~~~~~	~~~~~~~~	~~~~~~~	~~~~~~~
WSSIIA	~~~~~~	~~~~~~~	~~~~~~~~	~~~~~~~	~~~~~~~
	51				100
wSSIIB		CCCACTGCCG	CGCTACTCCC	CACTCCCACT	
wssIID	~~~~~~~	~~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~
wssiia	~~~~~~~~	~~~~~~~~	~~~~~~~~	~~~~~GCT	GCCACCACCT
	101				150
wssiib	CCGCcTGCGC	CGCGCTCTGG	GCGGACCAAC	CCGCGCATCG	TATCACGATC
wssiid	~~~~~~~~	~~~~~~~~	~~~~~~~~		
wssiia	CCGCCTGCGC	CGCGCTCTGG	GCGGAGGACC	AACCCGCGCA	TCGTACCATC
	151				200
wssiib		ATCCCGGCCG	CCCCCATGTC	стессессте	
wssiid	~~~~~~	~~~~~~~	~~~~~~~	~~~~~~	~~~~~~~
wssiia	GCCCGCCCG	ATCCCGGCCG	CCGCCATGTC	GTCGGCGGTC	GCGTCCGCCG
	201				250
wssiib	CGTCCTTCCT	CGCGCTCGCG	TCCGCCTCCC	CCGGGAGATC	ACGGAGGAGG
wssIID	~~~~~~~	~~~~~~~~	~~~~~~~~	~~~~~~~~	~~~~~~~~
wssiia	CGTCCTTCCT	CGCGCTCGCC	TCCGCCTCCC	CCGGGAGATC	ACGCAGGCGG
					2.00
TTD	251	aaaaamaaaa	20000202020		300
wssIIB wssIID	ACGAGGG I GA	GCGCGTCGCC	ACCCCACACC	GGGGCTGGCA	GGTTGCACTG
WSSIIA	GCGAGGGTGA	GCGCGCCGCC	ACCCCACGCC	GGGGCCGGCA	GGCTGCACTG
"55111	00010001011		Accentegee		00010011010
	301				350
wssiib	GCCGCCGTCG	CCGCCGCAGC	GCACGGCTCG	CGACGGAGCG	GTGGCCGCGC
wssiid	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~~	~~~~~~~
WSSIIA	GCCGCCGTGG	CCGCCGCAGC	${\tt GCACGGCTCG}$	CGACGGAGGT	GTGGCCGCGC
	351	~~-			400
wssIIB wssIID	GCGCCGCCGG	GAAGAAGGAC	GCGGGGAT	. CGACGACGC	CGCGCCCGCG
WSSIIA	GCGCCGCCGG	GAAGAAGGAC	CCCACCCTCC	ACCACCACCC	CGCGTCCGCG
MDDIIM	GCGCCGCCGG	GAAGAAGGAC	GCGAGGGICG	ACGACGACGC	CGCGTCCGCG
	401				450
wSSIIB	AGGCAGCCCC	GCGCACTCCG	CGGTGGCGCC	GCCACCAAGG	
wSSIID	~~~~~~~	~~~~~~~	~~~~~~	~~~~~~~	~~~~~~
wssiia	AGGCAGCCCC	GCGCACGCCG	CGGTGGCGCC	GcCACCAAGG	TCGCGGAGCG
	451				500
wssiib	GAGGGATCCC				
wssiid		CTCAACACC			
wssiia	GAGGGATCCC	GICAAGACGC	TCGATCGCGA	CGCCGCGGAA	GGTGGCGCGC
	501				550
wSSIIB	CGTCCCCGCC	GGCACCGAGG	CAGGAGGACG	CCCGTCTGCC	
wssIID	~~~~~~~				
WSSIIA	CGGCACCGCC	GGCACCGAGG	CAGGACGCCG	CCCGTCCaCC	GAGTATGAAC

### FIGURE 2-1

	551	ma			600
WSSIIB	GGCATGCCGG	TGAACGGTGA	AAACAAATCT	ACCGGCGGCG	GCGGCGCGAC
wssiid wssiia	000700000	man nacaman	~~~~~~~~	ACCGGCGGCG	
WSSIIA	GGCACGCCGG	IGAACGGIGA	GAACAAAICI	ACCGGCGGCG	GCGGCGCGAC
	601				650
wssIIB		GGGCTGCCCG	CACCCGCACG	CGCGCCCCAG	
wssIID				~~~~~~~	
wssiia				CGCGCCCCAT	
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	651				700
wSSIIB	AGAACAGAGT	ACCGGTGAAT	GGTGAAAACA	AAGCTAACGT	CGCCTCGCCG
wssIID	~~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~
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	701				750
wssiib	CCGACGAGCA	TAGCCGAGGT	CGCGGCTCCG	GATCCCGCAG	CTACCATTTC
wssIID	~~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~
wssiia	CCGACGAGCA	TAGCCGAGGT	CGTGGCTCCG	GATTCCGCAG	CTACCATTTC
	_				
	751				800
wssiib				CCCAGCCGAG	
wssiid				~CCAGCTGAG	
wssiia	CATCAGTGAC	AAGGCGCCGG	AGTCCGTTGT	CCCAGCCGAG	AAGCCGCCGC
	801				850
wSSIIB		Ctcaaamtrc	ataCcCtCaa	cttctGctCc	
wssiid				CCTCTGCTCC	
WSSIIA				CTTCTGCTCC	
	851				900
wssiib	actgtCaGCG	acGtGGaact	TgaActGAAg	aAGGGtgCgg	tCattgTcaA
wssIID	ACTGTCAGCG	ACGTGGAACA	AGAACTGAAG	AAGGGTGCGG	TCGTTGTCGA
wssiia	ATTGACAGCG	ATGTTGAACC	TGAACTGAAG	AAGGGTGCGG	TCATCGTCGA
	901				950
wSSIIB	-			GCCCGCAGCA	
wssIID				GCCtGCAGCc	
wssiia	AGAAGCTCCA	AACCCAAAGG	CTCTTTCGCC	GCCTGCAGCC	CCCGCTGTAC
	057				1000
··CCTTD	951 AACAAGACCT	MMCCCO N CMMC	**************	mmaammaa x	1000
wssIIB wssIID	AAGAAGACCT				
WSSIIA	AAGAAGACCT				
WODIIA	AAGAAGACCI	TIGGGACTIC	AAGAAAIACA	TIGGETTEGA	ddAdcccoid
	1001				1050
wssIIB	GAGGCCAAGG	ATGATGGCCG	GGCTGTTGCA	GATGATGCGG	
wssIID	GAGGCCAAGG				
wssiia	GAGGCCAAGG				
	1051				1100
wssiib	ACACCACCAG	AATCACGATT	CCGGGCCTTT	GGCAGGGGAG	AACGTCATGA
wssiid	ACACCACCAG				
wssiia	ACATCACCAG	AACCATGATT	CCGGACCTTT	GGCAGGGGAG	AACGTCATGA

	1101				1150
wSSIIB	ACGTGGTCGT	CGTGGCTGCT	GAATGTTCTC	CCTGGTGCAA	AACAGGTGGT
wSSIID	ACGTGGTCGT	CGTGGCTGCT	GAGTGTTCTC	CCTGGTGCAA	AACAGGTGGT
wssiia	ACGTGGTCGT	CGTGGCTGCT	GAATGTTCTC	CCTGGTGCAA	AACAGGTGGT
	1151				1200
wssIIB	CTTGGAGATG	TTGCCGGTGC	TTTGCCCAAG	GCTTTGGCGA	AGAGAGGACA
wssIID	CTqGGAGATG			GCTTTGGCaA	
wssiia	CTTGGAGATG	_		GCTTTGGCGA	
	011001101110	1100000100	11100001110	001110000.	,
	1201				1250
wssiib		СТТСТССТАС	СУУССТУТСС	GGACTATGAG	
WSSIID				GGACTATGAa	
wssiiA				GGACTATGAG	-
WSSIIA	ICGIGITATG	GIIGIGGIAC	CAAGGTATGG	GGACTATGAG	GAAGCCIACG
	1251				1300
		GGG333383G	mr	amaan an aan	
wssiib	ATGTCGGAGT	<del>-</del> -		CTGGACAGGA	
wssiid		CCGAAAATAC		CTGGACAGGA	TATGGAAGTG
wssiia	ATGTCGGAGT	CCGAAAATAC	TACAAGGCTG	CTGGACAGGA	TATGGAAGTG
	1301				1350
wssiib		ATGCTTATAT			TCATTGACGC
wssiid		ATGCTTaTAT	CGATGGAGTT	GATTTTGTGT	TCATTGACGC
wSSIIA	AATTATTTCC	ATGCTTATAT	CGATGGAGTT	GATTTTGTGT	TCATTGACGC
	1351				1400
wSSIIB	TCCTCTCTTC	CGACACCGCC	AGGAAGACAT	TTATGGGGGC	AGCAGACAGG
wssIID	TCCTCTCTTC	CGACACCGAG	AGGAAGACAT	TTATGGGGGC	AGCAGACAGG
wssiia	TCCTCTCTTC	CGACACCGCC	AGGAAGACAT	TTATGGGGGC	AGCAGACAGG
	1401				1450
wssiib	AAATTATGAA	GCGCATGATT	TTGTTCTGCA	AGGCCGCTGT	CGAGGTTCCA
wssIID	AAATTATGAA	GCGCATGATT	TTGTTCTGCA	AGGCCGCTGT	TGAGGTTCCA
wssiia	AAATTATGAA	GCGCATGATT	TTGTTCTGCA	AGGCCGCTGT	CGAGGTTCCT
	1451				1500
wssiib	TGGCACGTTC	CATGCGGCGG	TGTCCCTTAT	GGGGATGGAA	ATCTGGTGTT
wssiid	TGGCACGTTC	CATGCGGCGG	TGTCCCTTAT	GGGGATGGAA	ATCTGGTGTT
wSSIIA	TGGCACGTTC	CATGCGGCGG	TGTCCCTTAT	GGGGATGGAA	ATCTGGTGTT
	1501				1550
wSSIIB	TATTGCAAAT	GATTGGCACA	CGGCACTCCT	GCCTGTCTAT	CTGAAAGCAT
wSSIID	TATTGCAAAT	GATTGGCACA	CGGCACTCCT	GCCTGTCTAT	CTGAAAGCAT
wSSIIA	TATTGCAAAT	GATTGGCACA	CGGCACTCCT	GCCTGTCTAT	CTGAAAGCAT
	1551				1600
WSSIIB	ATTACAGGGA	CCATGGTTTG	ATGCAGTACA	CTCGGTCCAT	TATGGTGATA
wssIID	ATTACAGGGA	CCATGGTTTG	ATGCAGTACA	CTCGGTCCAT	TATGGTGATA
wSSIIA	ATTACAGGGA		- · · ·		
				<del></del>	
	1601				1650
wSSIIB		CTCACCAGGG	CCGTGGCCCA	GTAGATGAGT	
wSSIID				GTAGATGAAT	
WSSIIA	CATAACATCG				
II	CALINGTICS	COCHOOG	CCCIGGCCCA	CARONIGHMI	- CCCOIICAC

wssiib wssiid wssiia	CGAGTTGCCT	GAGCACTACC GAGCACTACC GAGCACTACC		CAGACTGTAC CAGACTGTAC CAGACTGTAC	GACCCCGTGG
wssiib wssiid wssiia	GTGGTGAACA	CGCCAACTAC	TTCGCCGCCG TTCGCCGCCG	GCCTGAAGAT	GGCGGACCAG
wssiib wssiid wssiia	GTTGTCGTGG	TGAGCCCCGG	GTACCTGTGG GTACCTGTGG GTACCTGTGG	GAGCTGAAGA	CGGTGGAGGG
wssiib wssiid wssiia	CGGCTGGGGG	CTTCACGACA	TCATACGGCA TCATACGGCA TCATACGGCA	GAACGACTGG	AAGACCCGCG
wssiib wssiid wssiia	GCATCGTCAA	CGGCATCGAC	AACATGGAGT AACATGGAGT AACATGGAGT	GGAACCCCGA	GGTGGACGCC
wssiib wssiid wssiia	CACCTCAAGT	CGGACGGCTA	CACCAACTTC CACCAACTTC CACCAACTTC	TCCCTGAGGA	CGCTGGACTC
wssiib wssiid wssiia	CGGCAAGCGG	CAGTGCAAGG	AGGCCCTGCA AGGCCCTGCA AGGCCCTGCA	GCGCGAGCTG	GGCCTGCAGG
wssiib wssiid wssiia	TCCGCGCCGA	CGTGCCGCTG	CTCGGCTTCA CTCGGCTTCA CTCGGCTTCA	TCGGCCGCCT	GGACGGGCAG
wssiib wssiid wssiia	AAGGGCGTGG	AGATCATCGC	GGACGCGATG GGACGCCATG GGACGCCATG	CCCTGGATCG	TGAGCCAGGA
wssiib wssiid wssiia	CGTGCAGCTG	GTGATGCTGG	GCACCGGGCG GCACCGGCCG GCACCGGCCG	CCACGACCTG	GAGAGCATGC
wssiib wssiid wssiia	TGCAGCACTT	CGAGCGGGAG	CACCACGACA CACCACGACA CACCACGACA	AGGTGCGCGG	GTGGGTGGGG

	2201				2250
wssilB	TTCTCCGTG	GGCTGGCGC#	CCGGATCAC	GCCGGCGCCG	ACGCGCTCCT
wssIID	TTCTCCGTGC				ACGCGCTCCT
wssiia	TTCTCCGTqc				ACGCGCTCcT
	1101000190	CCCTGGCGCF	CCGOATCACC	GCGGGCGCCG	ACCCCCTCCT
	2251				2300
wssiib	CATGCCCTCC	· CGGTTCGAGC	י רפיזפרפפאריז	GAACCAGCTC	
wssiid		CGGTTCGTGC			
wssiia	CATGCCCTCC	CGGTTCGAGC	: CGTGCGGGT1	GAACCAGCTt	TACGCCATGG
	2301				2350
wssIIB		COTOCOCOTO		TCGGTGGCCT	
wssIID		CGTCCCCGTC			
wssiia	CCTACGGCAC	CGTCCCCGTC	GTGCACGCCG	TCGGCGGGGT	GAGGGACACC
	2351				2400
wSSIIB	GTGCCGCCGT	TCGACCCCTT	СААССАСТСС	GGGCTCGGGT	
wssIID	GTGCCGCCGT			GGGCTCGGGT	GGACGTTCGA
wssiia					
WSSIIA	GTGCCGCCGT	TCGACCCCTT	CAACCACTCC	GGcCTCGGGT	GGACGTTCGA
	2401				2450
wSSIIB	CCGCGCAGAG	GCGCAGAAGC	TGATCGAGGC	GCTCGGGCAC	TGCCTCCGCA
wssiid		GCGCACAAGC		GCTCGGGCAC	
WSSIIA		GCGCACAAGC			
WDDIIM	CCGCGCCGAG	GCGCACAAGC	TGATCGAGGC	GCTCGGGCAC	TGCCTCCGCA
	2451				2500
wssIIB	CCTACCGGGA	СТАСААССАС	AGCTGGAGGG	GGCTCCAGGA	
wssIID				CCCTCCAGGA	
wssiia	CCTACCGGGA	CTACAAGGAG	AGCTGGAGGG	GcCTCCAGGA	GCGCGGCATG
	2501				2550
wssIIB	TCGCAGGACT	TCAGCTGGGA	GCATGCCGCC	AAGCTCTACG	
wssiid	TCGCAGGACT			AAGCTCTACG	
WSSIIA					
MOSITA	ICGCAGGACI	1 CAGC 1 GGGA	GCATGCCGCC	AAGCTCTACG	AGGACGTCCT
	2551				2600
wSSIIB	CGTCAAGGCC	AAGTACCAGT	GGTGAACGCT	AGCTGCTAGC	CGGTCCAGCC
wssIID				AGCTGCTAGC	
wssiia				AGCTGCTAGC	
WODIIA	CCTCAAGGCC	AAGIACCAGI	GGIGAACGCI	AGCIGCIAGC	CGCTCCAGCC
	2601				2650
wSSIIB	CCGCATGCG.	TGCATGA	CAGGATGGAA	TTGCGCATTG	CGCACGCAGG
wssiid				CTGCATTG	
wssiia				CTGCGCATTG	
"ODIIA	CCGCATGCGT	GCATGCatgA	gAddg IddAA	CIGCGCAIIG	CGCCCGCAGG
	2651				2700
wssiib	AAGGTGCCAT		. GGAGCGCCG	GCATCCGCGA	AGTACAGTGA
wSSIID				GCATCCGCGA	
wssiia				GCATCCGCGA	
					331301010n
	2701				2750
wSSIIB	CATGAGGT	GTGTGTGGTT	GAGACGCTGA	TTCC	GATCTGGTCC
wSSIID				TTCC	
wSSIIA				TTCCGATCTc	
					5

	2751				2800
wssiib	GTAGCAGAGT	AGAGCGGAGG	TAGGGAAGCG	CTCCTTGTTA	CAGGTATATG
wssiid	GTAGCAGAGT	AGAGCGGAGG	TATATGGGAA	TCTTAACTTG	GTATTGTAAT
AII22w	GTAGCAGAGT	AGAGCGGAcG	TAGGGAAGCG	CTCCTTGTTg	CAGGTATATG
	2801				2850
wssiib	GGAATGTTGT	TAACTTGGTA	TTGTAATTTG	TTATGTTGTG	TGCATTATTA
wssiid	TTGTTATGTT	GTGTGCATTA	TTACAATGTT	GTTACTTATT	CTTGTTAAGT
wssiia	GGAATGTTGT	CAACTTGGTA	TTGTAgTTTG	cTATGTTGTa	TGCgTTATTA
	2851				2900
wssiib	CAGAGGGCAA	CGATCTGCGC	CGGCGCACCG	GCCCAACTGT	TGGGCCGGTC
wssiid	CGGAGGCCAA	GGGCGAAAGC	TAGCTCACAT	GTCTGATGGA	TGCAAAAAAA
WSSIIA	caatgttgtt	acttattctt	gtTAAAAAAA	AAAAAAAAA	AAAA~~~~~
	2901				2950
wssiib	GCACAGCAGC	CGTTGGATCC	GACCGCCTGG	GCCGTTGGAT	CCCACCGAAA
wssiid	AAAAAAAAA			~~~~~~~	~~~~~~
wssiia	~~~~~~~	~~~~~~	~~~~~~~	~~~~~~~	~~~~~~
	2951	2965			
wSSIIB	ААААААААА	AAAAA			
wssiid	~~~~~~~	~~~~			
wssiia	~~~~~~~	~~~~			

WSSIIA	1	REAVASAAS	FLALASA	A SP-GRSRRR	A RVSAPPPHAG	AGRLHV	PPWPP-QRTA	51
WSSIIB	1	******	*****	+ ** <u>-</u> ****	T ****\$***T*	*****	**S**_***	
WSSIID		~		·	_ ~~ <b>~~~</b> ~~			
ZSSIIA	,1.	****AV*\$\$*	STF*****	* **G**~-**	* **GSS*F*T*	*-S*SFAFWA	**S**RAPRD	57
ZSSIJB							ARRG*P*DG*	
PEASSII							7R	
POTSSII							:Q	
							· ·	
		N -	Transit pep	tide cleav	age site			
WSSIIA.	52					PPDPWTTDP	DAAEGGAPAP	
WSSIIB	52	*********	*****CT-**	**D******	. ********	*******	********	111
WSSIID	-							110
ZSSIIA	58							97
ZSSIIB								
PFASSII							ERKKLVSSID	
POTSSII							ERRKVVSSIK	
1010511	20	MAKING CAR I	" DIAGGERYA- S	-DESKUALQ	/ ITEMSKTA	1.IQQDEDQQTH	. EKKYAASSIK	110
WSSIIA	110		DOMICTORALC	ENTERNACION	መጀመድረኛ በአበአ	DA BUDGGOM	Uningmeet	
WSSIIR							VPVNGENKAN	
WSSIID	T T T						******	170
ZSSIIA	0.0							
ZSSIIA							ALADV*I*SI	
					-ESEEAAKSS			97
PEASSII							QKGSSSSGSA	
POTSSII	11/	SSL*NA	KGTYDGGSGS	LSDVDIPDVD	KDYNVTVPST	A*TGITDVDK	NTPPAISHDF	172
	~ 55							
WSSIIA							PRLDIDSDVE	
WSSIIB							*GS*TV****	
WSSIID							*GS*TV****	
ZSSIIA							GIAPPT**	
ZSSIIB							DASAVKPEPA	
PEASSII							ASSKLHFNEQ	
POTSSII	173	*E*KREIKRD	LADERAPPLS	RS*IT*SSQI	SSTVSSKR	TL*VPPETPK	SSQETLL**N	230
						wssIIp1 Reg		
WSSIIA							WAVADDAGSE	
WSSIIB							R*****	
WSSIID							R*****	
ZSSIIA							RVG******	
ZSSIIB							A*P*T**AAS	
PEASSII							PSSKEV+NEA	
POTSSII	231	SRKSLVD*PG 1	KKIQSYMPSL	R	<b>-</b> *ESSAS	HVEQRNENLE	GSS*EANEET	277
					Region 1		Region 2	
WSSIIA.					CKTGGLGDV A		GHRVMVVVP 3	
WSSIIB							****** 3	
WSSIID	292 *:	****** - = * **	******	*****	******	******	****** 3	49
ZSSIIA							***** 2	
ZSSIIB							****** 2	
PEASSII							***** 3	
POTSSII							******A* 3	

•				~	a newaido o			
				sgp-	1 Peptide 3	TOADI EDUDA	FULVGGGBUE	400
WSSIIA	350	RYGDYEEAYD	VGVRKYYKAA	GODMEVNYFH	AYIDGVDFVF	IDAPDERARQ	EDIYGGSRQE	409
WSSIIB	349	****	****	*****	*****	****	****	400
WSSIID	350	*******PT*	******	*****	*****	****	******	343
ZSSIIA	283	******V**F*	*+I*****	***[*****	******	***************************************	D******	700
ZSSIIB	249	****E*A**R*	L***RR**V*	***S**T***	S********	VEP	NN****E*LD	262
PRASSII	303	H**N*A**H*	I****R**V*	*****T***	THEFT	**S*I**NLE	SN****N*LD	202
POTSSII	338	**DN*P*PQ*	S****I**VD	***AD+J.**Ö	AFTWING	.HSUM IG	NN****N*VD	331
					Domina			
					Region		VODUČI MOVT	460
AII22W	410	IMKRMILFCK	AAVEVPWHVP	CGGVPYGDGN	LVFIANDWHT	ALLIPVILICAL	YRDHGLMQYT	403
WSSIIB	409	****	*****	*****	****	*******	******	460
WSSIID	410	******	****	****	****	******	*******	407
ZSSIIA	343	*****	A******	****C****	****	****	*******	3.60
ZSSIIB	309	*L******	******YA*	***TV****	****	****	***N*****A	422
PRASSII	363	*LR**V****	*****	***IC****	****	*****	*******N**	422
POTSSII	398	*I***A***	**I*****	****C****	****	****	***N*I*N**	457
						~~***************************	* IDIA DOUBER!	500
WSSIIA	470	RSIMVIHNIA	HQGRGPVDEF	PFTELPEHYL	EHFRLYDPVG	GEHANYFAAG	LKMADQVVVV	523
WSSIIB	469	*****	*****	***	****	********	****	528
WSSIID	470	****	******	****	******	*****	******	529
ZSSIIA	404	**VL*****	******	*YMD*****	Q**E*****	****	****R**T*	462
ZSSIIB	369	**^\T++**	*******D*	VNFD*****I	D**K***NI*	*D*S*V****	**T**R**T*	428
PRASSII	423	**VL*****	*******ED*	NTVD*\$GN**	DL*KM****	***E*I****	*+T**RI*T*	482
POTSSII	458	**VL*****	******LED*	SYVD**P**M	DP*K*****	***E*I****	**T**R**T*	517
				Re	egion 4			~ A A
WSSIIA	530	SPGYLWELKT	VEGGWGLHDI	IRONDWKTRG	IVNGIDNMEW	NPEVDVHLK-	SDGYTNFSLG	588
WSSIIB	529	*****	******	*****	****	****	******	587
WSSIID	530	*****	******	*****	******	****A***	*****	588
ZSSIIA	163	*D******	******	**S****IN*	*******HO**	**R*****R-	********	52I
ZSSIIB	429	*N**M****	S******	*N*****LQ*	*****MS**	**A*****H-	**D***YTFE	48/
PEASSII	483	*H**A****	S******N*	*NES***F* <u>*</u>	****V*TKD*	**OF*AY*T-	*********	541
POTSSII	518	*H**S****	50******Q*	*NE****LQ*	*****TK**	***L***PR	****M*Y**D	577
								-
				Region '	Γ,		Region	
WSSIIA	589	TLDSGKRQCK	EALQRELGLQ	VRADVPLLGE	IGRLDGOKGV	EIIADAMPWI	VSODVOLVML	648
WSSIIB	588	*****	****	**G*****	*****	****	******	54/
WSSIID	500	*****	****	*****	****	****	******	648
ZSSIIA	ちつつ	****	A******E	**D*****	****	Dangarara	AG"	JUL
ZSSIIB	488	******	A****O****	**D****T**	*****	D****TH**	AG	241
PEASSII	E42	*******	A*******	**E***IIS*	******	DL**E*I**M	Waltana	90T
POTSSII	578	**QT**P***	A***K****	**D****I**	******	DL**E*V**M	MG* <u>****</u>	637
						ion 6		
WSSIIA	649	GTGRHDLESM	LRHFEREHHD	KVRGWVGFSV	rlahrit <u>aga</u>	DALLMPSRFE	PCGLNOLYAM	708
WSSIIB	54B	******	******	*****	**** <del>*</del> **	*****	****	707
WSSIID	649	*****	*0******	*****	***** <u>**</u>	******	*****	/08
ZSSĪIA	582	*******	*O*L****PN	****	PM******	*V*V****	****	041
ZSSIIB	548	****D*	**R**S**S*	***A*****	P***** <u>**</u>	+ + + + + + + + + + + + + + + + + + + +	****	607
PRASSII	602	*******	*KE**AO*C*	*I*S*****	KM*****	* ******	****	007
POTSSII	638	****R***O*	**Q**CQ*N*	*I******	KTS******	*T*****	**A*****	697

WSSIIA WSSIIB WSSIID ZSSIIA ZSSIIB PEASSII	Region 7  709 AYGTVDVVHA VGGVPDTVFP FDPFNHSGLG WTFDPAEAHK INTEALGHCLR TYRDYKESWR 768  708 ********* ***L***** *****************
<b>PQ</b> 15311	Oyo KI
WSSIIA WSSIIB WSSIID ZSSIIA ZSSIIB PEASSII POTSSII	769 GLQERGMSQD FSWEHAAKLY EDVLLKAKYQ W 799 768 ******** ******* ******* * 798 769 ******** ****** ****** * 799 702 S**A***** L**D***E** ****V**** * 732 668 ACRA**AE* L**D***V** ****V**** * 698 722 *I******* L**DN**QQ* *E**VA*** * 752 759 *I*T*C*T** L**DN**QQ* *E**IA*** * 788

### FIGURE 3-3

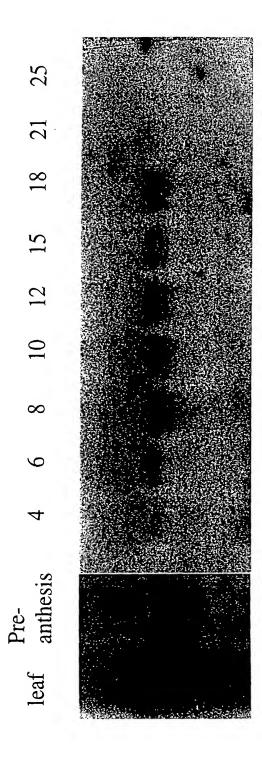


FIGURE 4

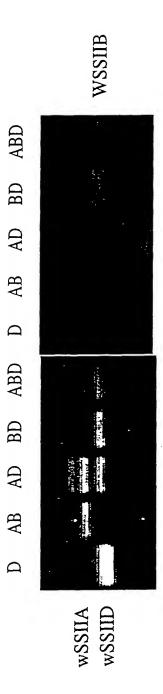


FIGURE 5

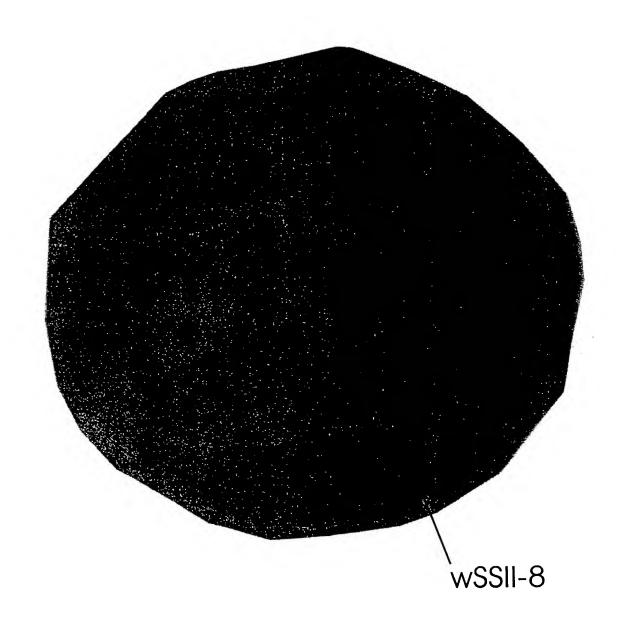
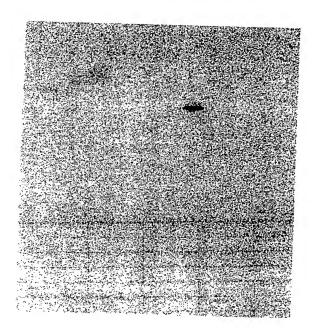


FIGURE 6

# 12345678910M

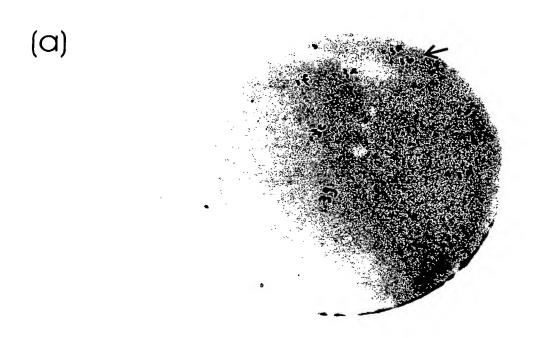


1				50	
maizeSSIII	MEMVLRSQSP	LCLRSGPVLI	FRPTVAGGGG	GTQSLLRTTR	FARRRVIRCV
potatoSSIII	~~~~~~~	~~~~~~~~	~~~~~~~	~~~~~~	~~~~~~~
wheatSSIII	MEMSLWPRSP	LCPRSRQPLV	V.VRPAGRGG	LTQPFLMNGR	FTRSRTLRCM
	51				100
maizeSSIII	VASPGCPNRK	S.RTASPNVK	VAAYSNYAPR	LLVESSSKKS	EHHDSSRHRE
potatoSSIII	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~~	~~~~~~~
wheatSSIII	VASSDPPNRK	SRRMVPPQVK	VISSRGYTTR	LIVEPSNENT	EHNNRDE
	101			,	150
maizeSSIII			RDVEIEVDLQ		
potatoSSIII			~~~~~~~		
wheatSSIII	ETLDTYNALL	STETAEWTDN	REAE	. TAKADSSQN	ALSSSIIGEV
	151				200
maizeSSIII			VLRNVAVREV		
potatoSSIII					
wheatSSIII	DVADEDILAA	DLTVISLSSV	MKKEVDAADK	ARVKED	AFELDXAS I I
	201				250
maizeSSIII		DEVERDMA	DVDILGLDLN	NATTEETDIM	_
potatoSSIII			~~~~~~~		
wheatSSIII			SVIVDVMDHN		
"IICACDOLLI	2101212112	IIID CQ D I DIC	5,2,5,,		
	251				300
maizeSSIII	DSPGNASSGR	TYGGVDELGE	LPSTSVDCIA	INGKRRSLKP	KPLPIVRFQE
potatoSSIII	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~
wheatSSIII	KARVEEDVFE	LDLSGNISSS	ATTVELDAVD	EVGPVQDKFE	ATSSGNVSNS
					•
	301				350
maizeSSIII	QEQIVLSIVD	EEGLIASSCE	. EGQPVVDYD	KQEENSTAFD	EQKQLTDDFP
potatoSSIII			~~~~~~~		
wheatSSIII	ATVREVDASD	EAGNDQGIFR	ADLSGNVFSS	STTVEVGAVD	EAGSIKDRFE
		•			
	351			~~	400
maizeSSIII		EPNNDIVGSS	KFLEQKQELD	GSYKQDRSTT	GLHEQDQSVV
potatoSSIII	mp.c.com.c.mc	A DAMADA TORR	VADODEEAD	T COMM CCCM	VDEIDDIAME
wheatSSIII	TDSSGNVSTS	APMWDAIDET	VADQDTFEAD	LSGNASSCAI	IREVDDVVDE
	401				450
maizeSSIII		CVPOOTOVND	QSIAGSHRQD	OSTAGAPROT	
potatoSSIII			~~~~~~		
wheatSSIII			ESGHEKHMAV		
***************************************					
	451				500
maizeSSIII	QSIVGSCKQH	ELIIPEPKKI	ESIISYNEID	QSIVGSH.KQ	DKSVVSVPEQ
potatoSSIII	RSLSCTSVSN	AITHLKIKPI	LGFVSHGTTS	LSVQSSSWRK	DGMVTGVSFS
wheatSSIII			LRLVRVEE		
	501				550
maizeSSIII			AESIIGVPEK		
potatoSSIII	ICANFSGRRR	RKVSTPRSQG	SSPKGFVPRK	PSGMSTQRKV	QKS . NGDKES

wheatSSIII		NQSIIGSYKQ	DKSIADVAGP	TQSIFGSSKQ	HRSIVAFPKQ
	553				
maizeSSIII	551	TOCTUMENT	NOSIVGLPKO	OOSTUUTUED	600
potatoSSIII		-	VETSDDDTKG		
wheatSSIII		· <del>-</del>	DLSAVSLPKQ		· · · =
			220111221112		
	601				650
maizeSSIII	.DLSIVGISN	EFQTKQLATV	GTHDGLLMKG	VEAKETSQKT	EGDTLQATFN
potatoSSIII			DDKDAVKLNK		.GFIIDSVIR
wheatSSIII	DRQDALYVNG		LEA	KEGDHTSEKT	DEDALHVKFN
	651				700
maizeSSIII		LTKEADETTT	IEKINDEDLV	MTEEOKSTAM	
potatoSSIII			GTKLYEILQV		
wheatSSIII			WKKVDEEHLY		
	701				750
maizeSSIII			LSEEESSWDE		-
potatoSSIII			ESNEIDDLDT		
wheatSSIII	ELSITEIG	MGRGDKIQHV	LSEEELSWSE	DEVQLIEDDG	QYEVDETSVS
	751				800
maizeSSIII	TEQDIQES	PNDDLDPQAL	WSMLQELAEK	NYSLGNKLFT	
potatoSSIII	ETGDSSLN	LRLEMEANLR	RQAIERLAEE	NLLQGIRLFC	FPEVVKPDED
wheatSSIII	VNVEQDIQGS	PQDVVDPQAL	KVMLQELAEK	${\tt NYSMRNKLFV}$	FPEVVKADSV
	801	*******	was mianun		850
maizeSSIII potatoSSIII			KGAFNGWKWR MGAFNEWRYR		
wheatSSIII			KGAFNGWKWR		
WIIGUGBBILL		THE STORY I	nom nomm	21 12 12 11 10 2	200 VIII DCICE
	851				900
maizeSSIII			ENNNNNDFVI		
potatoSSIII			DNNDGNDFSI		
wheatSSIII	YIPKEAYRLD	FVFFNGRTVY	ENNGNNDFCI	GIEGTMNEDL	FEDFLVKEKQ
	901				050
maizeSSIII		AERRROTDEO	RRMEEERAAD	KADRVOAKVE	950 VETKKNKI.CN
potatoSSIII			RRIEAEKAEI	-	
wheatSSIII			RRRKEARAAD		
					•
	951				1000
maizeSSIII	VLGLARAPVD		_		
potatoSSIII	LMVKATKTRD				
wheatSSIII	MLSLARTCVD	NLWYIEASTD	TRGDTIRLYY	NRNSRPLAHS	TEIWMHGGYN
	1001				1050
maizeSSIII	NWIDGLSFAE	RLVHHHDKDC	DWWFADVVVP	ERTYVLDWVF	
potatoSSIII	NWKDGLSIVK	KLVKSERIDG	DWWYTEVVIP	DQALFLDWVF	ADGPPKHAIA
wheatSSIII	NWXDGLSIVE	SFVKCNDKDG	DWWYADVIPP	EKALVLDWVF	ADGPAGNARN
m=i=c00TTT	1051	7 MT D 1-22-42		TIMBL CARE	1100
maizeSSIII potatoSSIII	YDNNGGHDFH YDNNHRQDFH				
wheatSSIII	YDNNARQDFH				
	-31		LOI WAQLIQIA	TIMBUNK	MIMMITTELL
	1101				1150

1101

maizeSSIII potatoSSIII wheatSSIII	EKTALLKTET	KEKTMRMFLV KERTMKSFLL KAKTMRRFLL	SQKHVVYTE.	PLEIHAGTTI PLDIQAGSSV XLKYVPGTTV	TVYYNPANTV
maizeSSIII potatoSSIII wheatSSIII	LNGKPEIWFR	CSFNRWTHRL	GVLPPQKMVQ GPLPPQKMSP GALPPQKMVK	AENGTHVRAT	VKVPLDAYMM
maizeSSIII potatoSSIII wheatSSIII	DFVFSEREDG	GIFDNKSGMD	YHIPVFGSIA YHIPVFGGVA YHIPVSDSIE	KEPPMHIVHI	AVEMAPIAKV
maizeSSIII potatoSSIII wheatSSIII	GGLGDVVTSL	${\tt SRAVQDLNHN}$	VEVILPKYGC VDIILPKYDC VEVILPKYDC	LKMNNVKDFR	FHKNYFWGGT
maizeSSIII potatoSSIII wheatSSIII	EIKVWFGKVE	${\tt GLSVYFLEPQ}$	NGMFGVGYVY NGLFSKGCVY NGMFGVGCVY	GCSNDGERFG	
maizeSSIII potatoSSIII wheatSSIII	LQGGFSPDII	${\tt HCHDWSSAPV}$	AWLHKENYAK AWLFKEQYTH AWLYKEHYSQ	YGLSKSRIVF	1400 TIHNLEFGAH TIHNLEFGAD TIHNLEFGAH
maizeSSIII potatoSSIII wheatSSIII	1401 HIGKAMRYCD LIGRAMTNAD YIGKAMTYCD	KATTVSPTYS	~	PHLGKFYGIL PHLHKFHGIV PHREKFYGIL	NGIDPDIWDP
maizeSSIII potatoSSIII wheatSSIII	LNDKFIPIPY	TSENVVEGKT	AAKRALQQKF AAKEALQRKL AAKRALQQKF	GLKQADLPLV	GIITRLTHQK
maizeSSIII potatoSSIII wheatSSIII	GIHLIKHAIW	RTLERNGQVV	LLGSAPDSRI LLGSAPDPRV LLGSAPDHRI	QNNFVNLANQ	LHSKYNDRAR
maizeSSIII potatoSSIII wheatSSIII	LCLTYDEPLS	HLIYAGADFI	LVPSIFEPCG LVPSIFEPCG IVPSIFEPCG	LTQLTAMRYG	SIPVVRKTGG
maizeSSIII potatoSSIII wheatSSIII	LYDTVFDVDH	DKERAQQCGL	EPNGFSFDGA EPNGFSFDGA EPNGFSFDGA	DAGGVDYALN	RALSAWYDGR
maizeSSIII potatoSSIII wheatSSIII	DWFNSLCKQV	MEQDWSWNRP	ALDYIELYRS ALDYLELYHA ALDYIELYHA	ARKLE*	



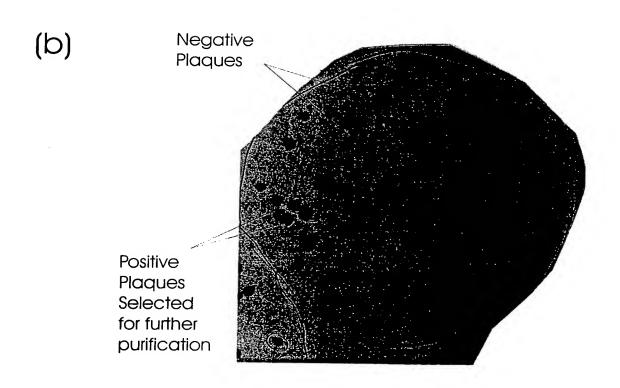
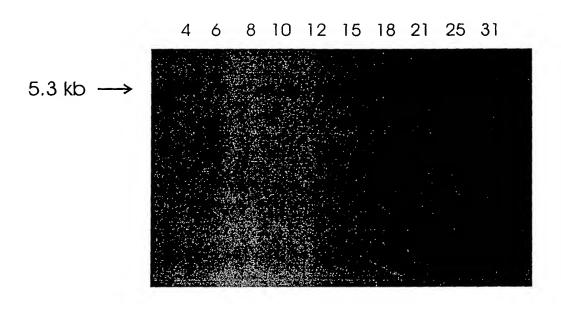


FIGURE 9

# (a) Wyuna



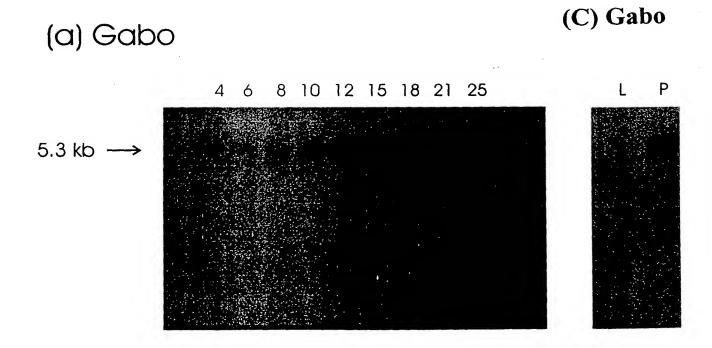
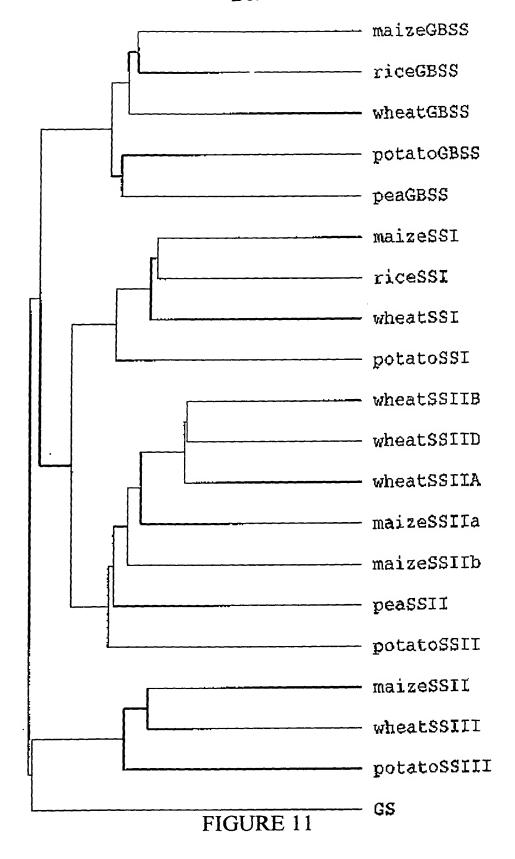


FIGURE 10



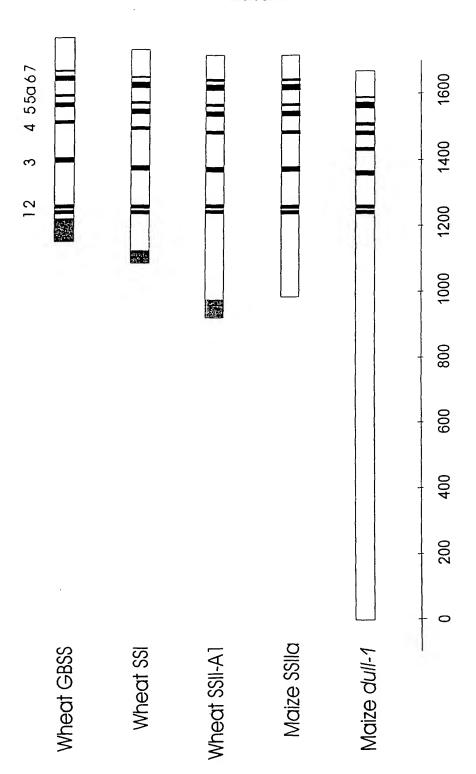


FIGURE 12

# FIGURE 13-

170 233 403 1276	260 323 493 1366	350 413 583 1456	440 503 673 1546	on 7 530 593 763 1636
90 DRVFVDHPCF *W*****SY *F**I*A*L* G**BDLTVY*	180 NGIYRAAKVA Y*V**DSRST H*LMQYTRSI -SRMASTR*V	270 GCELDNIMRL *LNELLSS*K *LHDIIRQND AGHGAIAPHR	360 DVMIASIPEI *LIKMA***- EIIADAM*W*	Region 450 TPCACASIGG *VPVVHG*** *VPVVHG*** SIPIVRK***
80 RYFHCYKRGV TF**E*RDN* N***A*ID**	Region 3  100  100  100  VCWDWHTGLL ACYLKSNYOS NGIYRAAKVA  *V****AS*V PVL*AAK*RP Y*V**DSRST  IA****AA** PV***AY*RD H*LMQYTRSI H*H**SSARV *HLX*EH*SQ -SRMASTR*V	260 ABBLISGEAR W*VTTAEGGQ W*LKTVBGGW TVSPTYSRDV	Region 5 340 VDRKVPLVAF IGRLEBOKGP *RED***IG* ****DX***I *PAD***IG* ****DG****VD**I*GI *T**TA***I	Region 6 420 440 450 HOMMAGADVL AVISREBECG LIOLOGMRYG TPCACASIGG *RIT***C*I* IMP******* *N***YA*O** *VPVVHG*** *RIT****A* INP******* *N**YA*A** *VPVVHAV** *LIY**S*FI I*P*I**** *T**VA*** SIPIVRK***
70 IKVDXYERV *PCFGGSHE* Y*!AGQDWE* -DGHLYQSFS		250 DKYLTVSPYY RI*TVSQG*S QV*VVSPG*L		430 AVISREBECG IMP******** INP***************************
60 SWNSE KALYTGKHIK G*RKY	150 SGPYGED:VP QNCM* D*NL**	240 NWMKAGILQA FLKG*VVTAD YFAAGLKMAD	330 EALQABVGLP AB**K*L** ****R*L**Q FG**QT	
50 DQYKDAWDT- LNGSSDKNYA GD*EE*Y*V- *CLNQSSVK-	140 ILNLDNNPYP **E*GGYI*G HVPCGGV**G	230 YDXPVEGRKI HALDKGEAVN **PVGGEHAN	320 EALEGKALNK DD*S***KC* TLDS**RQC* **AKRALQQ*	410 APLA V*VS VR**
Region 2 40 HRWNVISPRY *****YW*** *****YV***	130 LCQAALEVPR **Y**C*A*L F*K**V***W F*HS***P	220 PKSSFDFIDG BWVFPEMARR -BHYLEHPRL	310 TALEGKALNK FSLG NVVEG*	400 KSIBEKFP SKVRAVVRFN R*T*SSYK D*F*GW*G*S RHF*REHH D***GW*G*S FCRLADAL HG*YHGRVKL
Region 1 30 40 40 20 20 30 40 40 40 40 40 40 40 40 40 40 40 40 40	110 120 KIYGEDAGTD YEDNQQRFSL NFGA FG***F*YT* S RQBIMK*MI*	210 NLPDR G**PENYGAL E**	300 KRLAVNYDIT *C*PHH*SV- VH*KSDGYIN N*IP*P*TCE	390 LLKSIBEKFP WMR*T*SSYK M*RHF*REHH GDFCRLADAL
Region 1 20 KTGGLGDLLG *S*****VC* *******VA*	XIYGEDAGTD YEDNQQRFSL NFGA FG***F*YT* S RQBIMK*MI*	200 210 RFSFDDFAQL NLPDR LEPASTYPD* G**PEWYGAL *GPV*E*PFT E** AHYIGKAMTY CDK	Region 4 280 TGITIIVNGM DVSENDPTKD SVLNG****I *IND*N**T* WKTRG****I *NM**N*EV* EKFYG*L**I *PDI***YI*	Region 5a 370 IKEEDVOIVL LGTGKKKFER LIKSIBEKFP *MR****F*M **S*DPI**G WWR*T*SSYK V-SO***L*M ****RHDL*S M*RHF*REHH TL*SNG*V** **SAPDHRIQ GDFCRLADAL
10 FVGAEMAPWS -*TG*A**YA A**CS**C	100 LEKVRGKTKE -HRPGSLYGD RHRQEDIYGG **PQN*MFGV	190 FCIHNISYQG LV***LAH** MV****AH**	- · · · · · · · · · · · · · · · · · · ·	
81 144 314 1187	171 234 404 1277	261 324 494 1367	351 414 584 1457	
wgbss wss1 wss2 wss3	MGBSS WSS1 WSS2	wGBSS wSS1 wSS2 wSS3	#GBSS #SS1 WSS2 WSS3	wgbss wss1 wss2 wss3

	620	683	853	1726			710	773	943	1816
540	SEPGIVGBEI	<b>BOYEQIF*WA</b>	KLYED*LLKA	AARKF*					:	
530	DVLLELGVEG	PFGAKGEE GTGWAFSPLT VDKMLW*LRT AMSTFREHKP **E*LM*RGM TKDHTWDHAA BQYEQIF*WA	SQDPSWEHAA	PA*DYIELYH	630	000				
520	SWKGPAKNWE	**E*LM*RGM	**R*LQERGM	VMBQDWSWNR	909	0.50	•		•	
210	MVKNCMIQDL	AMSTFREHKP	CLRTYRDYKE	RDWFHSLCKR		1	:			:
200	KVVGTPAYHE	VDKMLW*LRT	AHKLIB*LGH	nraiganfda	909					
490	KVVTTLKRAV	GTGWAFSPLT	W*FD**E	ADSNGVDY*L	797	2		:		:
480	CNVVBPADVK	PFGAKGEE	PFNHSGLG	LEPNGFSFDG	787	2	:	:	:	
oncinuea)	WGBSS 531 LYDTIVEGKT GFHMGRLSYD CNVVRPADVK KVVTTLKRAV KVVGTPAYHB MVKNCMIQDL SWKGPAKNWE DVLLELGVEG SEPGIVGBBI	1 1 2 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	WSS2 764 VR**-*PPFDPFNHSGLGW*FD**E AHKLIB*LGH CLRTYRDYKE **R*LQERGM SQDPSWEHAA KLYED*LLKA	"SS3 1637 ****-*FDVD NDKDRAR*LG LEPNGFSFDG ADSNGVDY*L NRAIGARFDA RDWFHSLCKR VMBQDWSWNR PA*DYIELYH AARKF*	220	2	WGBSS 621 APLAMENVAA P*	WSS1 684 FVDQPYVM	wSS2 854 KYQW	wss3 1727
Region / (continued)	LYDTIVEGAT	WSS1 594 *R*****TFN	VR**-*PPFD	****-*FDVD	0 9 3	000	APLAMENVAA	FVDQPYVM	KYQW	:
	531	594	764	1637			621	684	854	1727
	wGBSS	wSS1	wS52	wSS3	e d	ממכ	WGBSS	4.551	wS52	w\$83

# FIGURE 13-2

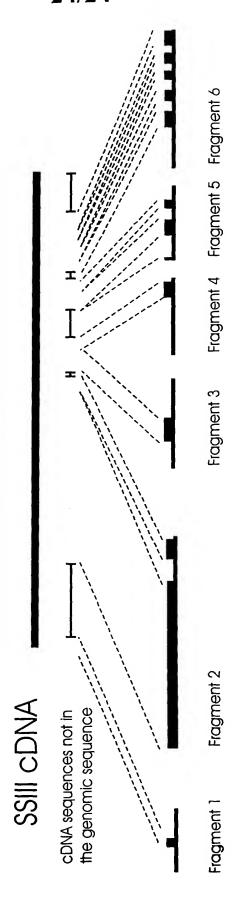


FIGURE 14

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